

Clinical Ophthalmic Oncology

Eyelid and Conjunctival Tumors

Jacob Pe'er

Arun D. Singh

Bertil E. Damato

Editors

Third Edition

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Preface

Ophthalmic tumors are rare and diverse so that their diagnosis can be quite complex. Treatment usually requires special expertise and equipment and, in many instances, is controversial. The field is advancing rapidly, because of accelerating progress in tumor biology, pharmacology, and instrumentation. Increasingly, the care of patients with an ocular or adnexal tumor is provided by a multidisciplinary team, consisting of ocular oncologists, general oncologists, radiotherapists, pathologists, psychologists, and other specialists.

For all these reasons, we felt that there was a need for the new edition of the textbook providing a balanced view of current clinical practice. Although each section of *Clinical Ophthalmic Oncology, 3rd Edition* now represents a standalone volume, each chapter has a similar layout with boxes that highlight the key features, tables that provide comparison, and flow diagrams that outline therapeutic approaches.

The enormous task of editing a multi-author, multivolume textbook could not have been possible without the support and guidance by the staff at Springer: Caitlin Prim, Melanie Zerah, ArulRonika Pathinathan, and Karthik Rajasekar. Michael D. Sova kept the pressure to meet the production deadlines.

It is our sincere hope that our efforts will meet high expectation of the readers.

Jerusalem, Israel
Oxford, UK
Cleveland, OH, USA

Jacob Pe'er, MD
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To my wife, Edith, and my children, Liron, Neta, and Doron, for years of support and patience.

Jacob Pe'er, MD

To my family, Frankanne, Erika, Stephen, and Anna.

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To my parents who educated me beyond their means, my wife, Annapurna, and my children, Nakul and Rahul, who make all my efforts worthwhile.

Arun D. Singh, MD

Contents

1 Eyelid Tumors: Examination Techniques	1
Catherine J. Hwang and Julian D. Perry	
2 Eyelid Tumors: Classification and Differential Diagnosis	7
Jacob Pe'er and Shahar Frenkel	
3 Benign Eyelid Squamous and Melanocytic Tumors	15
Lynn Schoenfield and Arun D. Singh	
4 Basal Cell Carcinoma	33
Mordechai Rosner and Ido Didi Fabian	
5 Squamous Cell Carcinoma	45
Mordechai Rosner and Ido Didi Fabian	
6 Sebaceous Gland Carcinoma	53
Mordechai Rosner and Ido Didi Fabian	
7 Eyelid Tumors: Cutaneous Melanoma	63
Jacob Pe'er and Robert Folberg	
8 Adnexal Tumors	71
Martina C. Herwig-Carl and Karin U. Loeffler	
9 Stromal Tumors	83
Geeta K. Vemuganti and Santosh G. Honavar	
10 Surgical Techniques	97
Andrew J. Rong, Jennifer I. Hui, and David T. Tse	
11 Systemic Associations	113
Matteo Scaramuzzi, Lucy T. Xu, Arun D. Singh, and Elias I. Traboulsi	
12 Conjunctival and Corneal Tumors: Examination Techniques	131
Jacob Pe'er and Shahar Frenkel	
13 Conjunctival and Corneal Tumors: Classification and Differential Diagnosis	137
Jacob Pe'er and Shahar Frenkel	

14	Conjunctival and Corneal Tumors: Benign Epidermal and Melanocytic Tumors	143
	Jacob Pe'er and Shahar Frenkel	
15	Conjunctival and Corneal Tumors: Ocular Surface Squamous Neoplasia	159
	Jacob Pe'er, Shahar Frenkel, and Arun D. Singh	
16	Conjunctival and Corneal Tumors: Primary Acquired Melanosis	185
	Jacob Pe'er and Robert Folberg	
17	Conjunctival and Corneal Tumors: Melanoma	197
	Jacob Pe'er and Robert Folberg	
18	Conjunctival Stromal Tumors	209
	Jacob Pe'er and Shahar Frenkel	
19	Caruncle Tumors	235
	Hans E. Grossniklaus, Daniel R. Capiz-Correa, and Jill R. Wells	
20	Pharmacotherapy for Conjunctival Malignancies	245
	Ghada Al Bayyat, Dan Arreaza-Kaufman, Anat Galor, Jacob Pe'er, and Carol L. Karp	
21	Sentinel Lymph Node Biopsy for Eyelid and Conjunctival Malignancies	261
	Oded Sagiv and Bitá Esmaeli	
22	Surgical Techniques	279
	Anat Galor, Bennie H. Jeng, Arun D. Singh, and Carol L. Karp	
23	Radiation Therapy: Conjunctival and Eyelid Tumors	287
	Christopher Fleming, Shlomo Koyfman, and Arun D. Singh	
24	Conjunctival and Corneal Tumors: Systemic Associations	295
	Matteo Scaramuzzi, Lucy T. Xu, Arun D. Singh, and Elias I. Traboulsi	
	Index	307

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Eyelid Tumors: Examination Techniques

1

Catherine J. Hwang and Julian D. Perry

Introduction

Neoplasia may develop within any eyelid structure. Examination of the eyelid is thought to be relatively straightforward, given its anterior location and the ability to visualize its anterior and posterior surfaces. However, examination including structure and function is critical to determine the layers of the eyelid involved and if there might be extension posteriorly into the orbit or medially into the lacrimal system. The examination of an eyelid tumor determines the need for any ancillary tests and the surgical plan.

