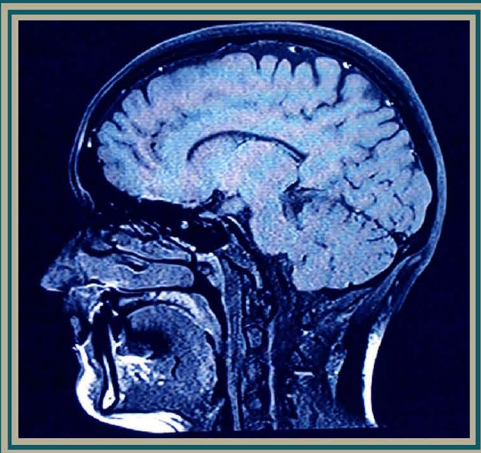


EIGHTH EDITION

*Scott-Brown's Otorhinolaryngology  
Head & Neck Surgery*



VOLUME 1

**Basic Sciences  
Endocrine Surgery  
Rhinology**

EDITED BY

**John C Watkinson  
Raymond W Clarke**

SECTION EDITORS

Louise Jayne Clark  
Adam J Donne  
R James A England

Hisham M Mehanna  
Gerald William McGarry  
Sean Carrie



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EIGHTH EDITION

**Otorhinolaryngology**  
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**Surgery**

**VOLUME 1**

Basic Sciences, Head and Neck Endocrine Surgery,  
Rhinology

**VOLUME 2**

Paediatrics, The Ear, Skull Base

**VOLUME 3**

Head and Neck Surgery, Plastic Surgery

# Scott-Brown's

# Otorhinolaryngology

# Head and Neck

# Surgery

EIGHTH EDITION

## VOLUME 1

### Editors

#### **John C Watkinson MSc (Nuclear Medicine; London) MS (London) FRCS (General Surgery) FRCS (ENT) DLO**

One-Time Honorary Senior Lecturer and Consultant ENT/Head and Neck and Thyroid Surgeon, Queen Elizabeth Hospital University of Birmingham NHS Trust and latterly the Royal Marsden and Brompton Hospitals, London, UK  
Currently Consultant Head and Neck and Thyroid Surgeon, University Hospital, Coventry and Warwick NHS Trust; and Honorary Consultant ENT/Head and Neck and Thyroid Surgeon, Great Ormond Street Hospital (GOSH)  
Honorary Senior Anatomy Demonstrator, University College London (UCL)  
Business Director, Endocrine MDT, The BUPA Cromwell Hospital, London, UK.

#### **Raymond W Clarke BA BSc DCH FRCS FRCS(ORL)**

Consultant Paediatric Otolaryngologist, Royal Liverpool Children's Hospital, Liverpool, UK  
Senior Lecturer and Associate Dean, University of Liverpool, UK.

### Section Editors

#### **Louise Jayne Clark MBChB MD(DIST) FRCS (ORL) FRCS (ENT)**

Queen Elizabeth University Hospital, Glasgow, UK.

#### **Adam J Donne PhD FRCS (ORL-HNS)**

Consultant Paediatric ENT Surgeon, Alder Hey Children's NHS Foundation Trust, Liverpool,  
Honorary Senior Lecturer, Liverpool University, UK.

#### **R James A England FRCS (ORL-HNS)**

Consultant Otolaryngologist Head and Neck and Thyroid Surgeon, Honorary Senior Lecturer,  
Hull and East Yorkshire NHS Hospitals Trust and Hull University, UK.

#### **Hisham M Mehanna PhD BMedSc (HONS) MBChB (hons) FRCS FRCS (ORL-HNS)**

Chair of Head and Neck Surgery and Director of the Institute of Head and Neck Studies and Education,  
School of Cancer Sciences, University of Birmingham, UK.

#### **Gerald William McGarry MD MBChB FRCS (ORL-HNS) FFSTed**

Consultant Surgeon, Head and Neck and Anterior Skull Base Surgery, Glasgow Royal Infirmary  
Greater Glasgow and Clyde NHS, Glasgow, UK  
Honorary Clinical Senior Lecturer, University of Glasgow, UK.

#### **Sean Carrie FRCS**

Consultant Otolaryngologist, Skull Base Surgeon, Newcastle-upon-Tyne Hospitals NHS Foundation, UK  
Honorary Senior Clinical Lecturer, Newcastle University, UK.



CRC Press

Taylor & Francis Group  
Boca Raton London New York

CRC Press is an imprint of the  
Taylor & Francis Group, an **informa** business

CRC Press  
Taylor & Francis Group  
6000 Broken Sound Parkway NW, Suite 300  
Boca Raton, FL 33487-2742

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CRC Press is an imprint of Taylor & Francis Group, an Informa business

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Printed on acid-free paper

International Standard Book Number-13: 978-1-138-09461-1 (Hardback; Volume 1)  
International Standard Book Number-13: 978-1-138-09463-4 (Hardback; Volume 2)  
International Standard Book Number-13: 978-1-138-09464-2 (Hardback; Volume 3)  
International Standard Book Number-13: 978-1-4441-7589-9 (Hardback; Set)  
International Standard Book Number-13: 978-1-138-19652-0 (International Student Edition; restricted territorial availability)

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Library of Congress Cataloging-in-Publication Data

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Names: Watkinson, John C., editor. | Clarke, Ray (Raymond), editor.  
Title: Scott-Brown's otorhinolaryngology and head and neck surgery : basic sciences, endocrine surgery, rhinology / John Watkinson, Ray Clarke.  
Other titles: Scott-Brown's otorhinolaryngology, head and neck surgery | Otorhinolaryngology and head and neck surgery.  
Description: Eighth edition. | Boca Raton : CRC Press, [2018] | Preceded by Scott-Brown's otorhinolaryngology, head and neck surgery. 7th ed. c2008. | Includes bibliographical references and index.  
Identifiers: LCCN 2017032760 (print) | LCCN 2017033968 (ebook) | ISBN 9780203731031 (eBook General) | ISBN 9781351399067 (eBook PDF) | ISBN 9781351399050 (eBook ePub3) | ISBN 9781351399043 (eBook Mobipocket) | ISBN 9781138094611 (hardback : alk. paper).  
Subjects: | MESH: Otolaryngology--methods | Otorhinolaryngologic Diseases--surgery | Head--surgery | Neck--surgery | Otorhinolaryngologic Surgical Procedures--methods.  
Classification: LCC RF20 (ebook) | LCC RF20 (print) | NLM WV 100 | DDC 617.5/1--dc23  
LC record available at <https://lcn.loc.gov/2017032760>

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<http://www.crcpress.com>

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# Contributors

**Nithin D Adappa MD**

Assistant Professor  
Department of Otorhinolaryngology, Head and Neck  
Surgery  
University of Pennsylvania  
Philadelphia, USA.

**Shahzada K Ahmed BSC(HONS) DLO FRCS (ORL-HNS) PHD**

Consultant Rhinologist and Skull Base Surgeon  
Queen Elizabeth Hospital  
Birmingham, UK.

**Waseem Ahmed MBBS BSC FRCS (ORL-HNS)**

Clinical Fellow in Otology  
Gloucestershire NHS Hospitals Foundation Trust  
Gloucester, UK.

**Osama Al Hamarneh MBBS MD FRCS (ORL-HNS)**

Interface Fellow in Head and Neck Surgical Oncology  
Department of Otolaryngology Head and Neck Surgery  
Royal Hallamshire Hospital  
Sheffield, UK.

**Sarah Al-Himdani MBChB MRCS (ENG)**

Plastic Surgery Speciality Registrar  
Plastic and Reconstructive Surgery Research  
Manchester Institute of Biotechnology  
University Hospital of South Manchester NHS  
Foundation Trust  
Institute of Inflammation and Repair, Faculty of Medical  
and Human Sciences  
University of Manchester, Manchester Academic Health  
Science Centre, Manchester, UK.

**Samah Alimam MBChB MRCP**

Haematology Specialty Registrar  
Central Manchester Foundation Trust  
Manchester, UK.

**Ursula Altmeyer MRCP FRCPath**

Consultant Microbiologist  
University Hospital Crosshouse  
NHS Ayrshire & Arran  
Scotland, UK.

**Ghassan Alusi PHD FRCS (ORL-HNS)**

Consultant Otolaryngologist  
ENT Department  
Barts and the London NHS Trust  
London, UK.

**Shahram Anari MD MSc FRCS (ORL-HNS)**

Consultant Otorhinolaryngologist/Head &  
Neck Surgeon  
Department of Otolaryngology  
Heartlands Hospital  
Birmingham, UK.

**Mario Andrea MD PHD**

Professor of Otolaryngology  
Faculty of Medicine  
University of Lisbon  
Lisbon, Portugal.

**Steve L Atkin MBBS PHD**

Professor of Medicine  
Weill Cornell Medical College Qatar  
Qatar Foundation  
Education City  
Doha, Qatar.

**Patrick R Axon MD FRCS (ORL-HNS)**

Consultant Otologist and Skull Base Surgeon  
Department of Otolaryngology  
Cambridge University Hospitals  
Cambridge, UK.

**Mo Aye MB BS FRCP FRCPE**

Consultant Endocrinologist  
Centre for Metabolic Bone Disease  
Hull Royal Infirmary  
Hull, UK.

**Ashraf Ayoub PHD BDS MDS FDSRCS FDSRCP**

Professor of Oral and Maxillofacial Surgery  
Dental School, School of Medicine  
College of Medical, Veterinary and Life Sciences  
University of Glasgow  
Glasgow, UK.

**John Ayuk MD MRCP**

Consultant Endocrinologist  
Department of Endocrinology  
Queen Elizabeth Hospital, Birmingham; and  
Honorary Senior Lecturer  
University of Birmingham  
Birmingham, UK.

**Eileen Baidam MBChB MRCP DCH FRCP FRCPC**

Consultant Paediatric Rheumatologist  
Alder Hey Children's Foundation NHS Trust  
Alder Hey, Liverpool; and  
Honorary Senior Lecturer  
Liverpool University  
Liverpool, UK.

**Paul Baines MD PHD MRCP FRCA FFICM**

Consultant in Paediatric Intensive Care  
Alder Hey Children's NHS Foundation Trust  
Liverpool, UK.

**Stephen Ball MRCS PHD**

NIHR Academic Clinical Lecturer & Speciality Registrar  
in ENT Surgery  
Newcastle University & Newcastle Hospitals  
Newcastle, UK.

**Martyn L Barnes MBBS MD MSC(OXON) FRCS-ORL(EDIN)**  
Rhinology and Anterior Skullbase Fellow to Dr Richard Douglas, North Shore Hospital, Auckland; and Honorary Senior Clinical Teacher University of Dundee; and Director to SurgTech Ltd Dundee, UK.

**Ardeshir Bayat BSC(HONS) MBBS MRCS(ENG, EDIN) PHD**  
Clinical Scientist and Associate Professor Plastic and Reconstructive Surgery Research Manchester Institute of Biotechnology University Hospital of South Manchester NHS Foundation Trust Institute of Inflammation and Repair, Faculty of Medical and Human Sciences University of Manchester, Manchester Academic Health Science Centre, Manchester, UK.

**Rajiv K Bhalla BSC (HONS) FRCS (ORL-HNS) MD PGCERT MED ED**  
Consultant ENT Surgeon and Rhinologist Department of Otolaryngology, Head and Neck Surgery Manchester Royal Infirmary Manchester, UK.

**Brian JG Bingham MBCHB FRCS ED GLAS**  
Consultant ENT Surgeon Department of Otolaryngology, New Victoria Hospital & Southern General Hospital; and Honorary Senior Lecturer in Otorhinolaryngology University of Glasgow Glasgow, UK.

**Kristien Boelaert MD PHD FRCP**  
Reader in Endocrinology Centre for Endocrinology, Diabetes and Metabolism Institute of Metabolism and Systems Research College of Medical and Dental Sciences University of Birmingham; and Honorary Consultant Endocrinologist Department of Endocrinology University Hospital Birmingham NHS Foundation Trust Birmingham, UK.

**James D Brierley MBBS FRCP FRCR FRCSP(C)**  
Professor Department of Radiation Oncology University of Toronto Princess Margaret Cancer Centre Toronto, Canada.

**Steven M Bromley MD FANN**  
Director South Jersey MS Center and Bromley Neurology PC Audubon New Jersey, USA.

**James V Byrne MD FRCS FRCR**  
Professor of Neuroradiology University of Oxford; and Consultant Neuroradiologist John Radcliffe Hospital Oxford, UK.

**A Simon Carney BSC (HONS) MBCHB FRCS FRACS MD**  
Associate Professor and Head of ENT Unit Flinders University and Flinders Medical Centre Adelaide, South Australia.

**Sean Carrie MB CHB FRCS FRCS (ORL)**  
Consultant Rhinologist and Hon Senior Lecturer Newcastle upon Tyne Hospitals NHS Foundation Trust Newcastle University Newcastle, UK.

**Ricardo L Carrau MD**  
Professor, Director of the Comprehensive Skull Base Surgery Program Department of Otolaryngology, Head & Neck Surgery The Ohio State University Wexner Medical Center Columbus, Ohio, USA.

**David Chadwick BM BCH FRCS(ED) MD**  
Consultant Endocrine Surgeon Chesterfield Royal Hospital Chesterfield, UK.

**Philip G Chen MD**  
Assistant Professor & Program Director Rhinology & Otolaryngology Department of Otolaryngology, Head and Neck Surgery University of Texas Health, San Antonio Texas, USA.

**Andrew Coatesworth FRCS (ORL-HNS)**  
Consultant ENT Surgeon York Teaching Hospital NHS Foundation Trust York Hospital York, UK.

**Steve Colley**  
Consultant Radiologist Queen Elizabeth Hospital Birmingham, UK.

**Rogan Corbridge MBBS BSC FRCS FRCS (ORL)**  
Consultant ENT Surgeon Oxford Centre for Head and Neck Oncology John Radcliffe Hospital Oxford, UK.

**Christopher Coulson PHD FRCS (ORL-HNS)**  
Consultant Otolaryngologist NIHR Clinical Lecturer, Otolaryngology Head and Neck Surgery School of Cancer Sciences, University of Birmingham Queen Elizabeth Hospital Birmingham, UK.

**Valerie Cunningham MBCHB FRCA FFICM**  
Consultant Anaesthetist Institute of Neurosciences Queen Elizabeth University Hospital Glasgow, UK.

**Dustin M Dalgorf MD FRCS**

Clinical Lecturer  
Rhinology and Skull Base Surgery  
Mount Sinai Hospital and St Joseph's Health Centre  
University of Toronto  
Toronto, Canada.

**Leigh Delbridge MD FRACS**

Emeritus Professor of Surgery  
University of Sydney  
Sydney, Australia.

**Oscar Dias MD PHD**

Professor of Otolaryngology  
Faculty of Medicine  
University of Lisbon  
Lisbon, Portugal.

**Leo FS Ditzel Filho MD**

Neurosurgeon  
Department of Neurosurgery  
Wexner Medical Center at The Ohio State University  
Columbus, Ohio, USA.

**Robert I Docking MBCHB FRCA**

Specialty Registrar, Anaesthesia and  
Intensive Care Medicine  
Glasgow Royal Infirmary  
Glasgow, UK.

**Richard L Doty PHD**

Professor and Director  
Smell & Taste Center  
University of Pennsylvania Medical Center  
Philadelphia, USA.

**Stergios Doumas MD DDS**

Maxillofacial/Head & Neck Fellow  
Leeds Teaching Hospitals NHS Trust  
Leeds, UK.

**Elizabeth Drewe MBBS PHD MRCP MRCPATH**

Consultant Clinical Immunologist  
Nottingham University Hospitals NHS Trust  
Nottingham, UK.

**Charles East FRCS**

Consultant Surgeon  
University College London Hospitals NHS Trust; and  
Honorary Senior Lecturer, University College London  
London, UK.

**Ron Eccles BSC PHD DSC**

Professor Emeritus  
Common Cold Centre  
School of Biosciences  
Cardiff University  
Cardiff, UK.

**Stephen R Eil BSC (HONS) MBBS CERTAVMED MD  
FRCS (EDIN) FRCS (ORL-HNS)**

Consultant ENT Surgeon  
Hull and East Yorkshire Hospitals NHS Trust; and  
Honorary Professor, University of Hull  
Hull, UK.

**R James A England FRCS (ORL-HNS)**

Consultant Otolaryngologist Head and Neck and  
Thyroid Surgeon; and  
Honorary Senior Lecturer  
Department of Otolaryngology, Head and Neck Surgery  
Hull and East Yorkshire Hospitals NHS Trust  
Hull, UK.

**Phillip Evans BSC(HONS) MSC MED FHEA**

Senior University Teacher  
College of Medical, Veterinary and Life Science  
University of Glasgow  
Glasgow, UK.

**Sabena Fareedi MBBS BSC MRCP FRCP**

Consultant Neuroradiologist  
Queen Elizabeth Hospital  
Birmingham, UK.

**Erin A Felger MD FACS**

Associate Program Director of General Surgery  
Assistant Professor of Clinical Surgery Georgetown  
University  
Endocrine Surgeon  
Medstar Washington Hospital Center  
Washington DC, USA.

**Juan C Fernandez-Miranda MD**

Associate Professor of Neurological Surgery  
Director of the Complex Brain Surgery Program  
Associate Director of the Center for Cranial Base Surgery  
Department of Neurological Surgery  
University of Pittsburgh School of Medicine  
Pittsburgh, USA.

**Jonathan M Fishman MA (CANTAB) PHD DOHNS FRCS  
(ORL-HNS)**

Clinical Lecturer, Academy of Medical Sciences  
University College London  
London, UK.

**Tira Galm FRCS (ORL-HNS) MBA**

Rhinology Fellow  
Queen Elizabeth Hospital  
Birmingham, UK.

**Eng Cern Gan MBBS MRCS (EDIN) MMED (ORL) FAMS**

Consultant, Department of Otorhinolaryngology, Head  
& Neck Surgery  
Changi General Hospital; and  
Senior Clinical Lecturer  
Yong Loo Lin School of Medicine  
National University of Singapore, Singapore.

**Paul A Gardner MD**

Neurosurgeon  
Department of Neurological Surgery  
University of Pittsburgh School of Medicine  
Pittsburgh, USA.

**Quentin Gardiner MB CHB FRCS (ENG & ED) FRCS (ORL)**

Consultant Rhinologist  
Ninewells Hospital and Medical School  
Dundee, UK.

**Neil JL Gittoes PHD FRCP**

Consultant Endocrinologist  
Department of Endocrinology  
Queen Elizabeth Hospital Birmingham; and  
Honorary Senior Lecturer  
University of Birmingham  
Birmingham, UK.

**Michael Gleeson MD FRCS**

Professor of Otolaryngology and Skull Base Surgery  
Institute of Neurology, University College London; and  
Consultant  
Guy's, Kings and St Thomas' and the National Hospital  
for Neurology and Neurosurgery; and  
Honorary Consultant Skull Base Surgeon  
Great Ormond Street Hospital for Sick Children  
London, UK.

**Scott M Graham MD**

Professor of Otolaryngology, Head and Neck Surgery; and  
Professor, Department of Neurosurgery  
The University of Iowa; and  
Director of the Division of Rhinology  
University of Iowa Hospital and Clinics  
Iowa City, Iowa, USA.

**John Greenman PHD**

Professor of Tumour Immunology & Scientific Director,  
Daisy Building  
Head, School of Biological, Biomedical and  
Environmental Sciences  
University of Hull  
Hull, UK.

**Angela Hague BSC PHD**

Senior Teaching Fellow  
School of Oral and Dental Sciences  
University of Bristol  
Bristol, UK.

**Chris Hansell BSC PHD**

Post-Doctoral Research Associate  
Institute of Infection, Inflammation and Immunity  
University of Glasgow  
Glasgow, UK.

**Barney Harrison MB BS MS FRCS**

Consultant Endocrine Surgeon  
Royal Hallamshire Hospital  
Sheffield, UK.

**Richard J Harvey MD**

Program Head and Professor  
Rhinology and Skull Base  
Applied Medical Research Centre  
University of New South Wales  
Faculty of Medicine and Health Sciences  
Macquarie University  
Sydney, Australia.

**Maurice Hawthorne FRCS**

Consultant Otolaryngologist, Head and Neck Surgeon  
James Cook University Hospital  
Middlesbrough, UK.

**Mária Hérincs MD MBBS**

Clinical Fellow  
Department of Endocrinology  
William Harvey Research Institute  
Barts and the London School of Medicine  
Queen Mary University of London  
London, UK.

**John Hill MBBS FRCS FRCS(ENT) FRCSEd FRCSEd FRCSEd FRCSEd**

Consultant Otolaryngologist  
Freeman Hospital  
Newcastle, UK.

**Claire Hopkins FRCS (ORL-HNS) DM**

Professor of Rhinology, King's College, London  
Consultant ENT Surgeon  
Guy's and St Thomas' NHS Trust  
London, UK

**Simon Holmes BDS MD BS FDS RCS FRCS**

Consultant Oral and Maxillofacial Surgeon  
Barts and the London NHS Trust  
London, UK.

**Pamela Howson BMEDSCI MBBS FRACS**

Breast, Endocrine and General Surgeon  
Concord and Auburn Hospitals  
Sydney, Australia.

**Camille A Huser BSC MSC PHD**

Post-Doctoral Research Associate  
MRC University of Glasgow Centre for Virus Research  
Glasgow, UK.