Presenting Symptoms

Eyelid neoplasia present with a limited spectrum of symptoms (Box 1.1). Most often, patients notice an abnormal eyelid appearance or asymmetry compared to the contralateral eyelid. The eyelid may harbor a distinct lesion, displaying elevation, ulceration, crusting, bleeding, altered pigmentation, telangiectasia, or other visible cutaneous or conjunctival changes. The patient may complain of loss of eyelashes or an irregularity along the eyelid margin.

History

The history begins with a description of the symptoms: severity, onset, and rate of progression. A targeted review of systems reveals additional clues to the etiology.

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Box 1.1 Symptoms of Eyelid Neoplasia

- Sensory: tenderness, itching, visual changes
- Motor: ptosis, lagophthalmos
- Structural: visible or palpable lesion, change in symmetry
- Functional: keratopathy or tearing
- Secondary: pigmentation, lymphadenopathy

Eyelid neoplasia may produce symptoms that occur with or without visible structural changes. Sensory symptoms such as pain, tenderness, itching, or vision symptoms due to keratopathy, induced astigmatism, or obstruction of vision may develop. Motor symptoms, such as blepharoptosis or lagophthalmos, may develop owing to involvement of the eyelid retractors and protractors or indirectly from a mass effect. Functional symptoms develop from mechanical keratoconjunctivitis, exposure keratopathy, or decreased lacrimal outflow.

Rate of Onset

Rapidity and progression help characterize the pathology. Most symptoms from eyelid tumors develop over weeks to months, but associated hemorrhage, infection, and inflammation may be acute. Both benign (e.g., angiomas, papillomas) and malignant (e.g., cutaneous malignancies, metastases) eyelid tumors can produce hemorrhage. Any eyelid tumor that blocks lacrimal outflow or causes diminished cutaneous integrity can result in infection. Eyelid tumors may also be associated with a significant inflammatory reactions.

Past Medical History

Because the majority of eyelid neoplasms are epidermal in origin, the past medical history should focus on risk factors for epidermal malignancy. Information should be obtained regarding family history of cutaneous malignancy, skin type, freckle density, eye color, hair color, and prior history of skin cancer. Patients

of Celtic or Scandinavian descent with blonde or red hair, blue eyes, and fair skin carry a greater risk for cutaneous malignancy [1, 2]. The history should also include immunosuppression, tobacco use, prior radiotherapy, sun exposure, and similar growths elsewhere on the skin.

Examination

The physical examination of an adult with suspected eyelid neoplasia does not end with direct visualization of the lesion. It should include a comprehensive inspection of the eyelid, ocular adnexa and orbit, eye, and other cutaneous lesions described in the history. Underlying conditions that may make reconstruction more challenging should be noted, including prominent globe, mid-face ptosis, hypoplastic orbital rim, lack of cutaneous or tissue redundancy, previous scarring from cutaneous malignancy repair or other surgery, asymmetry, lymph node enlargement, lagophthalmos, trichiasis, dry eye syndrome, and blepharitis.

Eyelid Examination

The patient should point out smaller lesions to the examiner using a hand mirror. The entire face should be evaluated to note Fitzpatrick skin type and any other cutaneous lesions. The eyelid examination should describe the appearance of the lesion, any associated anatomical deformities, and the results of palpation. The dimensions should be measured using a ruler or slit lamp beam. The eyelid examination should focus particularly on signs of malignancy, including telangiectasia, nodularity, pearly

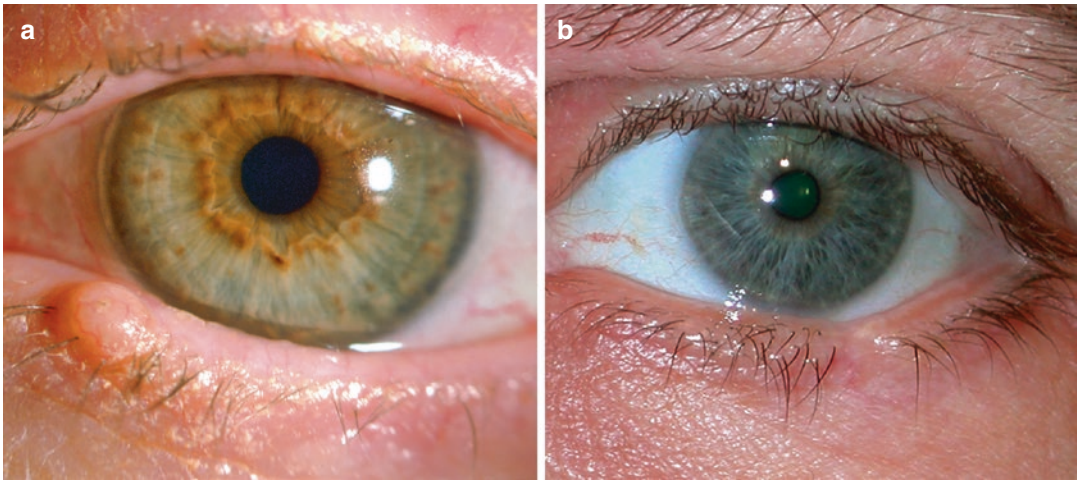


Fig. 1.1 Photograph of lower eyelid shows a benign eyelid nodule without loss of lashes (a) and loss of eyelid tissue with cilia loss secondary to a malignant tumor (b)

translucency, ulceration, bleeding, crusting, irregularity of the eyelid margin, meibomian gland effacement, misdirection of lashes or trichiasis, and loss of cilia (Fig.1.1; Box 1.2). Palpation results should describe the mobility of the lesion, as well as any fluctuance or associated tenderness. Color changes and irregularities should be noted.

Box 1.2 Signs of Malignant Eyelid Tumor

- Telangiectasia
- Nodularity, pearly translucency
- Ulceration, bleeding, crusting, margin notch
- Misdirection of lashes or trichiasis
- Loss of cilia
- Effacement of meibomian gland orifice

Function of the eyelid including levator excursion, orbicularis function, lagophthalmos, and lid lag in downgaze should be measured and noted. Horizontal eyelid laxity, blepharoptosis, cutaneous insufficiency, and other preexisting eyelid malpositions, scarring, and conditions should be noted, as they may challenge repair and will affect the reconstruction design. In addition, patients that relate these preoperative conditions to eyelid tumor surgery in the follow-up period can be reminded of the preoperative findings.

Ocular Adnexal Examination

Eyelid tumors may spread directly to the lacrimal gland, orbit, or lacrimal outflow apparatus. Conversely, primary tumors of these areas may occasionally present with only eyelid signs and symptoms. The structure and function of the orbit and ocular adnexal tissues in proximity to the

lesion should be evaluated. The examiner should palpate for preauricular, submandibular, cervical and supraclavicular adenopathy. Cranial nerves V and VII should be tested carefully to assess for involvement and possible perineural spread of an eyelid malignancy.

Eye Examination

The ocular examination should focus on detecting findings caused by, or associated with, the eyelid lesion. Slit lamp biomicroscopy may reveal signs of mechanical or exposure keratoconjunctivitis, or it may reveal signs of conjunctival spread of sebaceous cell carcinoma or cutaneous malignancy. During the evaluation of a pigmented eyelid lesion, the sclera and episclera should be observed for pigmentary changes as well. Direct intraocular extension of eyelid tumors is extremely rare, but funduscopy may reveal signs of ocular or orbital involvement (choroidal folds, venous congestion) in suspected cases.

Diagnostic Evaluation

Ancillary Laboratory and Imaging Studies

History and physical examination of a suspected eyelid tumor occasionally dictates ancillary testing. In cases of suspected eyelid granulomas, inflammatory labs may be indicated such as antinuclear cytoplasmic antibodies (p-ANCA/c-ANCA) and angiotensin-converting enzyme (ACE) to rule out more specific causes of inflammation. If the examination reveals associated orbital or lacrimal outflow signs, computed tomography (CT) or magnetic resonance imaging (MRI) may help to determine the extent of the lesion. Schirmer testing could be considered to document underlying dry eye disease. Lacrimal probing and irrigation should be performed for peri-punctal lesions, for lesions in proximity to the nasolacrimal drainage system, and for patients with preexisting epiphora. Photodocumentation of the lesion and periorbital should also be performed, especially prior to

biopsy. Marking the lesion and taking a photograph is helpful in localization if further procedures or monitoring are needed.

Dermatoscopy

Dermatoscopy is an in vivo noninvasive technique that may improve the clinical accuracy in diagnosing melanoma and other pigmented skin lesions [3]. Optical coherence tomography (OCT) may represent a new and promising technique for noninvasive investigation of skin tumors [4]. This modality may not only distinguish tumor tissue from normal tissue but may also visualize the epidermis, the dermoepidermal junction, and the dermis, as well as hair follicles, blood vessels, and sweat glands [5]. Although noninvasive techniques may improve diagnostic accuracy, the clinical diagnosis of eyelid tumors remains imperfect, and biopsy still represents the gold standard.

Biopsy

Based on clinical examination, the clinician is accurate in diagnoses in a high percent of patients anywhere from 83.7% to 96.9% [6, 7]. When the lesion is clinically misdiagnosed, it is often thought the lesion is benign but in fact histologically malignant. Malignant lesions can be clinically misdiagnosed as benign, especially when they are small and have nondescript surface features, thereby emphasizing the need for a confirmatory histology via incisional or excisional biopsy [6, 7].

The goal of biopsy is to determine the pathologic nature of the lesion, while minimizing adverse functional and cosmetic consequences. Tumor location and the presumptive clinical diagnosis largely dictate the approach and technique. Shave biopsy or excisional biopsy can be performed of lesions to determine pathology.

Biopsy-proven epidermal malignancies require margin-controlled excision and repair, with either frozen section control of Moh's micrographic surgery. Melanoma, sebaceous cell carcinoma, and Merkel cell carcinoma

require excision with wide margins. Some tumors, such as capillary hemangioma, may resolve spontaneously or require nonsurgical treatment (Chap. 10).

Treatment Planning

Information gathered from the history and eyelid examination determines the initial surgical plan for biopsy. This information also determines whether any special studies on the biopsy specimen are required. Any testing specific for the suspected diagnosis should be communicated to the pathologist in advance. For example, if sebaceous carcinoma is suspected, then the specimen usually is sent fresh for Oil Red O staining; however, it can be evaluated with immunohistochemistry on paraffin section with adipophilin and androgen receptor depending on the pathologist's preference [8]. If suspicion for lymphoproliferative disease exists, a fresh specimen for immunohistochemistry and cytology may be indicated. Such foresight may avoid inconclusive biopsy results, the need for an additional tissue biopsy, and lost time.