**S Musheer Hussain MB BS MSC (MANC) FRCS (EDIN) FRCS (ENT) FFST FRCS (ORL)**

Consultant Otolaryngologist Head and Neck Surgeon  
Honorary Professor of Otolaryngology and Consultant  
ENT Surgeon  
Licenced Teacher of Anatomy  
Ninewells Hospital & University of Dundee Medical School  
Dundee, UK.

**Waraporn Imruetaicharoenchoke MD**

Consultant Surgeon  
Division of Head Neck and Breast Surgery  
Department of Surgery  
Faculty of Medicine Siriraj Hospital  
Mahidol University  
Bangkok, Thailand.

**Richard M Irving MD FRCS (ORL-HNS)**

Consultant in Neurotology  
University Hospital Birmingham NHS Trust and Diana  
Princess of Wales (Birmingham Children's) Hospital; and  
Honorary Senior Lecturer  
University of Birmingham  
Birmingham, UK.

**Laura Jackson BSC PHD MBCHC FRCS(ORL-HNS)**

Specialty Registrar ENT  
New Cross Hospital  
London, UK.

**Andrew James BSC MB MCH MD FRCP**

Consultant Endocrinologist  
Newcastle upon Tyne Hospitals NHS Foundation Trust  
Newcastle University  
Newcastle, UK.

**Amin R Javer BSC MD FRCSC FARS**

Clinical Professor of Surgery  
University of British Columbia; and  
Director, St Paul's Sinus Centre  
Vancouver, Canada.

**Christopher M Jones BMEDSC(HONS) MB CHB(HONS) MRCP**

Wellcome Trust Clinical Research Fellow  
Faculties of Biological Sciences, Medicine & Health  
University of Leeds  
Leeds, UK.

**Elizabeth Jones MBCHB BSC (PHARMACOL) MRCPCH  
FRCPATH**

Consultant Haematologist  
Wirral University Teaching Hospital Trust  
Wirral, UK.

**Mustaffa Junaid MBBS BSC (HONS) MRCS DOHNS**

Honorary Research Associate  
Institute of Head and Neck Studies and Education  
(InHANSE)  
School of Cancer Sciences  
University of Birmingham, UK.

**Dipti Kamani MD**

Harvard Medical School  
Research Director Thyroid and Parathyroid Surgical  
Division  
Massachusetts Eye and Ear Infirmary  
Boston, Massachusetts, USA.

**Russell David Keenan MB CHB FRCP FRCPATH**

Consultant Paediatric Haematologist  
Alder Hey Children's Hospital  
Liverpool, UK.

**Bruno MR Kenway BMEDSCI MBBS DOHNS FRCS (ORL-HNS)**

Otology Fellow  
Department of Otolaryngology  
Cambridge University Hospitals  
Cambridge, UK.

**Nitish Khanna MBCHB BSC DTMH FRCPATH**

Consultant Microbiologist  
Deputy Director  
Scottish MRSA Reference Laboratory & Scottish  
Salmonella, Shigella and C.difficile Reference  
Laboratory  
Glasgow Royal Infirmary  
Glasgow, UK.

**Dae Kim MBCHB BDS MSC FRCS PHD**

Consultant ENT Surgeon  
Queen Alexandra Hospital  
Portsmouth, UK.

**Márta Korbonits MD PHD FRCP**

Professor  
Department of Endocrinology  
William Harvey Research Institute  
Barts and the London School of Medicine  
Queen Mary University of London  
London, UK.

**Schelto Kruijff PHD MD**

Consultant Oncology and Endocrine Surgeon  
University Medical Center  
Groningen, Netherlands.

**Kevin Kulendra BSC MBBS DOHNS FRCS (ORL-HNS)**

Consultant ENT Surgeon  
ENT Department, Oxford University Hospitals NHS Trust  
John Radcliffe Hospital  
Oxford, UK.

**Michael Kuo PHD FRCS (ENG) FRCS (ORL-HNS) DCH**

Consultant Otolaryngologist, Head and Neck Surgeon  
Birmingham Children's Hospital  
Birmingham, UK.

**B Nirmal Kumar MPHIL FRCS (ORL-HNS) FACADMED**

Consultant Otolaryngologist Head & Neck Surgeon  
Director of Medical Education  
Wrightington, Wigan and Leigh NHS Foundation Trust  
Wigan, UK.

**Sonia Kumar FRCS (ORL-HNS)**

Fellow in Paediatric ENT Surgery  
Great Ormond St Hospital for Sick Children  
London, UK.

**Samuel C Leong MPHIL FRCSED (ORL-HNS)**

Rhinologist and Anterior Skull Base Surgeon  
Aintree University Hospital NHS Foundation Trust  
Liverpool, UK.

**Andy Levy BMEDSCI MBBS PHD FRCP**

Professor of Endocrinology and Honorary Consultant  
Physician  
University of Bristol and University Hospitals  
Bristol NHS Foundation Trust  
Henry Wellcome Labs  
Bristol, UK.

**Josh Lodhia MBCHB MRCS**

Specialist Trainee  
Department of Cardiothoracic Surgery  
Hull and East Yorkshire Hospitals NHS Trust  
Hull, UK.

**David A Lowe BSC FRCSED FRCS**

Research Fellow  
Clinical Effectiveness Unit  
Royal College of Surgeons of England  
London, UK.

**Darlene E Lubbe MBCHB FCORL(SA)**

Principal Specialist  
Division of Otorhinolaryngology  
University of Cape Town  
Cape Town, South Africa.

**Valerie J Lund CBE MS FRCS FRCSed**

Professor of Rhinology  
University College London; and  
Honorary Consultant ENT Surgeon  
Royal National Throat Nose & Ear Hospital  
London, UK.

**Andrew Mackay MBChB FRCA EDIC FFICM**

Consultant, Anaesthesia and Intensive Care Medicine  
Victoria Infirmary  
Glasgow, UK.

**Gerald W McGarry MB ChB MD FRCSed FRCS (ORL-HNS)  
FFSTED**

Consultant Surgeon  
Department of Otolaryngology and Head and  
Neck Surgery  
Glasgow Royal Infirmary  
NHS Greater Glasgow and Clyde  
Glasgow, UK.

**Julian A McGlashan MBBS FRCS(ORL)**

Special Lecturer and Consultant  
Department of Otorhinolaryngology  
Queen's Medical Centre Campus  
Nottingham University Hospitals  
Nottingham, UK.

**Colin MacIver FRCSS FRD FRCS ED FRCS (OMFS)**

Consultant Maxillofacial/Head & Neck Surgeon  
Lead Clinician  
Maxillofacial Unit  
Queen Elizabeth University Hospital  
Glasgow, UK.

**Nick McIvor MBChB FRACS**

Head and Neck Surgeon  
Auckland Regional Head and Neck Service  
Auckland District Health Board  
New Zealand.

**Ian C Mackenzie BDS FDSRCS PHD**

Professor of Stem Cell Science  
Blizard Institute  
Barts and The London Medical School  
Queen Mary University of London  
London, UK.

**Alistair McNarry MA FRCA**

Consultant Anaesthetist  
Department of Anaesthesia  
Western General and St John's Hospitals  
NHS Lothian  
Edinburgh, UK.

**Samit Majumdar MBBS BMEDSCI(HON) FRCS(EDIN) FDS  
RCPS FRCS(ORL)**

Honorary Senior Lecturer and Consultant ENT Surgeon  
Fellow, Scottish Patient Safety Programme  
Ninewells Hospital & University of Dundee  
Medical School  
Dundee, UK.

**Stewart G Martin BSc (HONS) MSc PHD**

Associate Professor of Oncology  
MSc Course Director and Head of Translational  
Radiation, Biology Research Group  
University of Nottingham  
Academic Division of Clinical Oncology  
Nottingham University Hospitals  
Nottingham, UK.

**Louise Melia MBChB MRCS FRCS ORL-HNS**

Specialty Registrar  
Department of Otolaryngology  
Glasgow Royal Infirmary  
Glasgow, UK.

**Hisham M Mehanna PHD BMEDSC MBChB FRCS  
FRCS (ORL-HNS)**

Chair, Head and Neck Surgery; and  
Director, Institute of Head and Neck Studies and  
Education (InHANSE)  
Institute of Head and Neck Studies and Education  
Institute of Cancer and Genomic Sciences  
University of Birmingham  
Birmingham, UK.

**David Miles FRCP MD**

Professor  
Medical Oncology Consultant  
Mount Vernon Cancer Centre  
Northwood, London, UK.

**Omar Mirza MBChB MA MRCS DOHNS**

Specialty Registrar in ORL-HNS  
Wrightington, Wigan and Leigh NHS Foundation Trust  
Health Education North West  
Wigan, UK.

**Ram Moorthy FRCS (ORL-HNS)**

Consultant ENT/Head & Neck Surgeon  
Wexham Park Hospital  
Frimley Health NHS Foundation Trust  
Slough, UK.

**David AL Morgan FRCR**

Consultant Clinical Oncologist  
Department of Clinical Oncology  
Nottingham University Hospitals  
Nottingham, UK.

**David K Morrissey MBBS(HONS) FRACS**

Otolaryngology, Head and Neck Surgeon  
Department of Otolaryngology, Head and Neck Surgery  
The Queen Elizabeth Hospital  
Adelaide  
School of Medicine  
The University of Queensland  
St Lucia, Australia.

**Laura Moss FRCP FRCR LLM**

Consultant Clinical Oncologist  
Velindre Hospital  
Cardiff, UK.

**Sai HK Murng MB BS MRCP(UK) FRCPATH**

Consultant Immunologist  
Immunology Department  
NHS Greater Glasgow and Clyde Glasgow, UK.

**Michael F Murphy MD, FRCP, FRCPATH, FFPATH**

Professor of Transfusion Medicine  
University of Oxford; and  
Consultant Haematologist  
NHS Blood & Transplant and Oxford University  
Hospitals NHS Foundation Trust  
Oxford, UK.

**Michael Murray FRCA FFICM**

Consultant Anaesthetist  
Department of Anaesthesia  
Southern General Hospital  
Glasgow, UK.

**Jameel Muzaffar BA(HONS) MBBS(HONS) MSC DO-HNS  
MRCS(ENT)**

Specialist Registrar & Honorary Research Fellow  
Institute of Head and Neck Studies and Education  
(InHANSE)  
University of Birmingham & University Hospitals  
Birmingham NHS Foundation Trust  
Birmingham, UK.

**Salil Nair MD FRCS (ORL-HNS)**

Consultant Rhinologist & Honorary Senior Lecturer  
Manukau SuperClinic and Auckland University  
Hospitals  
Auckland, New Zealand.

**Paul Nankivell PHD FRCS (ORL-HNS)**

NIHR Clinical Lecturer, Otolaryngology Head and  
Neck Surgery  
School of Cancer Sciences, University of Birmingham  
Queen Elizabeth Hospital  
Birmingham, UK.

**Ravinder Singh Natt BSC DO-HNS FRCS (ORL-HNS)**

Consultant Otorhinolaryngologist/Head & Neck  
Surgeon  
Department of Otolaryngology  
Guy's and St Thomas' Hospitals  
London, UK.

**Kurt Busuttill Naudi DDS MED LTHE BCHD FDS RCSED MFDS  
RCPS MSURGDENT PGCAP FHEA**

Senior Clinical University Teacher / Honoray Consultant  
in Oral Surgery  
Dental School, School of Medicine  
College of Medical, Veterinary and Life Sciences  
University of Glasgow  
Glasgow, UK.

**Nigel KF Koo Ng MA BM BCH MFSTED FRCS (ORL-HNS)**

Rhinology and Facial Plastic Surgery Fellow  
St George's University Hospitals NHS  
Foundation Trust  
London, UK.

**Iain J Nixon MBCHB FRCS (ORL-HNS) PHD**

Consultant Otorhinolaryngologist Head and Neck  
Surgeon  
University of Edinburgh  
NHS Lothian Department of ENT/Head and Neck  
Surgery  
Edinburgh Royal Infirmary  
Edinburgh, UK.

**Gregory O'Neill BSC(HONS) MBCHB FRCR**

Consultant Radiologist  
Glasgow Royal Infirmary  
Glasgow, UK.

**Bradley A Otto MD**

Assistant Professor  
Director, Division of Rhinology  
Department of Otolaryngology, Head & Neck Surgery  
Wexner Medical Center at The Ohio State University  
Columbus, Ohio, USA.

**James N Palmer MD**

Associate Professor  
Department of Otorhinolaryngology, Head and Neck  
Surgery  
University of Pennsylvania  
Philadelphia, USA.

**Eric K Parkinson PHD**

Professor of Head & Neck Cancer  
Centre for Clinical & Diagnostic Oral Sciences  
Institute of Dentistry, Barts & The London School of  
Medicine and Dentistry  
Queen Mary University of London  
London, UK.

**Mihir R Patel MD**

Assistant Professor  
Department of Otolaryngology, Head & Neck Surgery  
Wexner Medical Center at The Ohio State University  
Columbus, Ohio, USA.

**Simon Paterson-Brown MBBS MPHIL MS FRCS(EDIN)  
FRCS(ENGL) FCS(HK)**

Consultant General and Upper Gastrointestinal Surgeon  
Royal Infirmary of Edinburgh; and  
Honorary Senior Lecturer  
Edinburgh University  
Edinburgh, UK.

**Sarah Payne MRCP PHD**

Medical Manager Oncology, Pfizer UK and Honorary  
Medical Oncology Consultant  
Guy's and St Thomas' NHS Foundation Trust  
Medical Oncology  
Mount Vernon Hospital  
London, UK.

**Kate Pendry BSC MBCHB FRCP FRCPATH**

Consultant Haematologist  
NHS Blood and Transplant  
Manchester Blood Centre  
Manchester, UK.



**Carl Philpott MB CHB DLO FRCS (ORL-HNS) MD PGCME**  
Professor of Rhinology & Olfactology and Head of  
Rhinology & ENT Research Group Professionalism  
Lead

Norwich Medical School; and  
Honorary Consultant Rhinologist and ENT Surgeon  
James Paget University Hospital  
Great Yarmouth, UK.

**Richard J Powell MMBS DM FRCP FRCPATH**  
Consultant and Professor in Clinical Immunology  
School of Life Sciences  
University of Nottingham  
Queen's Medical Centre  
Nottingham, UK.

**Daniel M Prevedello MD FACS**  
Professor  
Department of Neurological Surgery  
The Ohio State University  
Columbus, Ohio, USA.

**Alkis J Psaltis MBBS(HONS) PHD FRACS**  
Associate Professor, Division of Surgery, University of  
Adelaide; and  
Head of Department of Otolaryngology, Head and Neck  
Surgery  
The Queen Elizabeth Hospital  
Department of Surgery, Division of ENT  
The University of Adelaide  
Adelaide, Australia.

**M Shahed Quraishi OBE FRCS FRCS (ORL-HNS)**  
Consultant Otolaryngologist, Thyroid and Parathyroid  
Surgeon  
Director, ENT Masterclass®; and  
Visiting Professor Capital Medical University, Beijing; and  
Honorary Senior Lecturer in Surgical Oncology,  
University of Sheffield  
Doncaster Royal Infirmary  
Doncaster, UK.

**Parameswaran Rajeev BSC MBBS FRCSI MPHIL FRCS UK**  
Senior Consultant and Assistant Professor in Endocrine  
Surgery  
National University of Singapore  
National University Hospital, Singapore.

**Yujay Ramakrishnan MBBCHIR MA CANTAB FRCS  
(ORL-HNS)**  
ENT Skull Base Consultant  
Queen's Medical Centre  
Nottingham, UK.

**Gregory W Randolph MD FACS FACE**  
The Claire and John Bertucci Endowed Chair in Thyroid  
Surgical Oncology  
Harvard Medical School; and  
Director General and Thyroid/Parathyroid Surgical  
Divisions  
Massachusetts Eye and Ear Infirmary  
Boston, Massachusetts, USA.

**Baskaran Ranganathan DOHNS FRCS (ORL-HNS)**  
Consultant ENT  
Manchester University Foundation NHS Trust  
Manchester, UK.

**Urmila Ratnasabapathy FRCA**  
Consultant Neuroanaesthetist  
Department of Anaesthesia, Southern General Hospital  
NHS Greater Glasgow and Clyde Glasgow, UK.

**Penelope Redding MB BS FRCPATH**  
Consultant Microbiologist  
Southern General Hospital  
NHS Greater Glasgow and Clyde  
Glasgow, UK.

**Joanne Rimmer FRCS (ORL-HNS)**  
Specialist Registrar in Otolaryngology  
Royal National Throat Nose & Ear Hospital  
London, UK.

**Neil D Ritchie PHD MBChB MRCP(UK)(INFECTIOUS  
DISEASES)**  
Consultant Physician in Acute Medicine and  
Infectious Diseases  
Queen Elizabeth University Hospital  
Glasgow, UK.

**Alasdair Robertson FRCS ORL DLO MB CHB**  
Consultant ENT Surgeon  
NHS Greater Glasgow and Clyde  
Glasgow, UK.

**Andrew Robson FRCS (ORL)**  
ENT Consultant  
North Cumbria University Hospitals NHS Trust  
Director of Education ENTUK  
Carlisle, UK

**Robert Ross Russell MD FRCPCH FHEA**  
Consultant in Paediatric Respiratory Medicine  
Addenbrooke's Hospital  
Cambridge, UK.

**Raymond Sacks MBBCH FCS(SA)ORL FRACS FARS**  
Professor and Head of ORL/Head & Neck Surgery  
Macquarie University; and  
Clinical Professor and Head, ENT Surgery  
University of Sydney; and  
Past-President Australian and New Zealand Rhinologic  
Society  
Australia.

**Gregory P Sadler MD FRCS GEN SURG**  
Consultant Endocrine Surgeon  
Department of Endocrine Surgery  
John Radcliffe Hospital  
Oxford, UK.

**Hesham Saleh FRCS (ORL-HNS)**  
Consultant Rhinologist/Facial Plastic Surgeon and  
Honorary Senior Lecturer  
Charing Cross and Royal Brompton Hospital  
Imperial College  
London, UK.

**Bal Sanghera PHD MSC**

Clinical Scientist  
Paul Strickland Scanner Centre Mount Vernon Hospital  
Northwood, UK.

**Thozhukat Sathyapalan MD FACP FRCP**

Reader in Endocrinology and Honorary Consultant  
Head of Academic Endocrinology, Diabetes and  
Metabolism  
Hull York Medical School  
University of Hull  
Hull, UK.

**Bernhard Schick MD**

Head of Department of ENT  
Department of Otorhinolaryngology  
Saarland University Medical Center  
Hamburg, Germany.

**David M Scott-Coombes MS FRCS EBSQ**

Consultant Endocrine Surgeon  
University Hospital of Wales  
Cardiff, UK

**Christopher D Scrase MRCP(UK) FRCR**

Macmillan Consultant Clinical Oncologist and  
Honorary Senior Lecturer  
The Ipswich Hospital NHS Trust  
Ipswich, UK.

**Louise Selby BSC MBBS MRCPCH**

Paediatric Respiratory Registrar  
Royal Brompton Hospital  
London, UK.

**Neeraj Sethi MBCHB FRCS**

Clinical Research Fellow  
Leeds Institute of Cancer & Pathology  
Leeds, UK.

**Ashok R Shaha MD**

Professor of Head and Neck Surgery and Oncology  
Head and Neck Service  
Memorial Sloan Kettering Cancer Center New York  
New York, USA.

**Neil Sharma MBCHB PHD MRCS DOHNS**

NIHR Academic Clinical Lecturer  
Centre for Endocrinology, Diabetes and Metabolism  
Institute of Metabolism and Systems Research  
College of Medical and Dental Sciences  
University of Birmingham; and  
Specialty Registrar, Otolaryngology  
Department of Otolaryngology  
University Hospital Birmingham NHS Foundation Trust  
Birmingham, UK.

**Seiji B Shibata MD PHD**

Resident Physician  
Department of Otolaryngology, Head and Neck Surgery  
University of Iowa  
Iowa City, Iowa, USA.

**Uttam Shiralkar FRCS MS MRCPsych**

Consultant Psychiatrist and Psycho-oncologist  
Worcestershire Health and Care Trust  
and Birmingham University Hospital  
Birmingham, UK.

**Ricard Simo FRCS (ORL-HNS)**

Consultant Otorhinolaryngologist Head and Neck  
Surgeon  
Guy's and St Thomas' Hospital NHS Foundation Trust  
Honorary Senior Lecturer  
Guy's, King's and St Thomas' Medical and Dental  
School  
London, UK.

**Neil G Smart BSC(HONS) MBCHB FFARCSI MBA**

Consultant Anaesthetist  
Honorary Clinical Senior Lecturer  
Department of Anaesthetics  
Queen Elizabeth University Hospital  
Glasgow, UK.

**Joel Anthony Smith MD FRCS MBCHB BMEDSC**

Consultant Head Neck and Thyroid Surgeon  
Royal Devon and Exeter Hospital  
Exeter, UK.

**Wendy Smith BPHARM MBBS DLO FRCS (ORL-HNS)**

Consultant Otorhinolaryngologist  
Department of Otolaryngology  
Kettering General Hospital NHS Trust  
Kettering, UK.

**Carl H Snyderman MD MBA**

Professor  
Department of Otolaryngology and Neurological  
Surgery  
University of Pittsburgh School of Medicine  
Pittsburgh, USA.

**Iain RC Swan MD FRCS**

Consultant Otologist  
Glasgow Royal Infirmary  
Glasgow, UK.

**Andrew C Swift MB CHB CHM FRCS FRCSED**

Consultant Rhinologist and ENT Surgeon  
Honorary Senior Lecturer, University of Liverpool; and  
Honorary Senior Lecturer, Edge Hill University  
Aintree University Hospital  
Liverpool, UK.

**Mohammed-Iqbal Syed MS DLO FRCS (ORL-HNS)**

Consultant Otologist & Skull Base Surgeon and  
Honorary Senior Lecturer  
The Royal Infirmary of Edinburgh  
Edinburgh, UK.

**Mark Sywak MBBS MMedSci(Clin Epi) FRACS**

Associate Professor  
Head of Department, Endocrine Surgery and Surgical  
Oncology  
University of Sydney Endocrine Surgical Unit  
Royal North Shore Hospital  
Sydney, Australia.

**Moira Thomas MB CHB MRCP FRCPath**

Consultant Clinical Immunologist  
Queen Elizabeth University Hospital  
Glasgow, UK.

**Ian Todd MA PhD**

Associate Professor and Reader in Cellular  
Immunopathology  
Director of Studies for MOL  
Faculty of Medicine and Health Sciences  
Queen's Medical Centre  
Nottingham, UK.

**Neil S Tolley MD FRCS DLO**

Consultant ENT-Thyroid Surgeon  
St Mary's Hospital  
London, UK.

**Richard B Townsley MBBS BSc MRCS DOHNS**

Specialty Registrar, Otolaryngology, Head and Neck  
Surgery  
West of Scotland Deanery  
Crosshouse University Hospital  
Kilmarnock, UK.

**Richard W Tsang MD FRCSP(C)**

Professor  
Department of Radiation Oncology  
University of Toronto  
Princess Margaret Cancer Centre  
Toronto, Canada.

**Ralph P Tufano MD MBA FACS**

Charles W Cummings MD Professor  
Co-Director of the Johns Hopkins Hospital  
Multidisciplinary Thyroid Tumor Center; and  
Director of the Division of Head and Neck Endocrine  
Surgery  
Department of Otolaryngology, Head and Neck  
Surgery  
The Johns Hopkins University School of Medicine  
Baltimore, Maryland, USA.

**Jan HP van der Meulen PhD FFPH**

Reader in Clinical Epidemiology  
Health Services Research Unit, London School of  
Hygiene and Tropical Medicine  
London, UK.

**Daphne A Varveris MBChB FRCA**

Consultant Anaesthetist  
Department of Anaesthetics  
Queen Elizabeth University Hospital  
Glasgow, UK.

**Navin Vig MBBS BDS MRCS MFDS**

Clinical Research Fellow and Specialty Registrar (OMFS)  
Blizard Institute  
Barts and The London Medical School  
Queen Mary University of London  
London, UK.

**Sebastian Wallis MB CHB FRCS (ORL-HNS)**

ENT Consultant  
Department of Otolaryngology (Ear, Nose and Throat)  
York Hospital  
York, UK.

**Eric W Wang MD**

Associate Professor  
Department of Otolaryngology  
University of Pittsburgh School of Medicine  
Pittsburgh, USA.

**Adrian T Warfield FRCPath**

Consultant Histo-Cytopathologist, University Hospital  
Birmingham NHS Foundation Trust  
Honorary Senior Clinical Lecturer, University of  
Birmingham  
Birmingham, UK.

**John C Watkinson MSc MS FRCS DLO**

One-Time Honorary Senior Lecturer and Consultant  
ENT/Head and Neck and Thyroid Surgeon, Queen  
Elizabeth Hospital  
University of Birmingham NHS Trust and latterly the  
Royal Marsden and Brompton Hospitals, London, UK  
Currently Consultant Head and Neck and Thyroid  
Surgeon, University Hospital, Coventry and Warwick  
NHS Trust; and  
Honorary Consultant ENT/Head and Neck and Thyroid  
Surgeon, Great Ormond Street Hospital (GOSH)  
Honorary Senior Anatomy Demonstrator, University  
College London (UCL)  
Business Director, Endocrine MDT, The BUPA Cromwell  
Hospital, London, UK.

**Anthony P Weetman MD DSc**

Emeritus Professor of Medicine  
Department of Human Metabolism  
Faculty of Medicine, Dentistry and Health  
University of Sheffield  
Sheffield, UK.

**Martin O Weickert MD FRCP**

Consultant Endocrinology & Diabetes  
University Hospitals Coventry & Warwickshire NHS  
Trust (UHCW)  
Warwickshire Institute for the Study of Diabetes,  
Endocrinology and Metabolism; and  
Visiting Professor  
Centre for Applied Biological and Exercise Sciences  
Coventry University, Coventry, UK; and  
Honorary Associate Professor  
Division of Metabolic and Vascular Health  
Warwick Medical School, University of Warwick  
Coventry, UK.

**Paul S White MBCHB FRACS FRCS (ED)**

Consultant Rhinologist  
 Ninewells Hospital; and  
 Honorary Senior Lecturer, University of Dundee  
 Dundee, UK.

**William Whitmer PHD**

Senior Investigator Scientist  
 MRC/CSO Institute of Hearing Research – Scottish Section  
 Nottingham, UK.

**Mark Williams MA MRCP FRCPATH**

Senior Registrar in Haematology  
 Manchester Specialist Registrar Rotation  
 Manchester, UK.

**Wai Lup Wong BA (HONS) LL.M FRCP FRCR**

Paul Strickland Scanner Centre, Mount Vernon Hospital  
 Northwood  
 Honorary Senior Lecturer  
 University College London  
 London, UK.

**Timothy J Woolford MD FRCS(ORL-HNS)**

Consultant ENT Surgeon and Rhinologist  
 Department of Otolaryngology, Head and Neck Surgery  
 Manchester Royal Infirmary  
 Manchester, UK.

**Peter-John Wormald MD FRACS FRCS (EDIN) FCS (SA) MBCHB**

Chairman and Professor  
 Department of Surgery, Otolaryngology, Head and  
 Neck Surgery; and  
 Professor of Skull Base Surgery  
 Adelaide and Flinders Universities  
 Adelaide, Australia.

**Robert F Wynn BA MD MRCP FRCPATH**

Consultant Paediatric Haematologist  
 Director, Blood and Marrow Transplant Unit; and  
 Honorary Professor of Paediatric Haematology and  
 Cellular Therapy  
 Royal Manchester Children's Hospital and University of  
 Manchester  
 Manchester, UK.

**Emily Young MBCHB MPH FRCS (ORL-HNS)**

Neurotology Fellow  
 St Paul's Rotary Hearing Clinic  
 Vancouver, Canada.

**Karen Young MSC FRCS ORL-HNS**

Clinical Research Fellow  
 Department of Endocrinology  
 William Harvey Research Institute  
 Barts and the London School of Medicine  
 Queen Mary University of London  
 London, UK.

# Foreword

The eighth edition of *Scott-Brown* signals the beginning of a new and exciting era for ear, nose and throat surgeons, and also the end of 10 years of very hard work undertaken by John Watkinson and Ray Clarke, the Editors-in-Chief, their team of subeditors and, not least, the publishers. Whatever subspeciality the current generation of trainees decides to follow, they will all have to read and refer to *Scott-Brown* in order to complete their education and gain accreditation. It will be a constant companion and guide throughout their professional lives.

When asked to write the foreword for this edition, I was immediately reminded that I had read John Ballantyne and John Groves's third edition as a trainee, bought the fourth edition as a senior registrar, written chapters for Alan Kerr and Philip Stell in the fifth edition, edited the *Basic science* volume of the fifth edition and was ultimately Editor-in-Chief of the seventh edition. As each edition takes about 10 years to produce, that makes me very old indeed. John and Ray have one final task as Editors-in-Chief: to recommend their successors to the publishers. That was made easy for me as both of them had proved themselves more than capable with the previous edition, and the eighth edition is now their masterpiece. They can enjoy the next 10 years as thousands of surgeons worldwide recognize and thank them for their industry.

This edition reflects the continued expansion of our speciality into fields that *Scott-Brown* himself could

never have imagined. It lays the groundwork for the current generation to make their contribution that will, no doubt, be prompted by technological developments, an evidence base of what is wise and what is not, together with the experience gained by teamwork with other clinicians in today's multidisciplinary approach to patient care.

Simply looking at the table of contents it is clear to see that our role in endocrine surgery has increased dramatically over the last 10 years. The thyroid and parathyroids now account for 30 chapters. How would *Scott-Brown* have viewed that when the tonsils and adenoids justify just one chapter each, and the sore throat has a mere passing reference? Times have certainly changed and ENT surgery has grown up. We have reflected on our past practices, and the evidence base for our management protocols that was emphasized in the previous edition of *Scott-Brown* has been taken to heart.

I hope that this edition will find its way into every medical library in the world and onto every ENT surgeon's bookshelf. It will serve and guide surgeons throughout the English-speaking world, whether they live in high- or low-income countries. It is said that the tragedy of getting old is that we feel young. Reading these volumes makes me wish that I had my time all over again.

**Michael Gleeson**

# Preface

When we were asked to head up the editorial team for this, the eighth edition of *Scott-Brown*, we were mindful of Michael Gleeson's towering achievement in bringing the seventh edition to fruition. Michael delivered a much-loved text – conceived in the early post-war years when antimicrobials, the operating microscope and the National Health Service were all in their infancy – in an entirely new format that befitted modern surgical scholarship. Authors, editors and readers alike had become acutely conscious of the need to quote high-quality evidence to guide clinical decisions; the concept of grading clinical recommendations – and, by implication, acknowledging gaps in the evidence base of our practice – was born. Recognizing the enormity of Michael's contribution led us into the trap that has befallen every editor who has come before us; we grossly underestimated the task ahead. We had misjudged the pace of change. What began as an 'update' of some outdated chapters became a complete rewrite to reflect the advances that marked the decade between editions, but we were determined to keep the text to a manageable size! In the end, we have 330 chapters, but with a slightly smaller page count than the seventh edition.

The basic science knowledge that underpins our clinical practice is no longer focused just on anatomy and physiology; genetics, molecular biology, new techniques for auditory implantation, information technology, new medical therapies for many old disorders together with seismic changes in endoscopic technology and in medical imaging have transformed our specialty. Today's head and neck surgery would have been unrecognizable to the early authors and editors. Surgical oncologists have recourse to completely different treatment strategies than did their predecessors and now work as part of multidisciplinary teams. They deal with different disease patterns and vastly changed patient expectations. Thyroid and parathyroid surgery has become almost exclusively the domain of the otolaryngologist. Surgery of the pituitary fossa has come within our ambit, as has plastic and reconstructive surgery of the head and neck as well as aesthetic facial surgery. Neurotology, audio-vestibular medicine, rhinology and paediatric otolaryngology are accepted subspecialties, each with its own corpus of knowledge and skills and each warranting a sizeable section of this text. Contemporary otolaryngology is now a collection of subspecialty interests linked by common 'stem' training and a shared passion for looking after patients with disorders of the upper respiratory tract and the head and neck.

There is a view that a single text – even a multivolume tome of this size – cannot cover the entire knowledge base of modern clinical practice. The subspecialist will, of course, need recourse to supplementary reading. The pace of change shows no sign of slowing down, but there is still a need for a comprehensive working text embracing the whole spectrum of our workload. That was the task we set our authors and section editors; we think they have done our specialty proud.

In the new 'digital' editorial world authors create manuscripts on personal computers. They transmit chapters, figures, amendments and revisions across continents and

time zones with a few keystrokes. The bulky packages containing grainy photographic prints and the reams of paper with closely-typed and heavily scored text that accumulated on authors' and editors' desks are a distant memory. References, guidelines and systematic reviews are all available online; the editorial 'red pen' has been replaced by a cursor on the screen. This 'new age' has enabled us to look ever further for expertise. We are proud to have enlisted the support of authors from more than 20 countries for this edition. *Scott-Brown* always enjoyed particular affection and respect in Asia, Australia, Africa and the Middle East. It has been a joy to welcome authors in increasing numbers from many of these parts of the world. We are now a truly global specialty and the eighth edition fully reflects this.

What has not changed is the huge time commitment authors and editors need to make. That time now has to be fitted into an increasingly pressurized work environment. Revalidation, mandatory training, more intense regulatory scrutiny, expanding administrative burdens and ever-expanding clinical commitments leave little time for scholarship. Our section editors are all busy clinicians. They have generously given their time, first instructing authors, cajoling them and then editing their chapters, virtually all of which have been completely rewritten since the last edition. Each author was chosen because of his or her specific clinical and scientific expertise and none has disappointed. Authors and section editors receive no reward other than the satisfaction of knowing that they have made a contribution to teaching and learning in a specialty that has given us all so much professional satisfaction. We are profoundly grateful to them and hope that their endeavours spur the next generation of otolaryngologists to carry on this noble tradition. *Scott-Brown* simply wouldn't happen without this generous and dedicated commitment, unstintingly and graciously given.

It is impossible to produce a book like *Scott-Brown* without the contribution of many individuals working behind the scenes. We would like to express our gratitude to our Publishers, Taylor and Francis, and to the staff who have worked on this project from its early days in 2011 to publication in 2018. In particular we would like to mention Cheryl Brandt who with good humour and patience helped to reel in many of the 330 chapters. Miranda Bromage joined the team in 2016 and her publishing experience and enthusiasm for medical education have helped guide this new edition through its final phases to publication. Finally, we are indebted to Nora Naughton who has dedicated so much more than just her extensive publishing skills to this project. Nora's meticulous attention to detail, combined with her warmth and wisdom have encouraged us all at the end of this endeavour.

We are truly 'passing on the torch' of a huge amount of accumulated knowledge and wisdom; it is this that gives us, the Editors-in-Chief, the greatest pleasure.

Read on and enjoy, our thoughts are yours.

RWC  
JCW

I wish to acknowledge the love, happiness and inspiration that have been passed on to me by both my parents and grandparents. I recognize and value the friendship of my dear friend Ray Clarke who has been with me all the way on this rewarding and worthwhile endeavour. I would specifically like to thank Esme, Helen and William, without whom none of this would have been achievable. Their love and support has helped guide me through the years leading up to the publication of this tome, and my final thanks go to Angela Roberts and Sally Holden for their typing and editing skills.

JCW 2018

Thanks to my wife Mary for her patience and support. My parents, Emmet and Doreen Clarke, both sadly died during the preparation of this book. They would have been proud to have played a part in such a scholarly enterprise.

RWC 2018



*Black Hut on the River Test* – Pastel by W G Scott-Brown – circa 1970. Reproduced by kind permission of Mr Neil Weir, who was presented with the original by the artist.

# A Tribute to Bill Scott-Brown



Walter Graham ('Bill') Scott-Brown. 1897–1987

Walter Graham ('Bill') Scott-Brown was twenty-three when he arrived at Corpus Christi College Cambridge in 1919. One of the generation of young men whose entry to university and the professions was delayed by their participation in the First World War, he had joined the Gunners in 1915 as an 18-year-old. He considered himself blessed to have survived – although wounded – when so many of his contemporaries never returned from the Front. In those early post-WW1 years the medical school at St Bartholomew's ('Barts') in London was keen to attract 'gentlemen'. To this end a series of scholarships – 'Shuter's scholarships' – was established to lure those with humanities degrees from Oxford and Cambridge into medicine. It was via this scheme that the young Scott-Brown qualified MB, BCh in 1925. By now married to Margaret Bannerman, one of the very few women medical graduates of her generation, the two established a general practice in Sevenoaks, Kent. His work here involved looking after children with poliomyelitis, which was then commonplace, and his MD thesis was on polio-related bulbar palsy. It earned him the Copeman Medal for research from the University of Cambridge. While working in general practice, Bill pursued his interest in the then fledgling specialty of otolaryngology, securing fellowships from London and Edinburgh. Postgraduate training was haphazard; there were no structured programmes or even junior posts, so the young Scott-Brown was fortunate to be awarded a Dorothy Temple Cross Travelling Fellowship. Mrs Florence Temple Cross had set up these awards (now administered by the Medical Research Council) in memory of her daughter, who died in 1927 aged thirty-two.

They were made available to young physicians to help them travel to overseas centres specifically to study tuberculosis, then rampant and one of the commonest causes of death in young adults. The young Scott-Brown visited the leading pioneers of the day in Berlin, Vienna, Budapest, Stockholm, Copenhagen, Madrid and Venice. Here he developed his considerable endoscopy skills. He reported that his first bronchoscopies were done on a Venetian street entertainer who, for a few coins, would inhale sundry objects that the doctors would then dexterously retrieve from his main stem and segmental bronchi – without of course any anaesthesia!

Times were lean on Scott-Brown's return. Margaret ('Peggy') was now a popular and well-established GP who supported him as his private practice developed. Eventually he secured appointments at East Grinstead, the Royal National and Royal Free Hospitals. He had a thriving Harley Street practice and was the favoured otolaryngologist of the aristocracy. His reputation was such that he became laryngologist to the Royal family, was appointed Commander of the Victorian Order and was a particular favourite of the then Princess Royal, HRH Mary the Countess of Harewood.

By 1938 he was wealthy enough to purchase a farm in Buckinghamshire where he bred prize-winning short-horn cattle. Ironmongery and blacksmith work were hard to come by during the war years, so Scott-Brown prided himself on his ability to make his own agricultural implements, cartwheels and farm wagons in a makeshift forge he himself established on the farm. He would while away endless hours here at weekends following a busy week in London. An accomplished fly fisherman, he was part of the exclusive Houghton Club whose members fished the River Test in Hampshire, where he numbered aristocrats including the Prince of Wales among his circle.

Scott-Brown's celebrated textbook came about in the early 1950s, when he became ill with jaundice and heart trouble. He was advised to rest, and took 6 months off work. Not satisfied with editing what has become the standard UK textbook, he took up painting as well. He became a celebrated artist whose work is still prized in many private collections. One of his pastels is reproduced on the preceding page.

Bill Scott-Brown lived to be 90. He died in July 1987, six weeks after his beloved Peggy and just as the fifth edition of the celebrated textbook that still bears his name was going to press. His legacy lives on in the pages of this book, and we are proud to continue the tradition of scholarship and learning which he established all those years ago.

We would like to thank Martin Scott-Brown for his help in compiling the biography above.

John C. Watkinson and Raymond W. Clarke  
*London, 2018*



# Acknowledgements

*We acknowledge our debt of gratitude to the many authors who have contributed to previous editions of Scott-Brown's Otorhinolaryngology, and in particular to authors from the seventh edition, published in 2008.*

**Chapter 1, Molecular biology**, contains some material from 'Molecular biology' by Michael Kuo and Richard M Irving. The material has been revised and updated by the current author.

**Chapter 3, Gene therapy**, contains some material from 'Gene therapy' by Scott M Graham and John H Lee. The material has been revised and updated by the current author.

**Chapter 5, Radiotherapy and radiosensitizers**, contains some material from 'Radiotherapy and radiosensitizers' by Stewart G Martin and David AL Morgan. The material has been revised and updated by the current author.

**Chapter 11, Skin flap physiology**, contains some material from 'Skin flap physiology' by A Graeme B Perks. The material has been revised and updated by the current author.

**Chapter 15, Evaluation of the immune system**, contains some material from 'Evaluation of the immune system' by Elizabeth Drewe and Richard J Powell. The material has been revised and updated by the current author.

**Chapter 31, Recognition and management of the difficult airway**, contains some material from 'Recognition and management of the difficult airway' by Adrian Pearce. The material has been revised and updated by the current author.

**Chapter 34, Paediatric intensive care**, contains some material from 'Paediatric intensive care' by Helen Allen and Rob Ross Russell. The material has been revised and updated by the current author.

**Chapter 41, Epidemiology**, contains some material from 'Epidemiology' by Jan HP van der Meulen and David A Lowe. The material has been revised and updated by the current author.

**Chapter 44, Critical appraisal skills**, contains some material from 'Critical appraisal skills' by Martin Dawes. The material has been revised and updated by the current author.

**Chapter 45, Electrophysiology and monitoring**, contains some material from 'Electrophysiology and monitoring' by Patrick R Axon and David M Baguley. The material has been revised and updated by the current author.

**Chapter 46, Optical coherence tomography**, contains some material from 'Optical coherence tomography' by Mariah Hahn and Brett E Bouma. The material has been revised and updated by the current author.

**Chapter 92, Non-allergic rhinitis**, contains some material from 'Nonallergic perennial rhinitis' by Claus Bachert. The material has been revised and updated by the current author.

**Chapter 107, Nasal and facial fractures**, contains some material from 'Nasal fractures' by Brent A McMonagle and Michael Gleeson and 'Fractures of the facial skeleton' by Simon Holmes and Michael Gleeson. The material has been revised and updated by the current author.

**Chapter 109, Granulomatous conditions of the nose**, contains some material from 'Granulomatous conditions of the nose' by David J Howard and Valerie J Lund. The material has been revised and updated by the current author.