A detailed eyelid examination may also increase the efficiency of any anticipated surgery by determining the probable extent of tumor burden. For instance, if examination points to a larger, possibly infiltrating lesion rather than a smaller, localized process, the examination may dictate map biopsies to determine the extent of the lesion. Conversely, if examination shows a small, discreet lesion, then it may call for excisional biopsy to minimize the number of surgical interventions. Shave biopsy is also widely used and allows for examination of the tissue without significant disruption. The downside of shave biopsy is evaluation of the pathology at the depth of the lesion is sometimes inadequate. The examina-

tion results can direct the patient discussion to illuminate surgical risks and realities.

Conclusion

A systematic approach to the evaluation of suspected eyelid neoplasia allows the clinician to diagnose and treat these tumors efficiently and effectively. Current clinical diagnostic techniques remain inaccurate, and the threshold for biopsy should remain quite low. In the future, we hope less invasive diagnostic and therapeutic techniques will be available, as well as for improved preventive options and better early detection to limit the morbidity of these common tumors.

References

1. Gandini S, Sera F, Cattaruzza MS, et al. Meta-analysis of risk factors for cutaneous melanoma: III. Family history, actinic damage and phenotypic factors. *Eur J Cancer*. 2005;41:2040–59.
2. Cook BE, Bartley GB. Epidemiologic characteristics and clinical course of patients with malignant eyelid tumors in an incidence cohort in Olmsted County. *Minn Ophthalmol*. 1999;106:746–50.
3. Lallas A, Apalla Z, Chaidemenos G. New trends in dermatoscopy to minimize the risk of missing melanoma. *J Skin Cancer*. 2012;2012:820474. <https://doi.org/10.1155/2012/820474>. Epub 2012 Oct 8.
4. Khandwala M, Pennmetsa BR, Dey S, et al. Imaging of periocular basal cell carcinoma using en face optical coherence tomography: a pilot study. *Br J Ophthalmol*. 2010;94:1332–6.
5. Gambichler T, Jaedicke V, Terras S. Optical coherence tomography in dermatology: technical and clinical aspects. *Arch Dermatol Res*. 2011;303:457–73.
6. Kersten BC, Ewing-Chow D, Kulwin DR, et al. Accuracy of clinical diagnosis of cutaneous eyelid lesions. *Ophthalmology*. 1997;104:479–84.
7. Margo CE. Eyelid tumors: accuracy of clinical diagnosis. *Am J Ophthalmol*. 1999;128:635–6.
8. Schmitz EJ, Herwig-Carl MC, Holz RG, et al. Sebaceous gland carcinoma of the ocular adnex – variability in clinical and histological appearance with analysis of immunohistochemical staining patterns. *Graefes Arch Clin Exp Ophthalmol*. 2017;255(11):2277–85.



Eyelid Tumors: Classification and Differential Diagnosis

2

Jacob Pe'er and Shahar Frenkel

Introduction

In spite of being a small organ, the eyelids contain numerous histological elements that can be the origin of several types of benign or malignant tumors. In this chapter, we review the basic anatomy of the eyelid, outline a clinically relevant classification of eyelid tumors, and briefly discuss their differential diagnosis.

Anatomical Features

The eyelids are composed of four layers: skin and subcutaneous tissue, striated muscle (orbicularis oculi), tarsus, and conjunctiva [1]. The rest of the orbital entrance, which clinically may be considered as part of the eyelids, is covered, behind the skin and the orbicularis muscle, by the orbital septum that holds back the orbital fat.

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Eyelid Skin

The eyelid skin, especially the lower eyelid, is among the most sunlight-exposed anatomical structures. The eye and the eyelids are one of the most observed parts of the face, and therefore, eyelid tumors are usually diagnosed at an early stage. The eyelid skin is the thinnest in the body and lacks subcutaneous fat, but otherwise contains all other skin structures. In the pretarsal part, the skin and orbicularis oculi muscle are normally firmly attached to the tarsal plate, whereas in the preseptal part, they are more loosely attached. The skin epithelium is keratinized stratified squamous epithelium, the origin of all types of benign and malignant epidermal tumors. Melanocytes are spread in the basal layer of the epithelium and may give rise to melanocytic cutaneous lesions. The dermis contains also fibrous tissue, blood and lymphatic vessels, and nerves that can give rise to many types of fibrous tissue tumors, fibrohistiocytic tumors, vascular tumors, and neural tumors.

Adnexal Glands

The eyelids are rich in glandular tissue that may be the origin of various glandular tumors. Eccrine gland tumors may arise from the sweat glands of the eyelid skin as well as from the accessory lacrimal glands of Krause and

Wolfring. The glands of Moll can give rise to apocrine tumors. The sebaceous glands of Zeiss and the meibomian gland are the origin of sebaceous gland tumors.

Orbicularis Oculi

The entire orbital entrance is covered by the orbicularis oculi—a striated muscle that is divided into pretarsal and preseptal zones which are part of the eyelids and are involved in the eyelid movements and the orbital zone that is located over the external orbital bones.