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# Abbreviations

<b>1,25-DHCC</b>	1,25 dihydroxycholecalciferol	<b>AEF</b>	auditory-evoked cortical magnetic field
<b>2D</b>	two-dimensional	<b>AERD</b>	aspirin-exacerbated respiratory disease
<b>3D</b>	three-dimensional	<b>AES</b>	assigned educational supervisors
<b>4D CT</b>	four-dimensional computed tomography	<b>AF</b>	atrial fibrillation; <i>or</i> anterior fontanelle
<b>5-FdUMP</b>	5-fluoro-2 deoxyuridine monophosphate	<b>AFAP</b>	attenuated familial adenomatous polyposis
<b>5-FU</b>	5-fluorouracil	<b>AFIP</b>	Armed Forces Institute of Pathology
<b>5-FUMP</b>	5-fluorouridine monophosphate	<b>AFOI</b>	awake fibre-optic intubation
<b>6MP</b>	6-mercaptopurine	<b>AFRS</b>	allergic fungal rhinosinusitis
<b>18-FDG</b>	2-18-fluoro-2-deoxy-D-glucose	<b>AFU</b>	angiofollicular unit
<b>A</b>	adenine; <i>or</i> anterior	<b>AGHDA</b>	Assessment of Growth Hormone Deficiency in Adults
<b>AACE</b>	American Association of Clinical Endocrinologists	<b>AH</b>	adenohypophysis
<b>AAES</b>	American Association of Endocrine Surgeons	<b>AICA</b>	anterior inferior cerebellar artery
<b>AAGBI</b>	Association of Anaesthetists of Great Britain and Ireland	<b>AIDS</b>	acquired immunodeficiency syndrome
<b>AAOHNS</b>	American Academy of Otolaryngologists/Head and Neck Surgeons	<b>AIFR</b>	acute invasive fungal rhinosinusitis
<b>AAV</b>	adeno-associated virus	<b>AIFS</b>	acute invasive fungal sinusitis
<b>ABBA</b>	axillo-bilateral-breast approach	<b>AIH</b>	amiodarone-induced hypothyroidism
<b>ABG</b>	arterial blood gas	<b>AIP</b>	aryl hydrocarbon receptor-interacting protein
<b>ABPA</b>	allergic bronchopulmonary aspergillosis	<b>AIT</b>	amiodarone-induced thyrotoxicosis
<b>ABR</b>	auditory brainstem response	<b>AJCC</b>	American Joint Committee on Cancer
<b>ABRS</b>	acute bacterial rhinosinusitis	<b>AKT</b>	serine/threonine kinase
<b>AC</b>	air conduction; <i>or</i> alternating coupled; <i>or</i> adenylate cyclase	<b>ALLO</b>	alloimmunization
<b>ACE</b>	angiotensin-converting enzyme	<b>ALTB</b>	acute laryngotracheobronchitis
<b>ACGME</b>	Accreditation Council for General Medical Education	<b>ALS</b>	advanced life support; <i>or</i> amyotrophic lateral sclerosis; <i>or</i> acid labile subunit
<b>ACh</b>	acetylcholine	<b>ALSPAC</b>	Avon Longitudinal Study of Parents and Children
<b>AchR</b>	acetyl choline receptor	<b>ALTB</b>	acute laryngotracheobronchitis
<b>ACR</b>	America College of Rheumatology	<b>AML</b>	acute myeloid leukaemia
<b>ACT</b>	Aid for Children with Tracheostomies	<b>AMR</b>	antimicrobial resistance
<b>ACTH</b>	adrenocorticotrophic hormone	<b>AN</b>	acoustic neuroma; <i>or</i> auditory neuropathy; <i>or</i> audiovestibular nerve
<b>AD</b>	Alzheimer's disease	<b>ANA</b>	anti-nuclear antibody
<b>ADAM-33</b>	A disintegrin and metalloprotease 33k	<b>AN/AD</b>	auditory neuropathy/auditory dyssynchrony
<b>ADC</b>	apparent diffusion coefficient	<b>ANCA</b>	antineutrophil cytoplasmic antibody
<b>ADCC</b>	antibody-dependent cellular cytotoxicity	<b>ANSD</b>	auditory neuropathy spectrum disorder
<b>ADH</b>	antidiuretic hormone	<b>ANTS</b>	anaesthetists' non-technical skills
<b>ADP</b>	adenosine diphosphate	<b>AOM</b>	acute otitis media
<b>ADR</b>	adverse drug reaction	<b>AON</b>	anterior olfactory nucleus
<b>ADU</b>	avoidable, delayed or undertransfusion	<b>AP</b>	anterior-posterior; <i>or</i> action potential
<b>Ad-VEGF</b>	adenovirus-encoding vascular endothelial growth factor	<b>APACHE</b>	acute physiology and chronic health evaluation
<b>AEA</b>	anterior ethmoidal artery	<b>APC</b>	antigen presenting cell; <i>or</i> activated protein C; <i>or</i> argon plasma coagulation; <i>or</i> adenomatous polyposis coli
<b>AED</b>	aerodynamic equivalent diameter		

<b>APIT</b>	activated partial thromboplastic time	<b>BMD</b>	bone mineral density
<b>APOF</b>	aggressive psammomatoid ossifying fibroma	<b>BMI</b>	body mass index
<b>APTT</b>	activated partial thromboplastin time	<b>BMP</b>	bone morphogenetic protein; <i>or</i> bone morphogenic protein
<b>AQP2</b>	aquaporin 2	<b>BOR</b>	brachio-oto-renal; <i>or</i> branchioterrenal syndrome
<b>ARIA</b>	allergic rhinitis and its impact on asthma	<b>BP</b>	blood pressure
<b>ARR</b>	absolute risk reduction	<b>BPE</b>	bilateral parathyroid exploration
<b>ARS</b>	acute rhinosinusitis	<b>BSAC</b>	British Society for Antimicrobial Chemotherapy
<b>ARSAC</b>	Administration of Radioactive Substances Advisory Committee	<b>BTA</b>	British Thyroid Association
<b>ART</b>	advanced rotating tomograph; <i>or</i> antiretroviral therapy	<b>C</b>	cytosine
<b>ARIA</b>	allergic rhinitis and its impact on asthma	<b>CAD</b>	caspase-activated DNase
<b>ARS</b>	acute rhinosinusitis	<b>CAF</b>	carcinoma-associated fibroblasts
<b>ASA</b>	aspirin-induced asthma; <i>or</i> aspirin-sensitive asthma; <i>or</i> American Society of Anesthesiologists	<b>CAG</b>	curriculum advisory group
<b>a-SCC</b>	anterior semicircular canal	<b>cAMP</b>	3',5'-monophosphate
<b>AT</b>	ataxia telangiectasia; <i>or</i> auditory therapy or training; <i>or</i> autotransplantation	<b>CAP</b>	compound action potential; <i>or</i> category of auditory performance; <i>or</i> College of American Pathologists
<b>ATA</b>	American Thyroid Association	<b>CAPS</b>	cryopyrin-associated periodic syndromes
<b>ATC</b>	anaplastic thyroid carcinoma; <i>or</i> air traffic control	<b>CARD</b>	caspase recruitment domain
<b>ATD</b>	ascending tract of Deiters; <i>or</i> adult therapeutic dose	<b>CaSR</b>	calcium-sensing receptor
<b>ATIII</b>	antithrombin III	<b>CASTLE</b>	carcinoma with thymus-like elements
<b>ATP</b>	adenosine triphosphate	<b>CBD</b>	case-based discussion
<b>ATR</b>	acute transfusion reaction	<b>CBF</b>	ciliary beat frequency
<b>AVP</b>	arginine vasopressin	<b>CCD</b>	charge-coupled device
<b>AW</b>	anterior wall	<b>CCH</b>	C-cell hyperplasia
<b>BABA</b>	bilateral axillo-breast approach	<b>CCP</b>	cyclic citrullinated peptide
<b>BAC</b>	bacterial artificial chromosome	<b>CCR</b>	chemokine receptor
<b>BADS</b>	British Association of Day Surgery	<b>CCT</b>	Certificate of Completion of Training
<b>BAES</b>	British Association of Endocrine Surgeons	<b>CD</b>	cluster of differentiation; <i>or</i> colloid droplets; <i>or</i> compact disk; <i>or</i> Cowden's disease
<b>BAETS</b>	British Association of Endocrine and Thyroid Surgeons	<b>CDA</b>	cold dry air
<b>BAT</b>	basophil activation test	<b>CDAD</b>	<i>C. difficile</i> associated diarrhoea
<b>BCHD</b>	bone conduction hearing device	<b>CDC</b>	Centers for Disease Control and Prevention
<b>BCP</b>	biphasic calcium phosphate	<b>CDI</b>	<i>C. difficile</i> infection
<b>BCSH</b>	British Committee for Standards in Haematology	<b>CDK</b>	cyclin-dependent kinase
<b>BD</b>	Behcet's disease	<b>CEA</b>	carcinoembryonic antigen
<b>BDP</b>	beclomethasone dipropionate	<b>CEFTE</b>	carcinoma of the thyroid with Ewing family tumour elements
<b>BFU-E</b>	burst-forming unit erythroid	<b>CEPOD</b>	Confidential Enquiry into Perioperative Deaths
<b>BiPAP</b>	bilevel positive airway pressure	<b>CER</b>	control event rate
<b>BIPP</b>	bismuth and iodoform paraffin paste	<b>CESR</b>	Certificate of Eligibility for Specialist Registration
<b>BIS</b>	bispectoral index	<b>CEX</b>	clinical evaluation exercises
<b>BL</b>	Burkitt's lymphoma	<b>CF</b>	cystic fibrosis; <i>or</i> characteristic frequency
<b>BMA</b>	British Medical Association; <i>or</i> bone marrow aspirate	<b>CFD</b>	colour-flow duplex Doppler; <i>or</i> computational fluid dynamics
		<b>c-FOS</b>	fos proto-oncogene

CFTR	cystic fibrosis transmembrane conductance regulator	CREST	calcinosis, Raynaud's, oesophageal involvement, sclerodactyly, telangiectasis
CG	clinical governance	CRF	corticotrophin-releasing factor
CGD	chronic granulomatous disease	CRH	corticotrophin-releasing hormone
CGH	comparative genomic hybridization	CRH-R	corticotrophin releasing hormone receptor
CGIFS	chronic granulomatous invasive fungal sinusitis	CRP	C-reactive protein; <i>or</i> canalith repositioning procedure
CGRP	calcitonin gene-related peptide	CRS	chronic rhinosinusitis; <i>or</i> congenital rubella syndrome
CHI	Commission for Healthcare Improvement (UK)	CRSwNP	chronic rhinosinusitis with nasal polyps
CHOP-R	cyclophosphamide, doxorubicin, vincristine, prednisone and rituximab	CRSsNP	chronic rhinosinusitis without nasal polyps
CI	cochlear implant; <i>or</i> cardiac index; <i>or</i> confidence interval; <i>or</i> concha inferior	CRS	chronic rhinosinusitis; <i>or</i> congenital rubella syndrome
CICV	can't intubate can't ventilate	CS	corticosteroid; <i>or</i> cell salvage and autologous transfusions
CIFR	chronic invasive fungal rhinosinusitis	CSC	cancer stem cells
CIFS	chronic invasive fungal sinusitis	CSF	cerebrospinal fluid
CINCA	chronic infantile neurologic cutaneous articular syndrome	CSS	Churg-Strauss syndrome
CJD	Creutzfeldt–Jakob disease	CST	core surgical training
CK	cytokeratin	CT	computed tomography; <i>or</i> conventional thyroidectomy
CKD-MBD	chronic kidney disease mineral bone disorder	CTA	composite tissue allograft; <i>or</i> computed tomography angiography
CLL	chronic lymphatic leukaemia; <i>or</i> chronic lymphocytic leukaemia	CTLA	cytotoxic T-lymphocyte-associated antigen
CM	concha media; <i>or</i> cochlear microphonic; <i>or</i> cricothyroid muscle	CTM	cricothyroid membrane
CMAP	compound muscle action potential	Cu-ATSM	Cu(II)-diacetyl-bis-N4-methylthiosemicarbozone
CML	chronic myeloid leukaemia	DA	double adenoma
CMSO	complementary metal oxide detectors	DACH	diaminocyclohexane
CMT	Charcot—Marie—Tooth; <i>or</i> combined modality therapy	DAHANCA	Danish Head and Neck Cancer Study
CMV	cytomegalovirus	DAMPs	damage-associated molecular pattern molecules
CN	cranial nerve; <i>or</i> cochlear nuclei; <i>or</i> cochlear nerve	DAS	Difficult Airway Society
CNS	central nervous system	DAT	direct antiglobulin test
CO <sub>2</sub>	carbon dioxide	DAVF	dural arteriovenous fistulas
CoAA	coactivator AA	dB	decibel
COF	cemento-ossifying fibroma	DBM	demineralized bone matrix
COMET	Core Outcomes Measures in Effectiveness Trials	DCE MRI	dynamic contrast-enhanced MRI
CONSORT	Consolidated Standards of Reporting Trials	DCR	dacryocystorhinostomy
COPD	chronic obstructive pulmonary disease	DD	death domain
COSI	Client Oriented Scale of Improvement	DDAVP	desmopressin
COX-2	cyclo-oxygenase 2	DDE	Doctrine of Double Effect
CPA	cerebellopontine angle	DED	death effector domain
CPAP	continuous positive airway pressure	DFN3	deafness type 3
CPD	citrate phosphate dextrose; <i>or</i> continuing professional development	DFO-H	deferoxamine-hesperan
CPG	central pattern generator	DHI	dizziness handicap inventory
CRC	colorectal cancer	DHTR	delayed haemolytic transfusion reaction
CRD	component resolved diagnostics	DI	diabetes insipidus
CREB	cAMO binding element	DIC	disseminated intravascular coagulation
		DIO	deiodinases

DISC	death inducing signal complex	ECP	eosinophil cationic protein; <i>or</i> extra corporeal perfusion
DIT	diiodotyrosine; <i>or</i> di-iodothyronine	ECS	extracapsular spread
DLBCL	diffuse large B-cell lymphoma	EDTA	ethylenediaminetetraacetic acid
DM	diabetes mellitus; <i>or</i> decision making	EEA	endoscopic endonasal approach
DMARDS	disease-modifying anti-rheumatic drugs	EEG	electroencephalography; <i>or</i> electroencephalogram
DMSA	dimercapto succinic acid	EER	experimental event rate
DMSO	dimethylsulfoxide	EFRS	eosinophilic fungal rhinosinusitis
DNA	deoxyribonucleic acid	EFVPTC	encapsulated follicular variant of PTC
DNAR	do not attempt resuscitation	EGF	epidermal growth factor
dNTP	deoxynucleoside triphosphate	eGFR	estimated glomerular filtration rate
DOAC	direct oral anticoagulant	EGFR	epidermal growth factor receptor
DOHNS	Diploma in Otolaryngology – Head and Neck Surgery	ELISA	enzyme-linked immunosorbent assay
DOPS	direct observation of procedural skills	EMG	electromyography
DPA	Data Protection Act (UK)	EMT	epithelial-mesenchymal transition
DPOAE	distortion product otoacoustic emission	EMRS	eosinophilic mucin rhinosinusitis
DR	death receptor; <i>or</i> drug resistance	ENA	extractable nuclear antigen
DSA	digital subtraction angiography	ENoG	electroneurography
dsRNA	double-stranded RNA	ENS	empty nose syndrome
DTC	differentiated thyroid cancer	ENT	ear, nose and throat
DTD	DT-diaphorase	EORTC	European Organisation for Research and Treatment of Cancer
dTMP	deoxythymidine monophosphate	EPO	erythropoietin
dUMP	deoxyuridine monophosphate	EPOS	European position paper on rhinosinusitis and nasal polyps
DVT	deep vein thrombosis	EQ-5D	EuroQol
DWI	diffusion weighted image	ER	enhancement ratio; <i>or</i> endoplasmic reticulum
EAACI	European Academy of Allergology and Clinical Immunology	ER $\alpha$	oestrogen receptor alpha
EAC	external auditory canal; <i>or</i> external acoustic canal	ERAS	enhanced recovery after surgery
EAL	ethmoidal artery ligation	ERK	extracellular signal-regulated kinase
EAT	endoscopic-assisted thyroidectomy	ERS	European Rhinological Society
EBM	evidence-based medicine	ES	embryonic stem; <i>or</i> endolymphatic sac
EBNA	Epstein–Barr virus-associated nuclear antigen	ESBL	extended spectrum $\beta$ -lactamase
EBP	evidence-based practice	ESPAL	endonasal ligation of the sphenopalatine artery
EBRT	external beam radiotherapy	ESR	erythrocyte sedimentation rate
EBSLN	external branch of the superior laryngeal nerve	ESS	endoscopic sinus surgery; <i>or</i> Epworth Sleepiness Scale; <i>or</i> empty sella syndrome
EBUS	endobronchial ultrasound	ET-1	endothelin-1
EBV	Epstein–Barr virus	ETE	extrathyroidal extension
ECA	external carotid artery	ETT	endotracheal tube
ECAL	external carotid artery ligation	EUA	examination under anaesthesia
ECG	electrocardiogram	EUCAST	European Committee on Antimicrobial Susceptibility Testing
ECM	extracellular matrix	EVAL	ethylene-vinyl alcohol copolymer
ECMO	extracorporeal membrane oxygenation	EWTD	European Working Time Directive
ECochG	electrocochleography	Fab	fragment antigen binding
ECog	electrocochleogram	FACT	functional assessment of cancer therapy
ECOG	Eastern Cooperative Oncology Group (USA)	FADD	Fas-associated death domain

Fas-L	Fas ligand	G	guanine
FBC	full blood count	GABA	gamma-aminobutyric acid
Fc	fragment crystallizable	GABHS	group A beta-haemolytic streptococcus
FcεRI	high affinity IgE receptors	GAG	glycosaminoglycan
FD	fibrous dysplasia	GBI	Glasgow Benefit Inventory
FDA	Food and Drug Administration (USA)	GBM	anti-glomerular basement membrane
FDG	fluorodeoxyglucose; <i>or</i> 2-[18F] fluoro-2-deoxy-D-glucose; <i>or</i> F18-fluoro-2-deoxy-D-glucose	GC	glucocorticoid
FD-OCT	fourier domain optical coherence tomography	G-CSF	granulocyte-colony stimulating factor
FDG-PET	2-[18F] fluoro-2-deoxy-D-glucose positron emission tomography; <i>or</i> fluorine-18-labelled deoxyglucose positron emission tomography	GD	Graves' disease
FE	frontal ethmoidal cell	GERD	gastroesophageal reflux disease
FEA	finite element analysis	GH	growth hormone
FESS	functional endoscopic sinus surgery	GHABP	Glasgow Hearing Aid Benefit Profile
FEIBA	factor VIII inhibitor bypassing agent	GHRH	growth hormone-releasing hormone
F-ETNIM	fluorine-18 fluoroerythronitroimidazole	GHRP	growth hormone-releasing peptide
FFP	fresh frozen plasma	GI	gastrointestinal
FGF	fibroblast growth factor	GMC	ganglion mother cell; <i>or</i> General Medical Council (UK)
FHH	familial hypocalciuric hypercalcaemia	GM-CSF	granulocyte-macrophage colony stimulating factor
FIHP	familial isolated hyperparathyroidism	GMS	Gomori's methamine silver stain
FiO <sub>2</sub>	fraction of inspired oxygen	GnIH	gonadotrophin inhibitory hormone
FIPA	familial isolated pituitary adenomas	GnRH	gonadotrophin-releasing hormone
FISH	fluorescence <i>in situ</i> hybridization	GORD	gastro-oesophageal reflux disease
FITS	functional inferior turbinosurgery	gp	glycoprotein
FIV	feline immunodeficiency virus	GP	general practitioner
FLAIR	fluid attenuated inversion recovery	GPA	granuloma; <i>or</i> granulomatosis with polyangitis
FMF	familial Mediterranean fever	G protein	guanine nucleotide-binding regulatory protein
FMISO	fluorine-18 fluoromisonidazole	GRADE	Grading of Recommendation, Assessment, Development and Evaluation
fMRI	functional magnetic resonance imaging	GRB2	growth factor receptor binding protein 2
FMTC	familial medullary thyroid cancer	Gs	G-protein adenylate cyclase stimulator
FN	facial nerve	GSH	glutathione
FNA	fine-needle aspiration/aspirate	GSP	stimulatory G protein
FNAB	fine-needle aspiration biopsy	GTN	nitroglycerin
FNAC	fine-needle aspiration cytology	GTR	guided tissue regeneration
FNHTR	febrile non-haemolytic transfusion reactions	GTV	grow tumour volume
FOI	fibreoptic orotracheal intubation	GvHD	graft-versus-host disease
FRS	fungal rhinosinusitis	HA	hydroxyapatite
FS	folliculostellate; <i>or</i> frontal sinus	H&E	haematoxylin and eosin
FSH	follicle-stimulating hormone	H&N	head and neck
fT <sub>3</sub>	free T <sub>3</sub>	H <sub>2</sub>	histamine receptor type 2
fT <sub>4</sub>	free T <sub>4</sub>	HA	hydroxyapatite
FTC	frequency threshold curve; <i>or</i> follicular thyroid carcinoma	HAART	highly active antiretroviral therapy
FT-UMP	follicular tumour of uncertain malignant potential	HAI	hospital-acquired infection
FVPTC	follicular variant of papillary thyroid carcinoma	HBME-1	mesothelium-associated antibody
		Hb	haemoglobin
		HbA	adult haemoglobin

HBO	hyperbaric oxygen	HSC	haematopoietic stem cell
HBOT	hyperbaric oxygen therapy	HSCT	haemopoietic stem cell transplant
HCG	human chorionic gonadotrophin	HSD	hydroxysteroid dehydrogenase enzyme
HD	haemodialysis	HSE	handling and storage errors
HDL	high-density lipoprotein	HSV	herpes simplex virus
HDM	house dust mite	HSV-1	herpes simplex virus type 1
HDU	high dependency unit	HSV-2	herpes simplex virus type 2
He-Ne	helium-neon	HSV-TK	herpes simplex thymidine kinase
HEPA	high-efficiency particulate air	HTA	hyalinizing trabecular adenoma; <i>or</i> Health Technology Assessment
HES	hospital episode statistics	hTERT	human telomerase reverse transcriptase
HFJV	high-frequency jet ventilation	HTR	haemolytic transfusion reaction
HFT	hereditary familial telangiectasia	HTT	hyalinizing trabecular tumours
HGF	hepatocyte growth factor	HU	Hounsfield unit
HH	hypogonadotrophic hypogonadism	HUI	Health Utilities Index
HHI	Hearing Handicap Inventory; <i>or</i> hereditary hearing impairment	HVDT	health visitor distraction test
HHIE	Hearing Handicap Inventory for the Elderly	Hz	hertz
HHT	hereditary haemorrhagic telangiectasia	HZV	herpes zoster virus
HHV-6	human herpesvirus 6	IAC	internal auditory canal
HHV-8	human herpesvirus 8	IAR	intermittent allergic rhinitis
HI	hearing impaired	IBCT	incorrect blood component transfused
Hib	<i>Haemophilus influenzae</i> b	ICA	internal carotid artery
HIDS	hyper-IgD syndrome	ICAD	inhibitor caspase-activated DNase
HIT	heparin-induced thrombocytopenia	ICAM	intercellular adhesion molecule
HIV	human immunodeficiency virus	ICAM-1	intercellular adhesion molecule 1
HL	hearing loss; <i>or</i> hearing level; <i>or</i> hairy leukoplakia	ICD	International Classification of Disease
HLA	human leukocyte antigen	ICF	International Classification of Functioning
HLH	haemophagocytic lymphohistiocytosis	ICS	intra-operative cell salvage
HM	history of migraine; <i>or</i> hemifacial microsomia	ICU	intensive care unit
HMW	high molecular weight	IDD	intracellular death domain
HMWC	high molecular weight compound	IDT	infant distraction test; <i>or</i> intra-dermal test
HMWCK	high molecular weight cytokeratin	IFN	interferon
HNSCC	head and neck squamous cell carcinoma	IFN- $\alpha$	interferon-alpha
Ho-YAG	holmium yttrium aluminium garnet	IFN- $\beta$	interferon-beta
HPA	hypothalamic-pituitary-adrenal axis	IFN- $\gamma$	interferon-gamma
HPC	haemangiopericytoma	IFVPTC	invasive follicular variant of PTC
HPT	hyperparathyroidism	Ig	immunoglobulin
HPT-JT	hyperparathyroidism-jaw tumour	IGD	isolated GnRH-deficiency
HPV	human papillomavirus; <i>or</i> human herpes virus 8	IgE	immunoglobulin E
HPZ	herpes zoster	IGF	insulin-like growth factor
HR	hazard ratio	IGF-I	insulin-like growth factor 1
HRA	Human Rights Act	IGF-II	insulin-like growth factor II
HRQOL	health-related quality of life	IGFBP	insulin-like growth factor binding protein
HR	heart rate	IgG	immunoglobulin G
HRT	hormone replacement therapy	IgG $\kappa$ C	immunoglobulin G kappa chain
HRV	rhinoviruses	IGS	image-guided surgery
		IHC	immunohistochemistry; <i>or</i> inner hair cell
		IIT	iodide-induced thyrotoxicosis

IJV	internal jugular vein	K	Kirschner
IL	interleukin	KCCT	kaolin cephalin clotting time
IL-1	interleukin-1	KDOQI	Kidney Disease Quality Outcomes Initiative
IL-2	interleukin-2	keV	kilo electron volt
IL-3	interleukin-3	KOH	potassium hydroxide solution prep test
IL-5	high interleukin	KISS-R	kisspeptin receptor (also known as GR54)
IL-6	interleukin-6	KS	Kaposi's sarcoma
ILMA	intubating laryngeal mask airway	KTP	potassium titanyl phosphate
i.m.	intramuscular		
IMA	internal maxillary artery	LA	lymphangioma
IMAL	internal maxillary artery ligation	LAR	local allergic rhinitis
IMF	intermaxillary fixation	LAT	lateral aberrant thyroid
IMP	importin	LC1	liver cystolic 1
IMRT	intensity-modulated radiation therapy	LCM	laser capture microdissection
iNOS	inducible nitric oxide synthase	LDH	lactate dehydrogenase
INR	international normalized ratio; <i>or</i> interventional neuroradiology	LDL	low-density lipoprotein; <i>or</i> loudness discomfort level
IOM	institute of Medicine	LED	light-emitting diode
IONM	intra-operative neural/nerve monitoring	LFA	lymphocyte-function associated antigen
IOPTH	intra-operative PTH	LFJV	low-frequency jet ventilation
IOQPTH	intra-operative quick assay of intact parathyroid hormone	LH	luteinizing hormone
IOUS	intra-operative ultrasound scanning	LHB	lateral-head-back
IP	Inverted papilloma	LH-R	luteinizing hormone receptor
IPD	invasive pneumococcal disease	LINAC	linear accelerator
iPSCs	induced pluripotency stem cells	LKM	liver kidney microsomal
IPSS	inferior petrosal sinus sampling	LM	laryngeal mask
IRI	ischaemia-reperfusion injury	LMA	laryngeal mask airway
IRS	insulin receptor substrate; <i>or</i> Intergroup Rhabdomyosarcoma Study	LMW	low molecular weight
ISAAC	International Study of Asthma and Allergies in Childhood	LMWC	low molecular weight compound
ISCP	International Surgical Curriculum Programme	LMWH	low molecular weight heparin
ISL	<i>in situ</i> ligation	LOCR	lateral opticocarotid recess
IT	inferior turbinate	LOD	logarithm to the base 10 of the odds that the markers are linked at a recombination distance of N centimorgans
ITA	inferior thyroid artery	LOH	loss of heterozygosity
ITAM	immunoreceptor tyrosine-based activation motif	LP	lamina papyracea; <i>or</i> lichen planus; <i>or</i> lymphocyte predominant
ITP	idiopathic thrombocytopenic purpura	LPR	laryngopharyngeal reflux; <i>or</i> late phase reactions
ITU	intensive therapy unit	LR	likelihood ratio
i.v.	intravenous	LR-OCT	long-range optical coherence tomography
IVIg	intravenous immunoglobulin	LTRA	leukotriene receptor antagonists
		LW	lateral wall
JAK2	Janus kinase 2	M	metastases
JCST	Joint Committee on Surgical Training	M2	Matrix 2 ion channels
JIA	juvenile idiopathic arthritis	MAb	monoclonal antibodies
JLNS1	Jervell and Lange-Nielsen syndrome	MABP	mean arterial blood pressure
JNA	juvenile nasopharyngeal angiofibroma	MAC	membrane attack complex; <i>or</i> <i>Mycobacterium avium</i> complex
JNK	c-Jun N-terminal kinase		



<b>MAD</b>	mucosal atomization device	<b>MMR</b>	measles, mumps and rubella
<b>MAGE-3</b>	melanoma-associated antigen-3	<b>MNG</b>	multinodular goitre
<b>MALT</b>	mucosa-associated lymphoid tissue	<b>MOCR</b>	medial opticocarotid recess
<b>MAOI</b>	monoamine oxidase inhibitor	<b>MODS</b>	multiple organ dysfunction syndrome
<b>MAP</b>	minimum audible pressure	<b>MOE</b>	malignant otitis externa
<b>MAPK</b>	mitogen-activated protein kinase	<b>MOFT</b>	multiple oxyphil follicular tumour
<b>MAS</b>	mandibular advancement splint; <i>or</i> macrophage activation syndromes	<b>MOS</b>	Medical Outcomes Study
<b>MBL</b>	mannose-binding lectin	<b>MPA</b>	microscopic polyangiitis
<b>MC2R</b>	melanocortin 2 receptor	<b>MPTS</b>	Medical Practitioner's Tribunal Service
<b>MCP</b>	monocyte chemotactic protein	<b>MPO</b>	myeloperoxidase
<b>MCS</b>	mental component summary	<b>MPTP</b>	1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine
<b>MCT</b>	medullary thyroid carcinoma	<b>MR</b>	magnetic resonance
<b>MCV</b>	mean corpuscular volume	<b>MRA</b>	magnetic resonance angiography
<b>MDCT</b>	multi-detector row CT	<b>MRC</b>	Medical Research Council (UK)
<b>MDS</b>	myelodysplastic syndrome	<b>MRCs</b>	Member of the Royal College of Surgeons
<b>MDT</b>	multidisciplinary team	<b>MRI</b>	magnetic resonance imaging
<b>ME</b>	middle ear	<b>mRNA</b>	messenger ribonucleic acid
<b>MEG</b>	magnetoencephalography	<b>MRSA</b>	methicillin-resistant <i>Staphylococcus aureus</i>
<b>MEK</b>	MAPK/extracellular signal related kinase	<b>MS</b>	multiple sclerosis
<b>MEN</b>	multiple endocrine neoplasia	<b>MSBOS</b>	maximum surgical blood ordering schedule
<b>MeSH</b>	medical subject heading	<b>MSCs</b>	mesenchymal stem cells
<b>MET</b>	middle ear transducer	<b>MSF</b>	multi-source feedback
<b>M-FISH</b>	multifluor FISH	<b>MSSA</b>	methicillin-susceptible strains
<b>MGHD</b>	multiple gland hyperplasia disease	<b>MT</b>	maxilloturbinal; <i>or</i> middle turbinate
<b>MGO</b>	methylglyoxal	<b>MTC</b>	medullary thyroid carcinoma
<b>MGUS</b>	monoclonal gammopathy of uncertain significance	<b>mtDNA</b>	mitochondrial DNA
<b>MHC</b>	major histocompatibility complex	<b>mTORC1</b>	mammalian target of rapamycin 1
<b>MHP</b>	massive haemorrhage pack	<b>mTORC2</b>	mammalian target of rapamycin 2
<b>MI</b>	myocardial infarction	<b>MW</b>	medial wall
<b>mIBG</b>	metaiodobenzylguanidine; <i>or</i> iodine-123-metaiodobenzylguanidine	<b>N</b>	nodal
<b>MIBI</b>	sestamibi; <i>or</i> technetium-99m	<b>NADP</b>	nicotinamide adenine dinucleotide phosphate
<b>MIC</b>	minimum inhibitory concentration	<b>NADPH</b>	reduced form of nicotinamide adenine dinucleotide phosphate
<b>μOCT</b>	micro-optical coherence tomography	<b>NANIPER</b>	non-allergic non-infectious perennial rhinitis
<b>MIFC</b>	minimally invasive follicular carcinoma	<b>NAP4</b>	Fourth National Airway Project
<b>MIP</b>	minimally invasive parathyroidectomy; <i>or</i> maximum intensity projection; <i>or</i> macrophage inflammatory protein;	<b>NARES</b>	non-allergic rhinitis with eosinophilia syndrome
<b>miRNA</b>	micro ribonucleic acid	<b>NBCA</b>	n-butyl-2-cyanoacrylate; <i>or</i> N-butylcyanoacrylate
<b>MIST</b>	minimal invasive sinus techniques	<b>NBT</b>	nitro blue tetrazolium
<b>MIT</b>	monoiodotyrosine; <i>or</i> mono-iodothyronine; <i>or</i> minimally invasive thyroidectomy	<b>NCAS</b>	National Clinical Assessment Service (UK)
<b>MIVAT</b>	minimally invasive video-assisted thyroidectomy	<b>NCEPOD</b>	National Confidential Enquiry into Patient Outcome Death (UK)
<b>MLKL</b>	mixed lineage kinase domain-like protein	<b>NCIC</b>	National Cancer Institute of Canada
<b>MLTB</b>	microlaryngotracheobronchoscopy	<b>ncRNA</b>	non-coding ribonucleic acid
<b>MMC</b>	mitomycin C; <i>or</i> Modernising Medical Careers	<b>Nd-YAG</b>	neodymium-yttrium aluminium garnet
<b>MMP</b>	mucous membrane pemphigoid; <i>or</i> matrixmetalloprotease	<b>NET</b>	nerve excitability test; <i>or</i> neuroendocrine tumour

NF1	neurofibromatosis type 1	OPCS	Office for Population Censuses and Surveys (UK)
NF2	neurofibromatosis type 2	OPF	osteoplastic flap procedure
NH	normal hearing; <i>or</i> neurohypophysis	OR	occupational rhinitis
NHANES	National Health and Nutrition Examination Survey	OREP	olfactory event-related potential
NHL	non-Hodgkin's lymphoma	ORL	otorhinolaryngology
NHS	National Health Service (UK)	OSA	obstructive sleep apnoea
NHSLA	National Health Service Litigation Authority	OSATS	obstructive structured assessment of technical skill
NIBP	automatic non-invasive blood pressure	OTOF	otoferlin
NICE	National Institute for Health and Care Excellence (UK)	OXTR	oxytocin receptor
NIFTP	non-invasive follicular tumour with papillary-like nuclei	P	phosphate; <i>or</i> posterior
NIH	National Institutes of Health (USA)	PA	pernicious anaemia
NIM	nerve integrity monitor	PAC	P1 artificial chromosome; <i>or</i> pulmonary artery catheter; <i>or</i> parathyroid carcinoma
NIPF	nasal inspiratory peak flow	PACS	picture archiving and communication systems
NIS	Na <sup>+</sup> /I <sup>-</sup> symporter	PACU	post-anaesthesia care unit
NK	natural killer	PAF	platelet-activating factor
nLTP	non-specific lipid transfer protein	PBA	procedure-based assessment
NNT	number needed to treat	PBP	progressive bulbar palsy
NO	nitric oxide	PCA	patient-controlled analgesia
NO <sub>2</sub>	nitric dioxide	PCC	prothrombin complex concentrate; <i>or</i> Professional Conduct Committee (UK); <i>or</i> pheochromocytoma
NOAC	novel oral anticoagulant	PCD	primary ciliary dyskinesia
NOE	naso-orbito-ethmoid	PCOS	polycystic ovary syndrome
NOTSS	non-technical skills for surgeons	PCR	polymerase chain reaction
NOS	not otherwise specified	PCS	physical component summary
NP	nasopharynx; <i>or</i> nasopharyngeal	PD	Parkinson's disease
NPC	nasopharyngeal cancer; <i>or</i> nasopharyngeal carcinoma	PDCAT	poorly differentiated carcinoma of the thyroid
NPSA	National Patient Safety Agency (UK)	PDE	phosphodiesterase
NPT	near patient testing	PD1	programmed cell death protein 1
NPV	negative predictive value	PDGF	platelet-derived growth factor
NRTIs	nucleoside/nucleotide reverse transcriptase inhibitors	PDL	pulsed dye laser
NSAID	non-steroidal anti-inflammatory drug	PDR	Physicians' Desk Reference
NSF	national service framework; <i>or</i> nasoseptal flap	PDS	polydimethylsiloxane
NSHI	non-syndromic hearing impairment	PDT	photodynamic therapy
NTT	near-total thyroidectomy	PE	polyethylene; <i>or</i> pulmonary embolism; <i>or</i> pharyngo-oesophageal
O <sub>3</sub>	ozone	PEG	percutaneous endoscopic gastrostomy
OAE	otoacoustic emission	PER	persistent allergic rhinitis
OB	olfactory bulb	PET	polyethylene terephthalate; <i>or</i> positron emission tomography
OCT	optical coherence tomography	PET-CT	positron emission tomography/computed tomography
ODP	operating department practitioner	PET-MRI	positron emission tomography-MRI
OF	ossifying fibroma	PF	posterior fontanelle; <i>or</i> cisplatin/5-fluorouracil
OFDI	optical frequency domain imaging		
OIDA	observe; interpret; decide; act		
OMC	ostiomeatal complex		
OPC	oropharyngeal cancer; <i>or</i> oropharyngeal candidiasis		

PF4	platelet factor 4	PTP	post-transfusion purpura
PFAPA	periodic fever, aphthous stomatitis, pharyngitis and cervical adenitis	PTU	propylthiouracil
PFS	progression-free survival	PUOF1	pituitary-specific positive transcription factor 1
PG	paraganglioma	PVA	polyvinyl alcohol
PGA	polyglycolic acid	PVC	polyvinyl chloride
PGD2	prostaglandin D2	PW	posterior wall
PGE1	prostaglandin-E1	QALY	quality adjusted life year
PGI2	prostacycline; <i>or</i> prostaglandin I2	QOL	quality of life
PGL	persistent generalized lymphadenopathy	RA	retinoic acid
PHPT	primary hyperparathyroidism	RAE	Ring, Adair, Elwyn
PI	pulsatility index; <i>or</i> protease inhibitors	RAF	rapidly accelerated fibrosarcoma signal-regulated kinase
PI3K	phosphatidylinositol 3	RAI	radioactive iodine
PICU	paediatric intensive care unit	RANKL	regulation of nuclear factor $\kappa$ B ligand
PIII	parathyroid III	RANTES	regulated on activation, normal T-cell expressed and secreted
PIP	peak inspiratory pressure	RAS	recurrent aphthous stomatitis; <i>or</i> rat sarcoma protein family
PIV	parainfluenza virus; <i>or</i> parathyroid IV	RAST	radioallergosorbent test
PKA	protein kinase A	RAT	rapid antigen testing; <i>or</i> robotic-assisted thyroidectomy
PLAT	paraganglioma-like adenoma of the thyroid	RB	retinoblastoma
PLD	potentially lethal damage	RBC	red blood cell
PM	particulate matter	RCC	Rathke cleft cyst; <i>or</i> red cell count
PNH	paroxysmal nocturnal haemoglobinuria	RCoA	Royal College of Anaesthetists
PNIF	peak nasal inspiratory flow	RCT	randomized controlled trial
p.o.	by mouth	RET	rearranged during transfection
POGO	prescription of gain and output	RF	rheumatoid factor; <i>or</i> radio frequency
POMC	propiomelanocorticotrophin	RFLP	restriction fragment length polymorphism
PONV	postoperative nausea/vomiting	RFRP	RFamide-related peptide
PP	pyrophosphate	RFTVR	radiofrequency tissue volume reduction
PPD	purified protein derivative; <i>or</i> parathyroid proliferative disease	rFVIIa	recombinant factor VIIa
PPV	positive predictive value	rhBMP-7	recombinant human bone morphogenetic protein 7
PR3	proteinase 3	rhPTH	recombinant human parathyroid hormone
PRISMA	Preferred Reporting Items for Systematic Reviews and Meta-Analyses	rhTSH	recombinant human TSH
PRL	polypeptide hormone prolactin	RHD	Reported Hearing Disability
PRP	platelet-rich plasma	RIG	radiologically inserted gastrostomy
PRPP	5-phospho-alpha-D-ribose 1-diphosphate	RLN	recurrent laryngeal nerve
PSA	prostate-specific antigen; <i>or</i> pleomorphic salivary adenoma; <i>or</i> persistent stapedial artery	RLNP	recurrent laryngeal nerve palsy
PS-OCT	polarization-sensitive OCT	RNA	ribonucleic acid
PT	prothrombin time	RNP	ribonucleoprotein
PTAH	phosphotungstic acid haematoxylin	ROS	reactive oxygen species
PTC	papillary thyroid cancer; <i>or</i> psychophysical tuning curve	ROSE	rapid on-site evaluation
PTG	parathyroid gland	ROTEM	thromboelastography
PTH	parathyroid hormone	RP	rapid prototyping; <i>or</i> relapsing polychondritis
PTHrP	parathyroid hormone-related protein; <i>or</i> parathyroid hormone-related peptide	RR	relative risk
PTMC	papillary thyroid microcarcinoma		
pTNM	pathological tumour, nodes, metastases		

RRA	radioiodine remnant ablation	SLE	systemic lupus erythematosus
RRR	relative risk reduction	SLICC	Systemic Lupus International Collaborating Clinics
RSDI	Rhinosinusitis Disability Index	SLIT	sublingual immunotherapy; <i>or</i> sublingually in drops or tablets
RSOM	rhinosinusitis outcome measure	SLN	superior laryngeal nerve
RSTL	relaxed skin tension line	SMAS	superficial or subcutaneous musculoaponeurotic system
RSV	respiratory syncytial virus	SAMD	submucosal diathermy
RT	radiotherapy	SMR	submucosal resection
RTOG	Radiation Therapy Oncology Group	SMS	short message service; <i>or</i> indium-111 pentetretotide
rT3	reverse triiodothyronine	SNHL	sensorineural hearing loss
RTK	receptor tyrosine kinase	SNOT	sino-nasal outcome test
RUDS	reactive upper airways dysfunction syndrome	SNUC	sinonasal undifferentiated carcinoma
SA	solitary adenoma; <i>or</i> situational awareness	SO <sub>2</sub>	sulphur dioxide
SAD	supraglottic airway device	SOCS2	suppressors of cytokine signalling 2
SAGM	saline-adenine-glucose-mannitol	SOE	supraorbital ethmoid cells
SAPS	simplified acute physiology score	SOFA	sequential organ failure assessment
SBS	sick building syndrome	SOFT	solitary oxyphil follicular tumour
s.c.	subcutaneous	SOS	guanine nucleotide exchange factor (son of sevenless)
SCC	squamous cell carcinoma or cancer; <i>or</i> semicircular canal	SPA	sphenopalatine artery
SCCA	squamous cell carcinoma antigen	SPECT	single photon emission computed tomography
SCCHN	squamous cell carcinoma of the head and neck	SPET	single photon emission tomography
SCID	severe combined immunodeficiency	SPF	sphenopalatine foramen
SCIT	subcutaneous immunotherapy	SPLINTS	scrub practitioners list of intra-operative non-technical skills
SCL-90	Symptom Checklist-90	SPT	skin prick test; <i>or</i> station pull through
SCM	sternocleidomastoid muscle	SPTX	subtotal parathyroidectomy
SCN	severe congenital neutropenia; <i>or</i> solid cell nests	SRS	subacute rhinosinusitis
SEER	Surveillance, Epidemiology, and End Results (program)	SS	somatostatin
SERM	selective-oestrogen receptor modifier	ssDNA	single-stranded DNA
SETTLE	spindle cell tumour with thymus-like differentiation	SSC	superior semicircular canal; <i>or</i> Surviving Sepsis Campaign
SF-36	Medical Outcome Study Short-Form 36-Item Health Survey	SSEP	steady-state potential
SFF	speaking fundamental frequency; <i>or</i> solid free form fabrication	SSLP	simple sequence length polymorphism
SHC	SH2-containing protein	SSQ	speech, spatial and qualities
Shh	sonic hedgehog	SSTR1-5	somatostatin receptors 1-5
SHI	syndromic hearing impairment	SSRI	selective serotonin reuptake inhibitor
SHOT	serious hazards of transfusion	ST	superior turbinate; <i>or</i> subtotal thyroidectomy
SHPT	secondary hyperparathyroidism	STAT	signal transducer and activator of transcription
SIADH	syndrome of inappropriate antidiuretic hormone	STD	standard deviation
sIgE	specific immunoglobulin E	STRP	short tandem repeat polymorphism
SIGN	Scottish Intercollegiate Guidelines Network	SUV	standardized uptake value
SIP	sickness impact profile	SV	stroke volume
siRNA	small interfering ribonucleic acid	SVCO	superior vena caval obstruction
SLD	sublethal damage	SVS	selective venous sampling