Tarsus

The tarsi are firm plates composed of dense connective tissues that serve as the skeleton of the eyelids. The upper tarsal plates are much larger than the lower ones. The meibomian glands, large sebaceous glands, are embedded in the connective tissue of the tarsal plates. The superior tarsal muscle (Muller's muscle), a smooth muscle, is attached to the upper margin of the tarsus. A parallel muscle does not exist in the inferior tarsus, but the aponeurosis of the inferior rectus muscle attaches to the inferior edge of the inferior tarsus. The upper and lower orbital septum, a thin sheet of fibrous tissue, arises from the periosteum in the orbital rim and fuses with the levator aponeurosis superiorly and the lower margin of the lower tarsus inferiorly. All these histological structures can give rise to rare fibrous, striated, and smooth muscular and glandular tumors. The orbital fat behind the septum and the fat under the orbital part of the orbicularis oculi can be the origin of rare lipomatous tumors.

Palpebral Conjunctiva

The posterior eyelid surface is lined by the conjunctiva—a translucent mucous membrane that is composed of epithelium and subepithelial stroma—the substantia propria. The anatomical and histological features of the conjunctiva and

the possible tumors that can originate from this tissue are described elsewhere (Chap. 12).

Eyelid Margin

The eyelid margin is a flat area on the edge of each eyelid. The anatomical structures that are seen in the margin from the skin backwards are the eyelashes and their lash follicles, the gray line which consists of the tips of the pretarsal orbicularis muscle (the muscle of Riolan), the meibomian gland orifices, and the mucocutaneous junction just posterior to them.

Vascular System

The venous and lymphatic drainage is important in understanding the routes of possible eyelid tumor metastases. The eyelid has extensive vascularity that comes from two main sources—the internal carotid and external carotid arteries—with anastomoses between these two systems. The venous drainage is into the angular vein medially, superficial temporal vein laterally, and the orbital veins, anterior facial vein, and the pterygoid plexus posteriorly. The lymphatic drainage of the medial portions of the eyelids is into the submandibular lymph nodes and of the lateral portions into the superficial preauricular nodes and then into the deeper cervical nodes.

Nerve Supply

The sensory nerve supply to the eyelids is from the fifth cranial nerve, and the motor nerve supply to the striated muscles is from the third and seventh cranial nerves and to the smooth muscles from sympathetic nerves.

Classification of Eyelid Tumors

Tumors of the eyelid may be classified, like tumors in other organs, according to their tissue or cell of origin and as benign or malignant. In

most groups of tumors, unique histological subtypes behave differently in spite of being of the same cell of origin.

The classification of eyelid tumors that appears in this section is based primarily on the second edition of the World Health Organization (WHO) International Histological Classification of Tumors (Table 2.1) [2]. The epithelial tumor classification has been modified and divided into groups according to the tumor cell of origin. Some tumors that are missing from the WHO list have been added from other sources [3–5].

The vast majority of the eyelid tumors, benign and malignant, are of cutaneous origin, mostly epidermal. These tumors are divided into non-melanocytic and melanocytic tumors (Table 2.2). Benign epithelial proliferations, basal cell carcinoma, cystic structures, and melanocytic nevi represent about 85% of all eyelid tumors [6, 7]. The squamous cell carcinoma and the melanoma are relatively rare [7]. Tumors arising from adnexal structures (Table 2.3), fibrous tissue, fibrohistiocytic and muscular tumors (Table 2.4),

and other stromal tumors (Tables 2.5 and 2.6) are less frequent. Lymphoid tumors, hamartomas and choristomas, and inflammatory and infectious lesions that simulate neoplasms are listed in Table 2.7.

Differential Diagnosis

Various characteristics of the tumor and the patient's general health are important in making the correct diagnosis. The important features that should be noted in examining the eyelid tumor are the tumor location (upper or lower eyelid, inner or outer canthus); is it on the eyelid margin; the eyelid layer involved (skin, subcutaneous tissue, or palpebral conjunctiva); is the tumor solid or cystic; tumor size; the color of the lesion (pigmented or non-pigmented); skin color (red, pink, yellow, white, or blue); the tumor consistency (hard, soft, or rubbery); its surface (smooth, irregular, papillary, ulcerated, umbilicated, cratered, or keratinized); its shape (flat or raised, pedunculated, papillary); is the tumor thin or thick; is the tumor solitary or are there several or multiple tumors; is there loss of eyelashes; the patient's race, age, and gender; is the tumor movable with the skin or is it fixed to the subcutaneous layers; the existence of systemic diseases such as genetic diseases (e.g., neurofibromatosis) or systemic malignancies; and the existence of diseases or malignancies in the surrounding structures (the eyeball, conjunctiva, orbit, lacrimal drainage system, and neighboring skin).

Certain features of the tumor are suggestive of malignancy [5]. Development of a new lesion or changes in size, shape, color, or surface appearance of an existing lesion is suspicious for malignant conversion. Poorly defined borders, palpable induration beyond visible boundaries, loss of fine cutaneous rhytids, hypervascularity, ulceration, and destruction of the normal eyelid architecture are all worrisome. Lesions that are not freely mobile due to invasion of underlying structures and those associated with regional lymphadenopathy, hypesthesia, paresthesia or pain, indicating

Table 2.1 Major types of eyelid tumors

Category	Subtypes
Epidermal tumors	Non-melanocytic tumors
	Melanocytic tumors
Adnexal tumors	Sebaceous gland tumors
	Sweat gland tumors
	Lacrimal gland tumors
	Hair follicle tumors
	Cystic lesions
Stromal tumors	Fibrous tissue tumors
	Fibrohistiocytic tumors
	Lipomatous tumors
	Smooth muscle tumors
	Skeletal muscle tumors
	Vascular tumors
	Perivascular tumors
	Neural tumors
	Lymphoid, plasmacytic, and leukemic tumors
	Cartilage and bone tumors
	Hamartoma and choristoma
	Palpebral conjunctival tumors
	Secondary tumors
Metastatic tumors	
Inflammatory and infectious lesions that simulate neoplasms	