SVCO	superior vena cava obstruction	TNM	tumour, node, metastasis
SVR	systemic vascular resistance	TNO	transnasal oesophagoscopy
Syk	signal-propagating kinase	TOE	transoesophageal echocardiography; <i>or Trichophyton, Oidiomyces and Epidermophyton</i>
T	thymine; <i>or</i> tumour	TOF	tracheo-oesophageal fistula
T1WI	T1-weighted images	TPO	thyroid peroxidase; <i>or</i> thyroperoxidase
T2WI	T2-weighted images	TPP	thyrotoxic periodic paralysis
T3	triiodothyronine	TPTX	total parathyroidectomy
T4	thyroxine	TPZ	tirapazamine
TAA	tumour associated antigens	TRALI	transfusion-related acute lung injury
TACO	transfusion-associated circulatory overload	TRAM	transverse rectus abdominis myocutaneous
TAD	transfusion-associated dyspnea	TRH	thyrotrophin-releasing hormone
TAGVHD	transfusion-associated graft-versus-host disease	tRNA	transfer ribonucleic acid
TAM	tumour associated macrophages	TSA	tumour specific antigens
TB	tuberculosis; <i>or Mycobacterium tuberculosis</i>	TSH	thyroid-stimulating hormone; <i>or</i> thyrotrophin
TBG	thyroxine-binding globulin	TSHoma	TSH-secreting adenoma
TcT	cytotoxic	TSH-R	TSH receptor
<sup>99m</sup> Tc	technetium-99m	TT	thrombin time; <i>or</i> total thyroidectomy
Tc-99m (v) DMSA	pentavalent dimercaptosuccinic acid	TTF-1	thyroid transcription factor-1
TC	thyroid cartilage	TTP	thrombotic thrombocytopeniac purpura
TCI	target-controlled infusion	TUNEL	TdT-mediated nick end labelling
TCP	tricalcium phosphate	U	uracil
TCR	T-cell receptor	UCT	unclassifiable complications of transfusion
t.d.s.	3 times a day	UICC	International Union Against Cancer
TEG	thromboelastography	UIC	urinary iodine concentration
TFT	thyroid function test	UKRETS	UK Registry of Endocrine and Thyroid Surgery
TG	thyroglobulin	UMP	uridine monophosphate
TgAb	thyroglobulin antibodies	UNICEF	United Nations Children's Fund
TGF	transforming growth factor	UP	uncinate process
TGF- $\alpha$	transforming growth factor alpha	UPSIT	University of Pennsylvania Smell Identification Test
TGF- $\beta$	transforming growth factor beta	URTI	upper respiratory tract infection
TGF- $\beta$ 1	transforming growth factor beta 1	US	ultrasound; <i>or</i> ultrasonography
Th	T-helper	USH	Usher syndrome
Th1	T-helper 1 cell	USH1B	Usher syndrome type 1B
Th2	T-helper 2 cell	USS	ultrasound scan
THST	thyroid hormone suppression therapy	UV	ultraviolet
THW	thyroid hormone withdrawal	V2R	vasopressin type 2 receptor
TI	thyroid isthmusectomy	VA	Veterans' Affairs; <i>or</i> vestibular aqueduct
TIL	tumour infiltrating lymphocytes	VAC	vacuum-assisted closure
TIVA	total intravenous anaesthesia	VAS	visual analogue scale; <i>or</i> visual analogue score
TKI	tyrosine kinase inhibitor	VATS	video-assisted thoracoscopic surgery
TL	total lobectomy	VC	vocal cord
TME	tumour microenvironment		
TMC1	transmembrane channel-like gene 1		
TMJ	temporomandibular joint		
TNF	tumour necrosis factor		
TNF- $\alpha$	tumour necrosis factor alpha		

<b>VCAM-1</b>	vascular cell adhesion molecule-1	<b>WDTC</b>	well-differentiated thyroid cancer
<b>vCJD</b>	variant Creutzfeldt—Jakob disease	<b>WDT-UMP</b>	well-differentiated tumour of uncertain malignant potential
<b>VCP</b>	vocal cord paralysis	<b>WHAFFT</b>	worrisome histologic alteration following fine-needle aspiration of the thyroid gland
<b>VDA</b>	vascular disrupting agent	<b>WHO</b>	World Health Organization
<b>VEGF</b>	vascular endothelial growth factor	<b>WIFC</b>	widely invasive follicular carcinoma
<b>VEGFR</b>	vascular endothelial growth factor receptor	<b>WMD</b>	weighted mean difference
<b>VF</b>	vocal fold	<b>WP</b>	Woodruff's plexus
<b>VHI</b>	Voice Handicap Index	<b>WPBA</b>	workplace based assessments
<b>VHI-10</b>	Voice Handicap Index-10	<b>WS</b>	Waardenburg syndrome
<b>VHQ</b>	Vertigo Handicap Questionnaire	<b>XLA</b>	X-linked agammaglobulinaemia
<b>VILI</b>	ventilator induced lung injury	<b>XM</b>	crossmatch
<b>VN</b>	vestibular nuclei; <i>or</i> vagus nerve	<b>YAC</b>	yeast artificial chromosome
<b>VNO</b>	vomero nasal organ	<b>YAG</b>	yttrium aluminium garnate
<b>VOC</b>	volatile organic compound		
<b>VTE</b>	venous thromboembolism		
<b>VTFF</b>	vertex-to-floor		
<b>vWD</b>	von Willebrand disease		
<b>vWF</b>	von Willebrand factor		
<b>VZV</b>	varicella zoster virus		
<b>WBS</b>	whole-body scan		
<b>WDC-NOS</b>	well-differentiated carcinoma, not otherwise specified		



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## MOLECULAR BIOLOGY

Michael Kuo, Richard M. Irving and Eric K. Parkinson

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## SEARCH STRATEGY

Data in this chapter may be updated by a Medline search using the keywords: molecular biology, genetics and cell biology.

## INTRODUCTION

Molecular biology describes the study of the biochemical processes that govern the behaviour of cells. These processes form the fundamental mechanisms by which cell function, cell–cell interactions and cell turnover are regulated. Disruption of this regulation may lead to disease, whilst an understanding of these mechanisms allows the physician to attempt to predict disease behaviour and to explore methods of restoring this regulation at a molecular level. This chapter reviews the principles of molecular genetics and outlines aspects of the molecular biology of the cell in the context of otolaryngological disease processes and describes some of the techniques that form the backbone of current molecular biology. It should give the reader sufficient background knowledge of molecular biology to understand subsequent chapters discussing the molecular biology of specific otolaryngological conditions.

## MOLECULAR GENETICS: DNA STRUCTURE AND FUNCTION

Hereditary information in eukaryotes is stored in the form of double-stranded deoxyribonucleic acid (DNA) and is referred to as the genome. DNA forms a double-helix structure as a result of hydrogen bonds between complementary pairs of nucleotides, adenine (A) with thymine (T) and cytosine (C) with guanine (G). The nucleotides on each strand are organized linearly in triplets, known as codons. Each specific sequence determines

a single specific amino acid, for example ACU specifies threonine. However, as there are more triplet combinations (64) than commonly encountered amino acids (20), some proteins may be represented by different codons (e.g. lysine by AAA as well as AAG) and some codons (UAA, UGA and UAG) are ‘stop’ codons, constituting a signal for arrest of translation. The overwhelming majority of this DNA (99.9%) exists in the cell nucleus as the nuclear genome, which, in the human, is estimated to be 3000 megabase pairs in physical size and encodes 30 000–35 000 genes. The remaining DNA (16.6 kilobase pairs) forms the mitochondrial genome, encoding 37 genes. The mitochondrial genome and its potential role in cancer diagnostics will be discussed later.

Each DNA molecule is packaged into a chromosome by complex folding of the DNA around proteins. Diploid human cells contain 22 pairs of autosomes (1 to 22) and a pair of sex chromosomes (XX or XY) that determines the sex of the organism. One of each pair of chromosomes is maternally inherited and the other is paternally inherited. Each chromosome has a distinctive shape, size and banding pattern, but have the common appearance of two arms apparently separated by a constriction. The centromere is microscopically recognizable as the central constriction separating the chromosome into a long arm (q for queue) and a short arm (p for petit), but its biological role lies in anchoring the chromosome to the mitotic spindle for segregation during cell division. The ends of the chromosomes are capped by telomeres, which are specialized structures containing unique simple repetitive sequences. They maintain the structural integrity of the

chromosome and provide a solution for complete replication of the extreme ends of the chromosome. The conventional nomenclature for chromosomal locus assignment is given by the chromosome number, followed by the arm and finally the position on the arm, for example, 3p21 indicates position 21(two-one) on the short arm of chromosome three.

During normal cell division, DNA replication is achieved by the separation of the two strands by DNA helicase. Each separated single strand then acts as a template for polymerization, catalyzed by DNA polymerase, of nucleotides forming a new complementary strand and thus double-stranded DNA identical to the original dsDNA. As each daughter DNA consists of one original and one newly synthesized DNA strand, the process is known as semi-conservative replication. The specificity of the complementary relationship between the nucleotides on each strand forms the basis for many techniques of modern molecular biology and molecular cytogenetics.<sup>1</sup> The accuracy with which DNA replication takes place is remarkable with an estimated error rate of less than one in  $10^9$  nucleotide additions. Such accuracy is of vital importance to the individual as a permanent change in DNA, or mutation may cause inactivation of a gene essential to cell survival or cell cycle control. The high fidelity of DNA sequence replication is achieved by unidirectional 5'-to-3' direction of DNA replication, a rigorous DNA proofreading mechanism that detects mismatched DNA and efficient DNA repair pathways that excise and repair DNA damage. Failure of these mechanisms, such as is encountered in xeroderma pigmentosum, Fanconi's anaemia and ataxia telangiectasia, leads to accumulation of DNA replication errors and a high incidence of malignancies.

Although the human nuclear genome is  $3 \times 10^9$  base pairs in size, about 90% of it is non-coding, with all the genes being coded by the remaining 10% of the DNA. Within the non-coding DNA are dispersed short arrays of repeat units of pairs or triplets of nucleotides (di-/trinucleotides). The exact function of these microsatellite repeats is not entirely clear, but their existence and frequency of dispersion throughout the genome have greatly facilitated study of the genetics of tumours and many inherited disorders, which will be discussed later.

A gene is a region of the chromosomal DNA that produces a functional ribonucleic acid molecule (RNA). It comprises regulatory DNA sequences that determine when and in which cell types that gene is expressed, exons that are coding sequences and interspersed introns that are non-coding DNA sequences. These regulatory sequences often consist of CpG islands, short stretches of DNA rich in dinucleotides of cytosine and guanine. The methylation status of these CpG islands determines whether that gene is expressed in a particular cell or tissue, being unmethylated in tissues where the genes are expressed. As will be discussed later, aberration of this control is one of the mechanisms of tumour suppressor gene inactivation. However, these same genes can also be regulated by proteins that recognize methylated sequences called histones<sup>2</sup> and these in turn can be regulated by polycomb genes

such as *BM11*.<sup>3</sup> Transcription is the intra-nuclear process driven by RNA polymerase whereby one of the two DNA strands acts as a template for the synthesis of a single RNA strand which is complementary to the DNA, except that uracil replaces thymine in RNA. This primary RNA transcript then undergoes post-transcriptional processing, or splicing.<sup>4</sup> Traditional dogma held that one gene produces one protein and therefore splicing was considered to occur simply in order to remove the non-coding intronic sequences, producing messenger RNA (mRNA). It is now known that by 'alternative splicing', one gene can result in the production of several different but often related proteins in different tissues.<sup>5</sup>

The mature mRNA then migrates into the cytoplasm where it acts as a template for the synthesis of a polypeptide during translation, a process regulated and catalyzed by cytoplasmic ribosomes. Successive amino acids are added to the polypeptide chain according to the triplet code on the mRNA, which is recognized by the transfer RNA (tRNA), to which each corresponding amino acid is covalently bound. Translation is commenced upon recognition of an initiation codon (usually but not exclusively AUG/methionine) and terminated upon recognition of a stop codon. The polypeptide subsequently undergoes a variable degree of post-translational modification and/or cleavage to produce the mature protein product, which may have an intra-cellular role or may be exported to the endoplasmic reticulum and hence to the extracellular space to execute its function.

The mitochondrial genome is considerably smaller than the nuclear genome, but it deserves mention here because of the increasing recognition of the role of mitochondrial DNA (mtDNA) mutations in human disease. The mitochondrial genome is only 16.6kb in size, comprising 37 genes, which encode polypeptides that are principally involved in the respiratory chain. mtDNA is double-stranded but does not form a double-helix or chromosomes, but instead takes the form of a circular double-stranded DNA structure with a heavy and a light strand. Unlike the nuclear genome, which is inherited from mother and father, the mitochondrial genome of an individual is entirely maternally inherited.

## OTHER REGULATORS OF GENE EXPRESSION

In the last few years it has become apparent that gene expression is 'fine-tuned' by several classes of molecule. Small interfering RNAs (siRNAs) were first discovered in plants and are now widely used as research tools to suppress gene expression. However, natural versions of siRNAs exist (microRNAs- miRNAs) and are thought to be biologically significant regulators of gene expression.<sup>6</sup> miRNAs are small non-coding RNA molecules that function through base-pairing with mRNA, thus preventing their translation into proteins. There may be more than 1000 miRNAs in the human genome<sup>7</sup> and some of them are associated with oral cancer; they are also secreted into

body fluids making them candidates for the non-invasive detection of the disease.<sup>8</sup> In addition, long non-coding RNAs (ncRNAs) have been identified that are thought to regulate various aspects of gene expression, including transcription, splicing, translation, siRNA-directed gene regulation and epigenetic regulation.<sup>9</sup> Whilst the study of ncRNAs is still new they have been implicated in a number of diseases, including ageing and cancer.

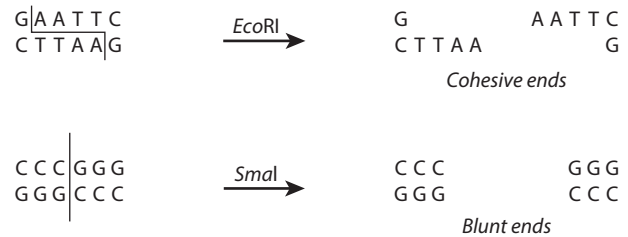
### KEY POINTS

- The double-stranded alpha helical structure of DNA, mainly located in the nucleus, consists of nucleotide triplets called codons which code for specific amino acids and stop signals, and forms the substrate for hereditary information in eukaryotes.
- The 22 pairs of autosomes and one pair of sex chromosomes, each with their distinctive shape, size and banding pattern, represent a complex folding of DNA around proteins to give the characteristic shape of a central constriction (centromere) separating the chromosome into a long arm (q) and a short arm (p) with a telomere cap at each end to maintain structural integrity.
- Chromosome locus nomenclature: chromosome number – 3p21 – position on chromosome arm.
- Semiconservative replication of DNA during normal cell division results in the separation of two strands of DNA by DNA helicase, each strand then acting as a template for polymerization by DNA polymerase. High fidelity is vital to prevent permanent change or mutations.
- A gene is a region of chromosomal DNA which produces functional RNA consisting of:
  - regulatory DNA sequences
  - exons, which are coding sequences
  - introns, which are non-coding sequences.
- Transcription is the intra-nuclear process driven by RNA polymerase whereby one of the two DNA strands acts as a template for single-stranded RNA synthesis complementary to the DNA, except that in RNA T is replaced by U. Splicing refers to post-transcriptional processing of RNA.
- Translation is the cytoplasmic process in which mRNA acts as a template for the synthesis of polypeptide by adding successive amino acids to the polypeptide chain, according to the triplet codon of the mRNA that is recognized by the tRNA to which the corresponding amino acid is covalently bonded. This process is regulated and catalyzed by cytoplasmic ribosomes. Post-translational modification produces mature proteins.
- Gene expression can be modulated by naturally-occurring miRNAs and ncRNAs.

## METHODS IN MOLECULAR BIOLOGY

### Basic techniques of DNA fragmentation and identification

Unlike RNA, DNA is extremely stable, which is understandable from the function that each has in the cell. For purposes of studying the DNA and in order to clone specific DNA, the DNA molecule needs to be divided into manageable fragments. Although the ability to cut (and also to join up) DNA molecules now appears to be a



**Figure 1.1 DNA cleavage by restriction endonucleases.** Derived from Ref. 10, with permission.

very straightforward process, it was only 1970 when the first restriction endonuclease was identified in a strain of *Haemophilus influenzae*, hence its name *HindII* (pronounced Hin-dee-two). It is believed that this restriction endonuclease acts *in vivo* in bacteria as an immune or host-defence system, recognizing non-self DNA in bacteriophages and cleaving them. By surveying many different bacteria, a wide range of restriction endonucleases is now available, each of which recognizes specific target sites based on sequences of four to eight nucleotides. As a specific, a seven nucleotide sequence (heptanucleotide) will occur less frequently than a four nucleotide sequence (tetranucleotide), statistically, endonucleases recognizing heptanucleotide targets will cut less frequently thereby yielding larger fragments than those recognizing tetranucleotides. As the DNA is double-stranded, the resultant fragments may have blunt ends or cohesive ('sticky') ends (Figure 1.1). The nature of the ends of DNA fragments thus generated impact upon the way in which they can be ligated (joined) into recombinant molecules. Ligation of DNA fragments with cohesive ends is more efficient than joining of blunt-ended fragments.

### Electrophoresis

Negatively charged phosphate groups on the DNA backbone confer a net negative charge on linear DNA. This allows fragments of different sizes to be resolved within a suitable gel matrix by the application of an electric current across the matrix. The DNA will migrate towards the positive electrode with the smaller fragments travelling faster than the larger fragments.<sup>10</sup> The size of the fragment can be estimated by the use of a graduated DNA ladder containing fragments of known molecular weight. The choice of the particular matrix depends on the fragment sizes that one is trying to resolve. Polyacrylamide gels can resolve differences of just one base pair between fragments of several hundred base pairs in size by virtue of a small pore size in the gel matrix. These gels can be used for DNA sequencing and resolution of alleles varying in only one dinucleotide repeat. Agarose gels can resolve fragment sizes from around 100 bp to 20 kb. Beyond that size, electrophoretic mobility is no longer proportional to fragment size. Resolution of fragments sizes in excess of 50 kb, such as larger bacterial artificial chromosomes (BAC) or yeast artificial chromosomes (YAC) require the use of pulsed field electrophoresis.

## Hybridization

Hybridization is the specific annealing of single DNA (or RNA) strands, the probe, to a DNA sample, the target. It serves to detect the presence of a specific sequence of DNA either in the cell or on a hybridization membrane and recognition that hybridization has occurred is achieved either by radioactively labelling the probe and localizing the radioactivity by autoradiography or by labelling the probe with fluorochromes that fluoresce when excited by light of specific wavelengths (Figure 1.2). Hybridization on a membrane requires the initial transfer of DNA on to a nitrocellulose membrane from an agarose gel. This elegantly simple process is eponymously known as Southern blotting after the scientist who described the process in 1975. Two other commonly used transfer techniques have their names derived from Southern blotting as jargon terms. Northern blotting is essentially the same process used for transfer of RNA to a membrane. Western blotting is one of the mainstays of protein analysis and involves the transfer of electrophoresed protein bands from a polyacrylamide gel on to a nitrocellulose or nylon membrane to which they bind strongly. Detection of the protein is usually achieved by the use of antibodies to specific antigens presented by the protein with the antibody being labelled radioactively, enzymatically or fluorescently.

## Cytogenetics and molecular cytogenetics

Although microscopy had already reached high levels of resolution in the early 1930s, the correct number of human chromosomes was not determined until 1958. The era of classical cytogenetics had thus begun. Cytogenetics is the study of chromosomal abnormalities and rearrangements. It currently has a major role to play in pre-natal diagnosis of Down syndrome and other congenital syndromes characterized by numerical chromosomal abnormalities. In the early part of this century, Theodore Boveri proposed that cancer arose from chromosomal alterations. This hypothesis was not proven until the consistent chromosomal translocation, t(9;22), was demonstrated in chronic myeloid leukaemia. Since that time, cytogenetic analysis has been the mainstay of genetic analysis in reticuloendothelial malignancies, being responsible for the identification of

consistent translocations in different leukaemias. Its use in solid tumours has been hampered by the difficulties of establishing short-term primary cultures from head and neck cancers for chromosomal analysis and the erratically acquired chromosomal changes in long-term cell lines, which may have occurred *in vitro*, influenced by culture conditions. Nevertheless, some studies have identified chromosomal areas consistently showing frequent breakpoints suggesting the location of putative tumour suppressor genes (including 3p21, 5p14, 8p11, 17p21, 18q21) and gain or amplification implying the presence of putative protooncogenes at other sites (including 3q, 5p, 8q, 11q13). Although the refinement of karyotyping has been radically enhanced by the introduction of 24-colour combinatorial multicolor FISH (M-FISH), the resolution and therefore utility of solid tumour karyotyping remains limited.<sup>11</sup>

Hybridization to target DNA in cells, using fluorescence detection, is known as fluorescence *in situ* hybridization (FISH). Fluorescence *in situ* hybridization allows the analysis of copy number of a known specific DNA sequence within intact nuclei. In reticuloendothelial malignancies and solid tumour-derived cell lines, the use of both single-copy probes and centromere alpha-satellite repeat probes on metaphase preparations has enhanced and refined classical karyotyping. Interphase FISH has been applied to solid tumour sections to assess the copy number of a known sequence in breast, prostate, bladder, brain, lung and head and neck tumours.

Fluorescence-labelled hybridization has also been combined with cytogenetics to produce the powerful technique of comparative genomic hybridization (CGH).<sup>12</sup> Comparative genomic hybridization permits the rapid medium resolution screening of the entire genome by comparatively hybridizing matched tumour and normal DNA from a patient, which are labelled with different fluorochromes, on to normal metaphase chromosome preparations. Under red-green dual filter fluorescence microscopy and computer-aided image analysis, areas of genetic 'neutrality' appear yellow, under-representation appears green, and over-representation appears red. Areas of genetic under-representation suggest the possibility of a tumour suppressor gene lying within that region while areas of over-representation may indicate the location of a putative oncogene. This technique has been applied to the rapid genetic analysis of many tumour types including squamous cell carcinomas of the head and neck. The advent of molecular cytogenetics has obviated the need for primary short-term cultures and refined the location of chromosomal aberrations in solid tumours.

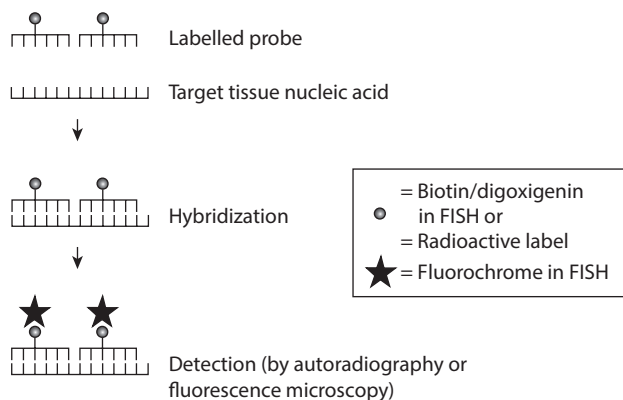


Figure 1.2 *In situ* hybridization.

## Polymerase chain reaction

Perhaps the single molecular technique that has had the most dramatic impact on molecular biology has been the polymerase chain reaction (PCR). The original problem lay in obtaining sufficient quantities of a particular DNA sequence such that DNA profiling (e.g. sequencing) and DNA manipulation (e.g. cloning) could be achieved. The only 'requirement' is that the sequences flanking the stretch of DNA of interest is known. With that proviso,

PCR achieves faithful and exponential amplification of a specific sequence of DNA by repeated cycles each consisting of dsDNA denaturation, hybridization of specific oligonucleotides (primers) and extension of the polynucleotide by rapidly altering the reaction temperature between segments of each cycle. dsDNA denaturation is achieved by raising the temperature of the reaction to 94°C for 30 seconds, thus disrupting the hydrogen bonds between the strands and exposing the hydrogen bond donor and acceptor groups to allow base pairing. The oligonucleotide primers are then allowed to hybridize to the denatured DNA (annealing) at around 55–65°C for 90 seconds before the reaction temperature is raised to 72°C to permit extension of the DNA strand by DNA polymerase in the presence of deoxynucleoside triphosphates (dNTPs). With each cycle resulting in the doubling of the copies of the DNA sequence, a 30-cycle PCR taking approximately two hours would amplify a single copy of a DNA sequence 268 million-fold (Figure 1.3). Although the PCR was originally described by Mullis and Faloona in 1987, one practical problem prevented its instant exploitation.<sup>13</sup> The DNA polymerase used in the original reaction was denatured during the DNA denaturation segment and therefore had to be added after each and every cycle. The solution came in 1989 when Lawyer isolated and characterized the DNA polymerase, Taq polymerase, from the thermophilic bacterium *Thermus aquaticus* which normally resided in temperatures above 95°C.<sup>14</sup> This polymerase was therefore ‘heat resistant’ and did not need to be replenished between cycles.

The PCR holds a central position in many molecular biological techniques as well as clinical diagnostic methods. The fundamental principle of DNA amplification has been adapted to amplify messenger RNA and to amplify areas where the initial flanking oligonucleotide sequences are not known. It is often described as a sensitive and powerful technique, but with great power comes the potential for corruption! In theory, a single copy of DNA can be amplified. Therefore, careless experimental technique may lead to contamination of the DNA sample with other DNA (e.g. from the skin of the investigator)

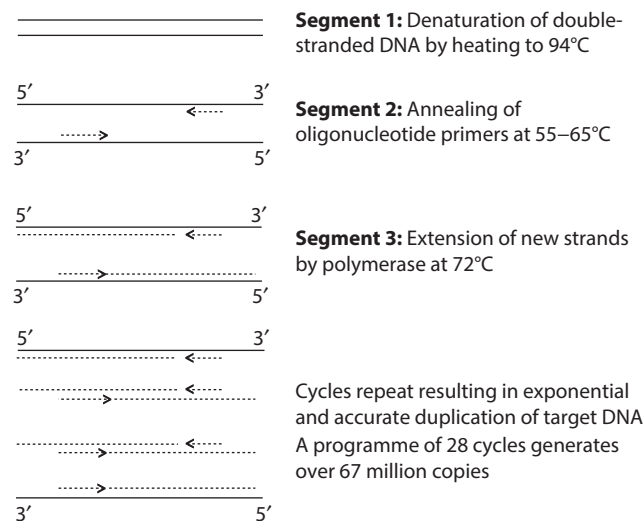


Figure 1.3 The polymerase chain reaction.

and consequently to an artefactual result. The Taq polymerase originally described in the technique does not have proofreading properties, but newer cloned enzymes such as Pfu polymerase incorporates a proofreading function to increase amplification fidelity for sequencing reactions.

The sensitivity of PCR also presented a problem for the analysis of genetic alterations in certain solid tumours. Squamous cell carcinomas of the head and neck are histologically often characterized by a large stromal element within the tumour. The genetic alterations in the tumour may not be present in the stromal tissue and thus total DNA extracted from the tumour will contain DNA from both benign and malignant tissue. This *in situ* contamination can now be eliminated by the use of laser capture microdissection (LCM) of tumours. LCM involves the placement of a laser-activated film over a tissue specimen. When areas of ‘pure’ tumour cells are identified, a focal laser pulse lifts the tissue on to the film in specimens down to 30 µm in diameter.<sup>15</sup>

### KEY POINTS

- Restriction endonucleases are enzymes that were initially identified in bacteria that can cut and join up DNA. They recognize specific target sites based on sequences of four and eight nucleotides.
- Electrophoresis is a technique for resolving the size of DNA fragments, which carry a negative charge from the phosphate groups on their backbone. Using a gel matrix with an electric current applied across it, the DNA will migrate to the positive electrode at a rate inversely proportional to its size.
- Hybridization is the specific annealing of single DNA or RNA strands (probe) to a DNA sample (target) to detect the presence of a specific sequence of DNA in the cell or hybridization membrane. Variants include the eponomously named Southern, Northern and Western blotting techniques.
- Cytogenetics is the study of chromosomal abnormalities and rearrangements important in the diagnosis of congenital syndromes characterized by numerical chromosomal abnormalities (e.g. Down syndrome and leukaemia types).
- FISH refers to fluorescence *in situ* hybridization which involves hybridization to target DNA cells using fluorescence detection and allows the analysis of copy number of a known specific DNA sequence within intact nuclei.
- PCR achieves faithful and exponential amplification of a specific sequence of DNA by repeated cycles each consisting of:
  - DNA denaturation by heating to 94°C to denature hydrogen bonds between strands
  - annealing (hybridization) of oligonucleotide primers to denatured DNA at 55–65°C
  - extension of DNA strand by DNA polymerase.

## MOLECULAR ABERRATIONS OF CELLULAR BIOLOGY

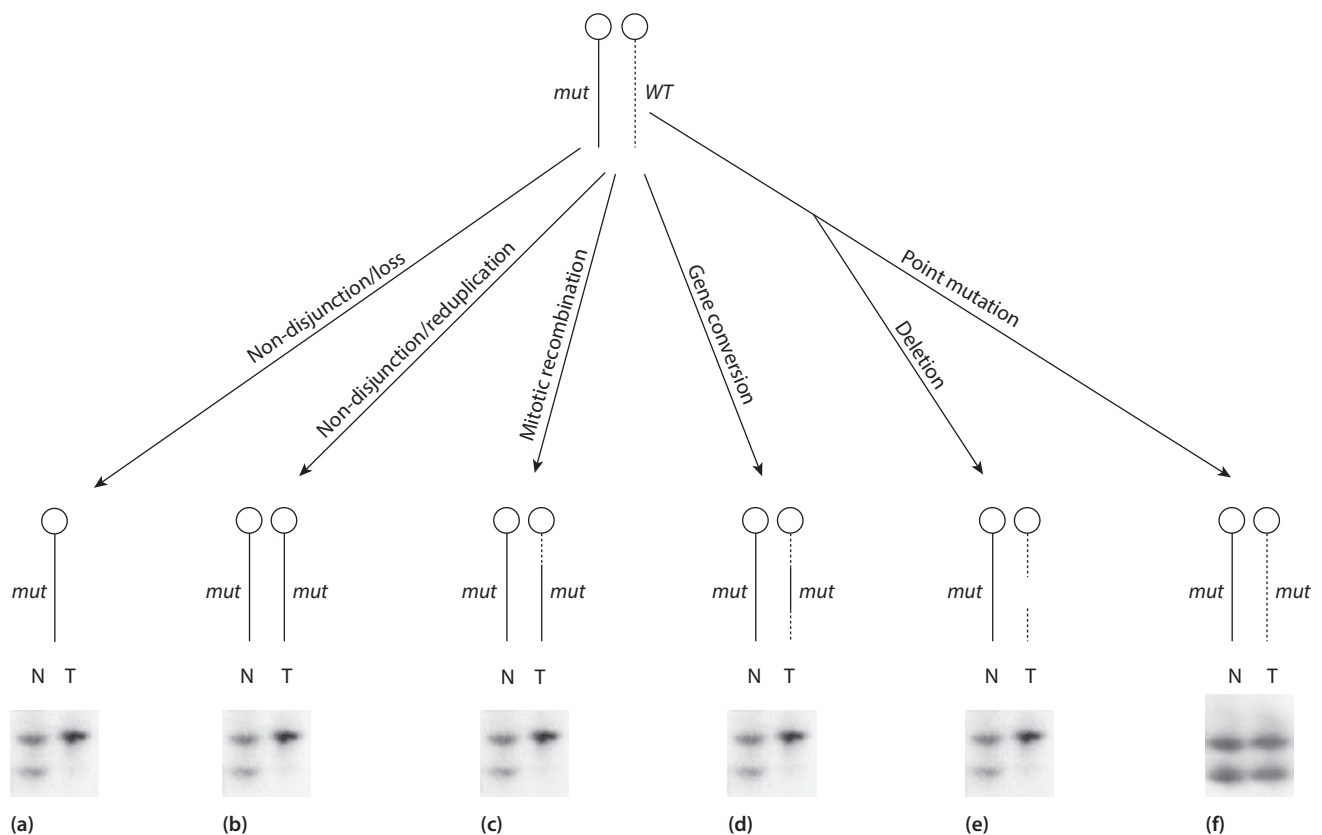
### Loss of heterozygosity and the expression of recessive mutant alleles

Retinoblastoma is a childhood cancer, which exhibits both hereditary and sporadic occurrence, with the inherited form transmitted as a highly penetrant autosomal

dominant trait. The proposition by Alfred Knudson in 1971, based upon a statistical analysis of the occurrence of retinoblastoma in children, that two genetic events were required to inactivate the gene mitigating against development of the cancer, was a major landmark in the understanding of tumour suppressor genetics.<sup>16</sup> In hereditary retinoblastomas, a single additional somatic event in a cell that carried the inherited mutation was sufficient to give rise to the disease while two somatic events were required to produce a sporadic retinoblastoma. This became known as Knudson's 'two-hit' hypothesis. The subsequent study on matched tumour and blood DNA from patients with sporadic retinoblastoma by Webster Cavenee not only proved Knudson's hypothesis but also established the paradigm for all subsequent investigations of tumour suppressor genes.<sup>17</sup> For the first time, the now widely accepted mechanisms of tumourigenesis were reconciled, viz. that neoplasms can arise in a multistep manner, that chromosomal events can lead to tumour formation and that chromosome loss with or without reduplication can lead to expression of recessive mutations. Perhaps even more strikingly, the authors presciently suggested that development of homozygosity for recessive mutant alleles at the *Rb-1* locus may give rise to the development of other tumours and that other additional dominant mutations may be involved in the development of retinoblastoma. Cavenee proposed the various chromosomal mechanisms that could

reveal recessive mutations and these are summarized for a putative tumour suppressor gene in **Figure 1.4**, adapted from the figure in his original paper. To these can now be added hypermethylation of the 5' CpG island resulting in transcriptional inactivation of the gene, discussed below.<sup>18</sup> The simplest way of revealing a recessive mutant allele is by deletion of the wild-type allele, resulting in hemizyosity at the particular locus on the remaining chromosome. It is inferred from this that areas of frequent allelic loss in tumours may represent the location of putative tumour suppressor genes and this hypothesis underpins the commonly employed method of molecular detection of allelic losses, loss of heterozygosity (LOH).

The practical exploitation of the concepts outlined above hinges on the presence of the previously described microsatellites, highly polymorphic non-coding DNA sequences, also referred to as simple sequence length polymorphisms (SSLP) or short tandem repeat polymorphisms (STRP), which are distributed approximately every 100 000 bp throughout the human genome. These microsatellites contain small dinucleotide or trinucleotide repeat units, the number of which may differ between the two alleles in a particular person. Microsatellite markers are now available which map thousands of these sequences to chromosomal loci. When DNA sequences containing these microsatellite markers are amplified by PCR in a person heterozygous for that



**Figure 1.4 Chromosomal mechanisms that could reveal recessive mutations.** In this example, before cell division, the tissue concerned carries a mutation in one copy of the hypothetical tumour suppressor gene. In each of the scenarios (a–f), the recessive mutation is revealed. If the individual is heterozygous for a microsatellite marker within or very close to the mutated gene, the hypothetical PCR results are given below each ideotype. The only mechanism that escapes observed loss of heterozygosity is F. *mut*, mutated; N, normal; T, tumour; *WT*, wildtype. After Ref. 17, with permission.

particular microsatellite, the PCR will yield two products of different lengths, which can be resolved on an electrophoretic gel. Where amplification of tumour DNA from such a subject yields only one product, the tumour is said to show LOH, implying allelic loss. Persons who are homozygous for a particular marker are said to be non-informative for that marker. The concept of examining the variation and extent of allelic deletion in tumours was introduced by Vogelstein in an analysis of colorectal carcinomas and termed allelotyping.<sup>19</sup> Allelotypes generated in this fashion have identified several areas of frequent allelic deletion from which some of the responsible tumour suppressor genes have been cloned or identified. The most common areas of loss in HNSCC are at chromosome 9p21, 17p13, 13q14, 4p, 5q21 and several discrete regions on 3p and 8p.<sup>20, 21</sup>

## Inactivation of genes and oncogenic transformation

Allelic deletion is only one mechanism by which a copy of a gene can be inactivated. As there are two copies of each gene, inactivation of the gene requires inactivation of both copies of the gene, ‘the second hit’. This may occur as a result of a genetic mutation or transcriptional silencing. Conversely, a protooncogene may be converted into an oncogene by a simple increase in the copy number of the gene (gene amplification) resulting in an overproduction of protein or by point mutations that affect the control of protein activity.

Not all mutations result in alteration in function of a gene. DNA mutation may occur as a result of base substitutions, as well as nucleotide insertions and deletions. Insertions and deletions of nucleotides are very rare in coding DNA, but if they occur they may produce a shift in the ‘reading frame’ that dramatically alters the coding downstream from the mutation. Base substitution is a more common form of mutation in coding DNA, which may have a range of consequences on the function of the gene; a loss of function (e.g. *p53*, *Rb*), a gain in function, often due to stabilization of the active protein (e.g. *c-erbB2*, *K-ras*) or no net functional effect. Since an amino acid may be encoded by different codons with a ‘third base wobble’ (e.g. GUA, GAC, GUG and GUU all encode valine), a substitution of the third base would result in no change to the amino acid. This is known as a silent substitution. At the other extreme is the nonsense mutation, whereby base substitution results in a stop codon which leads to truncation of the polypeptide and a dramatic reduction in function.

Loss of function in the presence of a normal (wildtype) allele may occur as a result of transcription silencing. The methylation status of CpG islands surrounding the promoter regions of genes determines whether a particular gene is expressed within a cell such that methylation of a CpG island ‘switches off’ the gene that it is regulating. CpG islands regulating housekeeping genes (i.e. genes encoding proteins essential for cell survival) are unmethylated in all cell types whereas those regulating cell-specific

genes (e.g. muscle-specific actin gene in muscle cells), are only unmethylated in the relevant cell. Many tumour suppressor genes (e.g. *CDKN2A* (p16<sup>INK4A</sup>), *APC*, *RASSF1A*) are now recognized to be inactivated by hypermethylation of the promoter region in malignant tumours.

## Gene sequencing

Recently a variety of next generation sequencing methods have been applied to the field. One method is to construct a library of DNA fragments and to use biotinylated RNA baits to capture the exomic sequences using streptavidin coated magnetic beads prior to loading them onto the sequencing machine following digestion of the RNA bait and a few cycles of PCR. This strategy was able to obtain 10 or more sequencing reads for over 90% of the targeted bases and this approach and similar strategies have revealed the mutational landscape of head and neck squamous cell carcinoma, including the frequent mutation of the *NOTCH1* gene, which had not been identified previously.<sup>22–24</sup>

## Proteomics and metabolomics

As the Human Genome Mapping Project powers its way towards a high-resolution sequence of the entire genome, the natural progression of research has been towards the elucidation of the repertoire of proteins that control cell signalling and cell growth. Proteomics is the profiling of proteins in cells and serum by two-dimensional gel analysis separating proteins by charge and mass, by X-ray crystallography and by mass spectrometry.<sup>25</sup> Proteomics has the ability to identify post-translational modifications, some of which are cancer cell-specific, which would not be detected by genetic or expression profiling. The power of this approach is particularly evident in cancer studies as it has the ability to compare the entire protein pattern of tumour tissue and normal tissue in a manner analogous to comparative genomic hybridization.

However, the functional consequences of changes to the proteome, especially some enzymes, can only be inferred and a more direct approach is to measure the levels of small molecules, or metabolites. Metabolomics is the global analysis of metabolites but is a relatively new discipline and no single technique is suitable for the analysis of all different types of molecule. Therefore, a mixture of techniques such as gas chromatography, high-pressure liquid chromatography and capillary electrophoresis are used to separate metabolites and the molecules are then identified using methods such as mass spectrometry. Metabolomics has already been applied to the detection of oral cancer in saliva<sup>26</sup> and serum.<sup>27</sup>

## Telomerase

Aberration of telomere biology has a now well-recognized role to play in cancer development. The length of the telomere is maintained in germline cells by the enzyme telomerase that is absent in normal somatic cells.<sup>28</sup> This observation led to the theory that progressive shortening



of telomeres with each cell division leads to cell senescence and ageing of the organism. Several mechanisms for this have been proposed. A critical shortening of the telomere may leave a non-telomere-free DNA end that signals cell cycle arrest. Telomere loss could also extend to the deletion or inactivation of genes located subtelomericly, again leading to cell cycle arrest. Two important consequences of critical telomere shortening in cells that survive the cell crisis are genomic rearrangements and chromosomal loss, which may in turn lead to malignant transformation. This is particularly relevant to rapidly dividing and regenerating tissues, such as skin and mucosa. Paradoxically, while some of these survivors will subsequently go into cell cycle arrest due to critical telomere length shortening, a proportion will then express telomerase activity. Such activity is capable of cell immortalization by maintaining telomere length indefinitely and its role in cancer is now unquestionable. Telomerase activity has been demonstrated in 80% of oral cancers and around 50% of oral leukoplakia lesions.<sup>29</sup> Even among premalignant lesions, telomerase activity was associated with degree of dysplasia, suggesting that telomerase activation occurs during the late stage of oral premalignancy. The potential for anti-telomerase drugs as a novel treatment remains promising, especially as they are now thought to target cancer stem cells.<sup>30</sup>

### KEY POINTS

- Practical techniques based on loss of heterozygosity of tumour DNA are based on Knudson's two-hit hypothesis, which states that two genetic events are required to inactivate the gene that prevents the development of a condition.
- The number of microsatellite repeats (i.e. non-coding base pairs dispersed throughout the genome) may differ between two alleles in the same person making that person heterozygous for that particular allele. If tumour DNA from such a subject when amplified by PCR yields only one product, the tumour is said to show loss of heterozygosity implying allelic loss.
- Inactivation of genes can occur by:
  - allelic deletion
  - genetic mutation
  - transcriptional silencing
  - conversion of protooncogene to oncogene due to gene amplification.
- Some mutations do not result in change of function. They can be silent or nonsense mutations.
- Next generation sequencing has produced a mutational landscape of head and neck cancer.
- Proteomics refers to the profiling of proteins in cells and serum by 2D gel analysis, X-ray crystallography and by examination of protein interactions. This can be used to identify post-translational modifications, some of which are cancer cell specific and are not detected by genetic profiling.
- Metabolomics can be used to detect small molecules accumulating in the tissues and body fluids of oral cancer patients.
- Critical telomere shortening or inefficient telomere DNA repair can lead to both cell cycle arrest and cell immortalization due to expression of telomerase, which, for example, has been demonstrated in 80% of oral cancers.