Table 2.2 Classification of epidermal tumors of the eyelid

Category	Subtypes	
Non-melanocytic	Benign	Squamous cell papilloma
		Seborrheic keratosis
		Inverted follicular keratosis
		Reactive hyperplasia (pseudoeplithiomatous hyperplasia)
	Premalignant	Actinic (solar) keratosis
		Intraepithelial neoplasia
		Sebaceous nevus (of Jadassohn)
		Xeroderma pigmentosum
	Malignant	Basal cell carcinoma
		Squamous cell carcinoma
		Mucoepidermoid carcinoma
		Keratoacanthoma
Melanocytic	Epithelial pigmentation	Ephelis or freckles
		Lentigo simplex
		Solar lentigo
	Benign	Junctional nevus
		Intradermal nevus
		Compound nevus
		Spitz nevus
		Balloon cell nevus
		Blue nevus
		Cellular blue nevus
		Oculodermal nevus of Ota
	Premalignant	Congenital dysplastic nevus
		Lentigo maligna (melanotic freckle of Hutchinson)
	Malignant	Melanoma arising from nevi
		Melanoma arising in lentigo maligna
		Melanoma arising de novo

Table 2.3 Classification of adnexal and cystic tumors of the eyelid

Category	Subtypes	
Sebaceous gland tumors	Benign	Sebaceous gland hyperplasia Sebaceous gland adenoma
	Malignant	Sebaceous gland carcinoma
Sweat gland and lacrimal gland tumors	Benign	Syringoma
		Papillary syringadenoma
		Eccrine spiradenoma
		Eccrine acrospiroma
		Pleomorphic adenoma (benign mixed tumor)
		Eccrine cylindroma
		Apocrine adenoma
		Other benign tumors
	Malignant	Sweat gland (eccrine) adenocarcinoma
		Mucinous sweat gland adenocarcinoma
		Apocrine gland adenocarcinoma
		Adenoid cystic carcinoma
		Porocarcinoma
Hair follicle tumors	Benign	Trichoepithelioma
		Trichofolliculoma/trichoadenoma
		Trichilemmoma
		Pilomatrixoma (calcifying epithelioma of Malherbe)
	Malignant	Carcinoma of hair follicles

Table 2.3 (continued)

Category	Subtypes	
Other cystic lesions	Benign	Epidermal inclusion cyst
		Sebaceous cyst
		Retention cyst
		Eccrine hidrocystoma
		Apocrine hidrocystoma
		Trichilemmal cyst
		Other benign cystic lesion

Table 2.4 Classification of fibrous, fibrous histiocytic, and muscular tumors of the eyelid

Origin	Type	Tumor	
Fibrous	Benign	Fibroma	
		Keloid	
		Nodular fasciitis	
		Proliferative fasciitis	
		Fibromatosis	
	Malignant	Fibrosarcoma	
		Congenital fibrosarcoma	
Fibrous histiocytic	Benign	Xanthelasma	
		Xanthoma	
		Dermatofibroma	
		Xanthogranuloma	
		Fibrous histiocytoma	
		Juvenile xanthogranuloma	
		Necrotic xanthogranuloma	
		Reticulohistiocytoma	
	Intermediate	Atypical fibroxanthoma	
		Dermatofibrosarcoma protuberans	
		Angiomatoid fibrous histiocytoma	
		Malignant	Malignant fibrous histiocytoma
			Malignant giant cell fibrous histiocytoma
Malignant fibroxanthoma			
Smooth muscle	Benign	Leiomyoma	
		Angiomyoma	
	Malignant	Leiomyosarcoma	
Skeletal muscle	Benign	Rhabdomyoma	
	Malignant	Rhabdomyosarcoma	

lymphatic or perineural spread are also suspicious for malignancy. Lesions associated with chronic inflammation that respond partially or temporarily to topical corticosteroids or antibiotics also may harbor malignancies. However, one should keep in mind that on the one hand malignant tumors can appear without any worrisome signs, while totally benign tumors can express some of the abovementioned features.

Table 2.5 Classification of vascular and perivascular tumors of the eyelid

Category	Subtypes		
Vascular	Benign	Nevus flammeus (port wine stain)	
		Papillary endothelial hyperplasia	
		Capillary hemangioma	
		Cavernous hemangioma	
		Venous hemangioma	
		Epithelioid hemangioma (angiolymphoid hyperplasia)	
		Arteriovenous malformation	
		Lymphangioma	
		Malignant	Angiosarcoma
			Lymphangiosarcoma
Kaposi's sarcoma			
Perivascular	Benign	Hemangiopericytoma	
		Glomus tumor	
	Malignant	Malignant hemangiopericytoma	
		Malignant glomus tumor	

Table 2.6 Classification of neural, lipomatous, cartilage, and bone tumors of the eyelid

Category	Subtypes	
Neural	Benign	Traumatic neuroma
		Neurofibroma
		Plexiform neurofibroma
		Schwannoma (neurilemoma)
		Others, e.g., neuroglial choristoma
		Malignant
		Merkel cell tumor
Lipomatous	Benign	Lipoma
		Others, e.g., hibernoma
	Malignant	Liposarcoma
Cartilage and bone	Benign	Chondroma
		Osteoma
	Malignant	Chondrosarcoma
		Mesenchymal chondrosarcoma
	Osteosarcoma	