## MAPPING AND IDENTIFICATION OF GENES ASSOCIATED WITH DISEASE

The traditional approach to understanding the molecular biology of disease was to look for and characterize the abnormal protein. Diabetes mellitus and sickle cell anaemia are two conditions that were elucidated at the molecular level in this way. Finding the protein when present in minute quantities, in an inaccessible site such as the inner ear, or when nothing is known of its mode of action, is, however, virtually impossible. This is the case with most of the genes involved in cellular growth control and also in inner ear function. Our approach to understanding such conditions relies on determining the chromosomal location, isolating and sequencing the gene so that the structure of the protein can be determined. This is known as the reverse genetic or positional cloning technique.

Strategies employed in determining the sites of these disease genes include cytogenetic analysis, molecular genetic analysis (typically LOH studies) and linkage analysis. Where a clearcut inheritance of a disease can be demonstrated, then linkage analysis can be useful in determining the location of the responsible gene. The key advance that has made this approach increasingly successful has been the construction of a high-resolution linkage map of the entire human genome. This map consists of genetic markers spaced less than 1 centimorgan apart and the process of linkage involves determining which of these the disease gene is close to. Analysis of the coinheritance of the genetic marker and the disease trait in families allows one to determine the probability of their physical proximity or linkage to each other. The statistical probability of linkage is expressed in terms of LOD scores. LOD is short for logarithm to the base 10 of the odds that the markers are linked at a recombination distance of  $N$  centimorgans. An LOD score of 3.0 favours linkage while a score of  $-2.0$  effectively rules out linkage.

Successful linkage analysis typically requires large pedigrees with many individuals known to have the disease trait and likewise many confirmed to be free of the disease. The process involves initial family analysis with clinical evaluation and accurate diagnosis. DNA samples are then obtained from each individual and genotyping and linkage analysis can then be carried out. Linkage and molecular deletion analysis generate genetic markers either side of the disease gene location. These genetic markers can be used to identify 'chunks of genomic DNA' cloned into yeast (yeast artificial chromosomes (YAC)) or bacteria (bacterial artificial chromosomes/P1 artificial chromosomes (BAC/PAC)) and organized into libraries. By a number of methods, these 'chunks' of genomic DNA can be directionally organized to produce a physical map of the region of interest from which candidate disease genes can be identified. The Human Genome Mapping Project has, to a great extent, physically mapped the human genome such that in some cases, when the region of interest is sufficiently small, all genes known to be between the markers can be considered positional candidates. The candidates are then analysed in disease

families until one is found that demonstrates mutations in affected individuals.

To facilitate the identification of the disease gene from a number of candidates within a particular region, several strategies are available. One such approach exploits our knowledge of animal models of the diseases that parallel

human inherited disorders. Another potential source of candidate genes is to identify and characterize genes that are expressed within a particular tissue. Genes expressed within the cochlea, for example, represent the genes critical for cochlear function and thus are excellent candidate genes for hereditary hearing loss.

## FUTURE RESEARCH

Although giant strides have already been made in the delineation of molecular events underlying disease processes, the knowledge of these events is far from comprehensive. A clearly identifiable adenoma–carcinoma sequence facilitated the correlation between genetic and histopathological progression models in colorectal cancer.<sup>19</sup> Several similar progression models have been proposed in head and neck squamous cell carcinoma evolution from epithelial dysplasia through to frank carcinoma, but the heterogeneity of carcinomas in different subsites precludes a molecular genetic progression model of equal rigour to that in colorectal cancer. Nevertheless, *p16* and *p53* inactivation have been identified as the two most frequent and critical events in head and neck carcinogenesis. An understanding of the sequence of genetic aberrations that occur in pathogenesis of tumours can assist in the prediction of progression from premalignant conditions to frank malignancy. An example of this is seen in the accumulation of genetic abnormalities in oral epithelium leading to the progression of oral dysplasia to oral squamous cell carcinoma. By analysing for loss of heterozygosity at 19 microsatellite loci in a series of oral dysplastic lesions and oral malignancies, Rosin et al.<sup>31</sup> showed that LOH at 3p and/or 9p occurred in virtually all cases of oral dysplasia which progressed to frank malignancy implying a ‘gatekeeper’ role of genes at those loci in the development of oral cancer, in themselves imparting a 3.8-fold increased relative risk of progression to squamous carcinoma. Furthermore, additional LOH at 4q, 8p, 11q or 17p imparted a 33-fold increased relative risk for developing cancer.<sup>31</sup> In certain cases, molecular genetic profiling has added a new dimension to age-old debates. For example, discordant genetic anomalies in inverted sinonasal papillomas and sinonasal carcinomas lend weight to the opinion that the former does not undergo malignant degeneration into the latter.<sup>32</sup>

Traditional methods of allelotyping and mutational analysis remain extremely time-consuming and labour-intensive. Recent years have seen the development of gene arrays (or ‘gene chips’) that allow the rapid assessment of the genetic profile of tumours or cell lines.<sup>33</sup> cDNA of up to 30 000 genes can be placed in arrays on glass slides on to which DNA and RNA samples can be allowed to hybridize using an automated two-colour fluorescence method for detection.<sup>34</sup> Hybridization with DNA can yield extensive genomic information, while hybridization with RNA takes genetic profiling a step further by semiquantitatively analysing the expression of genes rather than simply the presence or absence of the DNA. As the genetic aberrations and pathways associated with specific tumour development become better dissected, ‘designer’ chips can be produced that allow for screening for known mutations of those involved genes.<sup>35</sup> Array technology has also been married with molecular cytogenetics to produce array-based comparative genomic hybridization, which obviates the need for metaphase chromosome spreads for CGH.

The clinician might be forgiven for an expression of cynicism regarding the ‘molecular revolution’, feeling that little has changed despite ambitious predictions. The translation of molecular biology research into clinical practice – ‘from

laboratory to bedside’ – is without doubt the greatest challenge facing the clinician scientist. Progress so far has been largely at the level of disease classification, premorbid diagnosis and patient counselling. However, several discrete arms to translational research have hinted at the future role of genetic analysis determining the likely effectiveness of existing anti-cancer treatments. There are also isolated examples of effective novel therapeutic interventions.

Molecular analysis played a vital role in differentiating neurofibromatosis type I from neurofibromatosis type II, and it now seems difficult to understand how we ever confused the two in the first place. Such clarity of diagnosis is also anticipated for many syndromic and non-syndromic causes of sensorineural hearing loss as the genes are characterized and the clinical features analysed in more detail. This may enable clinicians to determine in which cases hearing loss will be progressive and target monitoring more appropriately. Diagnostic genetic testing will also become increasingly available to families with hereditary hearing loss and clinicians will need to be conversant with the ethical issues of testing and the implications of the results. By characterizing hearing loss at the molecular level we are also developing a greater understanding of inner ear physiology as the critical proteins involved in hearing become elucidated. This may lead to the development of agents able to repair and regenerate the diseased inner ear. The administration of these agents is already being evaluated using miniature infusion pumps.

It is hoped that in the future molecular characterization of tumour tissue will provide for a more accurate diagnosis and prognosis than conventional histopathological analysis. This may lead to a more rational treatment plan based on molecular features. Some genotype–phenotype correlations have already been made, particularly with respect to radio- and chemoresistance in tumours, but confirmation is needed with larger sample sizes as to their applicability into clinical practice. Refinement of the staging of tumours by probing of resection margins and the detection of histological subclinical metastatic disease in regional lymph nodes have been elegantly demonstrated by Brennan et al.<sup>36</sup> at Johns Hopkins. However, the technical complexity of the methodology has precluded its widespread application in the clinical arena. The development of technically simpler but equally sensitive techniques will inevitably render molecular staging routine in oncological practice. Although surveillance for tumour recurrence is less of a problem in the upper aerodigestive tract when compared with intra-abdominal malignancies, the emergence of molecular markers in saliva and serum will have a role in follow-up protocols for head and neck cancers.<sup>37</sup>

A comprehensive understanding of the molecular mechanisms underlying disease processes gives the potential of therapeutic restoration of DNA or protein function, as well as exploitation of the genetic abnormality for targeting of therapy. An example of the latter is the use of ONYX-015. ONYX-015 is an E1B 55-kDa gene-deleted adenovirus engineered to selectively replicate in and lyse *p53*-deficient cells while sparing normal cells. This tumour-targeting property has led

to a successful phase II trial of concomitant ONYX-015 and cisplatin/5-FU in the treatment of recurrent HNSCC, which indicated substantial objective responses as well as a high proportion of complete responses.<sup>38</sup> Post-treatment biopsies of tumour masses confirmed the replication selectivity of the virus *in vivo*.

A consideration of some of the landmarks in molecular biology (e.g. enzymatic DNA cleavage (1970) and PCR (1987)) readily indicates that molecular biology is a relatively young science. However, the rapid development of molecular techniques of such power and diversity as comparative genomic hybridization and gene array technology has permitted the dissection of molecular pathways and the identification of their disruption in disease processes. Some of these processes in specific diseases will be discussed in subsequent chapters. The future in the laboratory lies in the extension of our understanding of these molecular processes and the future in the clinic lies in the exploitation of

these molecular techniques in better informing disease classification, staging and treatment:

- ▶ Development of a molecular genetic progression model for head and neck squamous cell carcinoma of similar rigour to that for colorectal carcinoma.
- ▶ Development of gene arrays or gene chips that allow the rapid assessment of the genetic profile of tumours and can allow semiquantitative analysis of gene expression.
- ▶ Designer chips may be produced that allow for screening for unknown mutations of involved genes.
- ▶ Potential role for genetic analysis in determining the likely effectiveness of existing anti-cancer treatments.
- ▶ Providing more accurate diagnosis and prognosis than conventional histopathological techniques.
- ▶ Potential therapeutic restoration of DNA or protein function, as well as exploitation of a genetic abnormality for targeting therapy (e.g. use of ONYX-015).

## KEY POINTS

- Strategies employed in determining the sites of disease genes that code for the abnormal proteins include cytogenetic, molecular genetic and linkage analysis.
- Whilst genomics and other omics has generated a large body of data, cancers are complicated tissues. Future research may shift towards understanding the cancer environment and how perhaps to target this in future therapeutic strategies.

## REFERENCES

1. Hampson K, Bourn D. Principles of molecular genetics. *J Laryngol Otol* 1998; 112(2): 128–31.
2. Cruickshanks HA, Adams PD. Chromatin: a molecular interface between cancer and aging. *Curr Opin Genet Dev* 2011; 21(1): 100–06.
3. Schwartz YB, Pirrotta V. A new world of polycombs: unexpected partnerships and emerging functions. *Nat Rev Genet* 2013; 14(12): 853–64.
4. Sharp PA. Splicing of messenger RNA precursors. *Science* 1987; 235(4790): 766–71.
5. McKeown M. Alternative mRNA splicing. *Annu Rev Cell Biol* 1992; 8: 133–55.
6. Wu BH, Xiong XP, Jia J, Zhang WF. MicroRNAs: new actors in the oral cancer scene. *Oral Oncol* 2011; 47(5): 314–19.
7. Bentwich I, Avniel A, Karov Y, et al. Identification of hundreds of conserved and nonconserved human microRNAs. *Nat Genet* 2005; 37(7): 766–70.
8. Park NJ, Zhou H, Elashoff D, et al. Salivary microRNA: discovery, characterization, and clinical utility for oral cancer detection. *Clin Cancer Res* 2009; 15(17): 5473–77.
9. Perez P, Jang SI, Alevizos I. Emerging landscape of non-coding RNAs in oral health and disease. *Oral Dis* 2014; 20(3): 226–35.
10. Broomfield A, Bourn D. Basic techniques in molecular genetics. *J Laryngol Otol* 1998; 112(3): 230–34.
11. Speicher MR, Gwyn Ballard S, Ward DC. Karyotyping human chromosomes by combinatorial multi-fluor FISH. *Nat Genet* 1996; 12(4): 368–75.
12. Kallioniemi A, Kallioniemi OP, Sudar D, et al. Comparative genomic hybridization for molecular cytogenetic analysis of solid tumors. *Science* 1992; 258(5083): 818–21.
13. Mullis KB, Faloona FA. Specific synthesis of DNA *in vitro* via a polymerase-catalyzed chain reaction. *Methods Enzymol* 1987; 155: 335–50.
14. Lawyer FC, Stoffel S, Saiki RK, et al. Isolation, characterization, and expression in *Escherichia coli* of the DNA polymerase gene from *Thermus aquaticus*. *J Biol Chem* 1989; 264(11): 6427–37.
15. Emmert-Buck MR, Bonner RF, Smith PD, et al. Laser capture microdissection. *Science* 1996; 274(5289): 998–1001.
16. Knudson Jr AG. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci USA* 1971; 68(4): 820–23.
17. Cavenee WK, Dryja TP, Phillips RA, et al. Expression of recessive alleles by chromosomal mechanisms in retinoblastoma. *Nature* 1983; 305(5937): 779–84.
18. Merlo A, Herman JG, Mao L, et al. 5' CpG island methylation is associated with transcriptional silencing of the tumour suppressor p16/CDKN2/MTS1 in human cancers. *Nat Med* 1995; 1(7): 686–92.
19. Vogelstein B, Fearon ER, Kern SE, et al. Allelotype of colorectal carcinomas. *Science* 1989; 244(4901): 207–11.
20. Ah-See KW, Cooke TG, Pickford IR, et al. An allelotype of squamous carcinoma of the head and neck using microsatellite markers. *Cancer Res* 1994; 54(7): 1617–21.
21. Nawroz H, van der Riet P, Hruban RH, et al. Allelotype of head and neck squamous cell carcinoma. *Cancer Res* 1994; 54(5): 1152–55.
22. India Project Team of the International Cancer Genome Consortium. Mutational landscape of gingivo-buccal oral squamous cell carcinoma reveals new recurrently-mutated genes and molecular subgroups. *Nat Commun* 2013; 4: 2873.
23. Agrawal N, Frederick MJ, Pickering CR, et al. Exome sequencing of head and neck squamous cell carcinoma reveals inactivating mutations in NOTCH1. *Science* 2011; 333(6046): 1154–57.
24. Stransky N, Egloff AM, Tward AD, et al. The mutational landscape of head and neck squamous cell carcinoma. *Science* 2011; 333(6046): 1157–60.
25. Pandey A, Mann M. Proteomics to study genes and genomes. *Nature* 2000; 405(6788): 837–46.
26. Sugimoto M, Wong DT, Hirayama A, et al. Capillary electrophoresis mass spectrometry-based saliva metabolomics identified oral, breast and pancreatic cancer-specific profiles. *Metabolomics* 2010; 6(1): 78–95.
27. Tiziani S, Lopes V, Gunther UL. Early stage diagnosis of oral cancer using 1H NMR-based metabolomics. *Neoplasia* 2009; 11(3): 269–76, 4p following.
28. Greider CW. Telomeres, telomerase and senescence. *Bioessays* 1990; 12(8): 363–69.
29. Mutirangura A, Supiyaphun P, Trirekanan S, et al. Telomerase activity in oral leukoplakia and head and neck squamous cell carcinoma. *Cancer Res* 1996; 56(15): 3530–33.
30. Joseph I, Tressler R, Bassett E, et al. The telomerase inhibitor imetelstat depletes cancer stem cells in breast and pancreatic cancer cell lines. *Cancer Res* 2010; 70(22): 9494–504.
31. Rosin MP, Cheng X, Poh C, et al. Use of allelic loss to predict malignant risk for low-grade oral epithelial dysplasia. *Clin Cancer Res* 2000; 6(2): 357–62.
32. Califano J, Koch W, Sidransky D, Westra WH. Inverted sinonasal papilloma: a molecular genetic appraisal of its putative status as a precursor to squamous cell carcinoma. *Am J Pathol* 2000; 156(1): 333–37.

33. Lakhani SR, Ashworth A. Microarray and histopathological analysis of tumours: the future and the past? *Nat Rev Cancer* 2001; 1(2): 151–57.
34. Schena M, Shalon D, Davis RW, Brown PO. Quantitative monitoring of gene expression patterns with a complementary DNA microarray. *Science* 1995; 270(5235): 467–70.
35. Perou CM, Sorlie T, Eisen MB, et al. Molecular portraits of human breast tumours. *Nature* 2000; 406(6797): 747–52.
36. Brennan JA, Mao L, Hruban RH, et al. Molecular assessment of histopathological staging in squamous-cell carcinoma of the head and neck. *N Engl J Med* 1995; 332(7): 429–35.
37. Boyle JO, Mao L, Brennan JA, et al. Gene mutations in saliva as molecular markers for head and neck squamous cell carcinomas. *Am J Surg* 1994; 168(5): 429–32.
38. Khuri FR, Nemunaitis J, Ganly I, et al. A controlled trial of intratumoral ONYX-015, a selectively-replicating adenovirus, in combination with cisplatin and 5-fluorouracil in patients with recurrent head and neck cancer. *Nat Med* 2000; 6(8): 879–85.

## FURTHER READING

Alberts B, Johnson A, Lewis J, et al. *Molecular biology of the cell*. 4th ed. Oxford: Garland Science; 2002.

Latchman D (ed.). *Basic molecular and cell biology*. 3rd ed. London: BMJ Publishing Group; 1997.

Primrose SB, Twyman RM, Old RW. *Principles of gene manipulation*. 6th ed. Oxford: Blackwell Science; 2002.

Strachan T, Read AP. *Human molecular genetics*. 2nd ed. Oxford: BIOS Scientific Publishers; 2001.



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# GENETICS IN OTOLOGY AND NEUROTOLOGY

Mohammed-Iqbal Syed

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## SEARCH STRATEGY

Data in this chapter may be updated by a Medline search using a variety of relevant generic keywords, including: DNA, genetics, hereditary hearing loss, connexin 26, neurofibromatosis 2 and familial paragangliomas.

## INTRODUCTION

Advances in molecular genetics have helped to identify causative genes and proteins responsible for pathologies; this knowledge is pertinent to molecular target therapy and promises novel therapeutic interventions.

This chapter aims to review the mechanisms and principles of molecular genetics of hearing impairment, vestibular schwannoma, and glomus tumours, and will keep the reader abreast of new developments that may be relevant to identifying and treating these diseases in clinical practice.

## MOLECULAR GENETICS OF HEARING LOSS

Hearing loss is the most frequent sensory impairment in humans, with significant social and psychological implications. Permanent childhood hearing impairment of a moderate or greater degree (i.e. detection thresholds 40 decibels hearing level averaged across 0.5, 1, 2 and 4 kHz) is present at birth in about 1.6 per 1000 live births, of which approximately 1.0 in 1000 are bilateral impairments and 0.6 in 1000 are unilateral impairments.<sup>1,2</sup> However, studies have shown that the prevalence of permanent childhood hearing impairment continues to increase through infancy, and by the school entry hearing screen (4–5 years of age) possibly affects 3.5 per 1000 children.<sup>2</sup> It is estimated that at least

two thirds of cases of childhood-onset hearing loss have a genetic cause, with the remaining third caused by environmental factors (e.g. cytomegalovirus infection, meningitis, acquired conductive loss, and the impact of extracorporeal membrane oxygenation (ECMO)).<sup>3</sup> Improved clinical knowledge, awareness, and advances in antibiotics and vaccines has led to a decline in hearing loss resulting from environmental factors. Likewise, significant progress in the genetics of hereditary hearing impairment (HHI) has improved our understanding of the causes and early detection of HHI.

## Classification

The most useful distinction in hereditary hearing impairment (HHI) is syndromic versus non-syndromic. When SNHL occurs in isolation it is termed non-syndromic and when it is accompanied by other systemic disturbances it is termed syndromic. The majority of hearing impairments are non-syndromic (~70%), whereas a minority are syndromic (~30%).

Non-syndromic HHI is further classified by mode of inheritance:

- Autosomal recessive ~80%
- Autosomal dominant ~18%.<sup>4</sup>

Rare modes of transmission include X-linked and mitochondrial transmission, which account for the remaining 2% of hearing impairment.