Table 2.7 Classification of lymphoid tumors, hamartomas, choristomas, and inflammatory and infectious lesions that simulate neoplasms

Category	Subtypes
Lymphoid	Benign lymphoid hyperplasia
	Lymphoma
	Plasmacytoma
	Leukemic infiltration
Hamartomas and choristomas	Dermoid cyst
	Phakomatous choristoma
	Ectopic lacrimal gland
Inflammatory and infectious lesions	Chalazion
	Pyogenic granuloma
	Verruca vulgaris
	Molluscum contagiosum
	Others
Others	e.g., myxoma

Epidermal Non-melanocytic Tumors

The most common benign epithelial tumor is the squamous papilloma that is often sessile or pedunculated with papillary shape and keratinized surface (Table 2.2). Squamous papillomata may be multiple. Other epithelial tumors, including the premalignant actinic keratosis or small squamous cell carcinoma may look similar. Basal cell carcinoma comprises over 90% of all malignant eyelid tumors [7]. Its common location is the lower eyelid and medial canthus; it is usually firm and often has an ulcerated center. Other ulcerated eyelid tumors, such as keratoacanthoma or the more rare papillary syringadenoma, should be differentiated from BCC. Features of keratoacanthoma, such as rapid growth and possible spontaneous regression, can help in its diagnosis. Staging of carcinomas of the eyelid skin and adnexa can be found in the AJCC Cancer Staging Manual [8].

Epidermal Melanocytic Tumors

The most common pigmented eyelid lesions are the nevi, which are usually flat or mildly elevated and can appear anywhere in the eyelid in any size, and when appearing on the eyelid margin

can be sessile (Table 2.2). Congenital nevi usually appear at birth and acquired nevi between the ages of 5 and 10 years. Nevi should be differentiated on the one hand from flat epithelial pigmentation such as ephelis or freckles and, on the other hand, from the flat premalignant lentigo maligna or from malignant melanoma that is relatively rare in the eyelids.

Adnexal and Cystic Tumors

The eyelid adnexa include many different glands that are the origin of various benign and malignant tumors (Table 2.3). These include cystic lesions such as eccrine and apocrine hidrocystoma that are totally benign and may be transparent or have a distinct color like the blue apocrine hidrocystoma. On the other hand, there are very malignant solid sebaceous gland carcinomas that may resemble chalazion but unlike chalazion cause loss of eyelashes.

Stromal Tumors

The stromal eyelid tumors usually have a smooth surface, being under the skin (Tables 2.4, 2.5, and 2.6). The tumor elevation may have normal skin color, but many of the tumors will have a distinct color. Xanthomatous lesions are usually yellow. Most hemangiomas, diffuse or localized, are red. Subcutaneous varix is soft and blue, and Kaposi's sarcoma is blue or red. Merkel cell tumor is red or violaceous. Eyelid lymphoma can be manifested as a smooth, firm subcutaneous nodule. Sometimes also subcutaneous tumors can be sessile or even ulcerated, so such phenomena, which are usually seen in epidermal tumors, should not exclude them.

Inflammatory and Infective Simulating Conditions

In the differential diagnosis of eyelid tumors, we should include lesions that simulate tumors (Table 2.7). The most common simulating lesions

are inflammatory lesions such as chalazion or pyogenic granuloma (a misnomer for granulation tissue) or infectious viral lesions such as molluscum contagiosum or verruca vulgaris that is clinically and histologically similar to squamous papilloma. Many dermatological diseases such as amyloidosis and malakoplakia or connective tissue disease and systemic metabolic diseases such as hemachromatosis may, sometimes, simulate eyelid tumors and should be differentiated from them.

References

1. Bedrossian EH. Chapter 5: Embryology and anatomy of the eyelid. In: Tasman W, Jaeger EA, editors. *Duane's foundation of clinical ophthalmology, ocular anatomy, embryology and teratology*, vol. 1. Philadelphia: Lippincott Williams & Wilkins; 2004. p. 1–24.
2. Campbell RJ, Sobin LH. Tumours of the eyelid. In: *Histological typing of tumours of the eye and its adnexa*, World Health Organization international histological classification of tumors. 2nd ed. Berlin: Springer; 1998. p. 3–9.
3. Shields JA, Shields CL. *Atlas of eyelid and conjunctival tumors*. Philadelphia: Lippincott Williams & Wilkins; 1999. p. 3–189.
4. Hassan AS, Nelson CC. Benign eyelid tumors and skin diseases. *Int Ophthalmol Clin*. 2002;42:135–49.
5. Soparkar CN, Patrinely JR. Eyelid cancers. *Curr Opin Ophthalmol*. 1998;9:49–53.
6. Kersten RC, Ewing-Chow D, Kulwin DR, et al. Accuracy of clinical diagnosis of cutaneous eyelid lesions. *Ophthalmology*. 1997;104:479–84.
7. Cook BE, Bartley GB. Epidemiologic characteristics and clinical course of patients with malignant eyelid tumors in an incidence cohort in Olmsted County. *Minn Ophthalmol*. 1999;106:746–50.
8. Esmaeli B, Dutton JJ, Graue GF, et al. Chapter 64: Eyelid carcinoma. In: Amin MB, et al., editors. *AJCC Cancer staging manual*. 8th ed. New York: Springer; 2017. p. 779–85.



Benign Eyelid Squamous and Melanocytic Tumors

3

Lynn Schoenfield and Arun D. Singh

Introduction

The eyelid consists of six layers with epidermis externally and palpebral conjunctiva internally. Between these two (from outer to inner) are dermis, loose subcutaneous layer, orbicularis muscle, and tarsal plate. The epithelium consists of squamous cells and melanocytes primarily with smaller numbers of Langerhans cells and Merkel cells. The presence of Langerhans cells is important to recognize, as they, like melanocytes, are positive for the immunohistochemical stain S100. Benign tumors of the eyelid include a variety of nonpigmented and pigmented epidermal tumors, which arise from squamous and melanocytic cells, respectively, adnexal tumors (Chap. 4), stromal tumors (Chap. 5), and benign lymphoid proliferations. Important to note is that not all clinically pigmented lesions are melanocytic, since squamous cell proliferations can include scattered melanocytes or melanin pigment, thus giving a pigmented appearance to a lesion. The benign epidermal tumors of the eyelid are similar to those observed in the other sun-exposed areas

of the skin, but they may also include conjunctival tumors as well. Some of these tumors represent manifestations of systemic disease (Chap. 11). A classification of the epidermal eyelid tumors is presented in Table 3.1. Only the description of the most common and frequently observed benign tumors, along with their corresponding premalignant lesions and tumor-like nonneoplastic lesions, is included in this chapter.

Squamous (Non-melanocytic) Tumors

Squamous Cell Papilloma

Squamous papillomas are the most common benign tumors typically occurring in middle-aged or older adults. The clinical appearance is that of a pedunculated or sessile nodular growth with a variably convoluted surface with or without hyperkeratosis. They are often multiple, present at the lid margin, and are skin-colored (Fig. 3.1a) [1].

Microscopically a papilloma consists of benign squamous hyperplastic (acanthotic) epithelium with variable hyperkeratosis or parakeratosis overlying an expanded fingerlike fibrovascular core, which creates the exophytic nodule. They sometimes have overlapping features with seborrheic keratosis (Fig. 3.1b). If symptomatic, surgical excision may be performed.

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Table 3.1 Classification of epidermal tumors of the eyelid, excluding adnexal tumors

Types	Subtypes	
Non-melanocytic	Benign	Squamous cell papilloma
		Seborrheic keratosis
		Inverted follicular keratosis
		Molluscum contagiosum
		Reactive hyperplasia (pseudoeplitheliomatous hyperplasia)
	Potentially premalignant	Actinic (solar) keratosis
		Intraepithelial neoplasia
		Sebacous nevus (of Jadassohn)
	Malignant	Basal cell carcinoma
Squamous cell carcinoma		
Melanocytic	Benign epithelial pigmentation or hypermelanosis	Ephelis or freckles
		Lentigo simplex
		Solar lentigo
	Benign	Junctional nevus
		Intradermal nevus
		Compound nevus
		Spitz nevus
		Balloon cell nevus
		Blue nevus and cellular blue nevus
		Oculodermal nevus of Ota
		Seborrheic keratosis
	Potentially premalignant	Congenital dysplastic nevus
		Lentigo maligna (melanotic freckle of Hutchinson)
	Malignant	Melanoma arising from nevi
		Melanoma arising in lentigo maligna
Melanoma arising de novo		

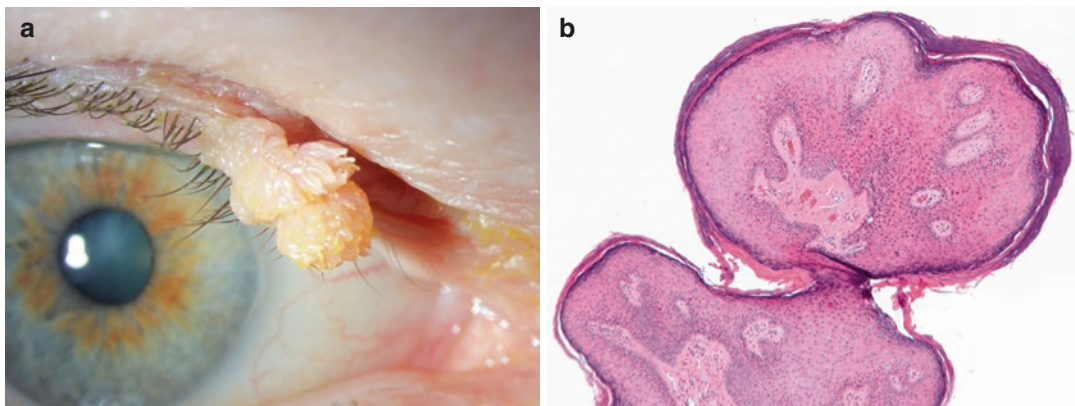


Fig. 3.1 Squamous papilloma. Clinical appearance. (a) Polypoid lesion consisting of benign squamous epithelium with variable acanthosis and hyperkeratosis overlying

a fibrovascular core ((b) hematoxylin and eosin; original magnification 4×)

Seborrheic Keratosis

Seborrheic keratoses are commonly acquired skin lesions which can occur on the eyelid affecting middle-aged and elderly patients. They have

also been referred to as basal cell papilloma, seborrheic wart, and senile verruca. The clinical appearance varies considerably in terms of size (few millimeters to several centimeters) and degree of pigmentation making it sometimes